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(54) Title: DECHI ATION OF LUNG TISSUE RV H	EDGE	HOG-LIKE POLYPEPTIDES AND FORMULATIONS AND USE

(54) Title: REGULATION OF LUNG TISSUE BY HEDGEHOG-LIKE POLYPEPTIDES, AND FORMULATIONS AND USES RELATED THERETO

#### (57) Abstract

The present application relates to a method for modulating the growth state of a lung tissue, or a cell thereof, e.g., by ectopically contacting the tissue, in vitro or in vivo, with a hedgehog therapeutic, a ptc therapeutic, or an FGF-10 therapeutic in an amount effective to alter the rate (promote or inhibit) of proliferation of cells in the lung tissue, e.g., relative to the absence of administeration of the hedgehog therapeutic or ptc therapeutic. The subject method can be used, for example, to modulate the growth state of epithelial and/or mesenchymal cells of a lung tissue, such as may be useful as part of a regimen for prevention of a disease state, or in the treatment of an existing disease state or other damage to the lung tissue.

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# Regulation of Lung Tissue by Hedgehog-like Polypeptides, and Formulations and Uses Related Thereto

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#### Background of the Invention

Pattern formation is the activity by which embryonic cells form ordered spatial

arrangements of differentiated tissues. The physical complexity of higher organisms arises during embryogenesis through the interplay of cell-intrinsic lineage and cell-extrinsic signaling. Inductive interactions are essential to embryonic patterning in vertebrate development from the earliest establishment of the body plan, to the patterning of the organ systems, to the generation of diverse cell types during tissue differentiation (Davidson, E., (1990) Development 108: 365-389; Gurdon, J. B., (1992) Cell 68: 185-199; Jessell, T. M. et al., (1992) Cell 68: 257-270). The effects of developmental cell interactions are varied. Typically, responding cells are diverted from one route of cell differentiation to another by inducing cells that differ from both the uninduced and induced states of the responding cells (inductions). Sometimes cells induce their neighbors to differentiate like themselves

(homoiogenetic induction); in other cases a cell inhibits its neighbors from differentiating like itself. Cell interactions in early development may be sequential, such that an initial

inductive interactions occur not only in embryos, but in adult cells as well, and can act to establish and maintain morphogenetic patterns as well as induce differentiation (J.B. Gurdon (1992) *Cell* 68:185-199).

Members of the *Hedgehog* family of signaling molecules mediate many important short- and long-range patterning processes during invertebrate and vertebrate development- In the fly a single *hedgehog* gene regulates segmental and imaginal disc patterning. In contrast, in vertebrates a *hedgehog* gene family is involved in the control of left-right asymmetry, polarity in the CNS, somites and limb, organogenesis, chondrogenesis and spermatogenesis.

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The first hedgehog gene was identified by a genetic screen in the fruitfly Drosophila melanogaster (Nüsslein-Volhard, C. and Wieschaus, E. (1980) Nature 287, 795-801). This screen identified a number of mutations affecting embryonic and larval development. In 1992 and 1993, the molecular nature of the Drosophila hedgehog (hh) gene was reported (C.F., Lee et al. (1992) Cell 71, 33-50), and since then, several hedgehog homologues have been isolated from various vertebrate species. While only one hedgehog gene has been found in Drosophila and other invertebrates, multiple Hedgehog genes are present in vertebrates.

The various *Hedgehog* proteins consist of a signal peptide, a highly conserved N-terminal region, and a more divergent C-terminal domain. In addition to signal sequence cleavage in the secretory pathway (Lee, J.J. et al. (1992) Cell 71:33-50; Tabata, T. et al. (1992) Genes Dev. 2635-2645; Chang, D.E. et al. (1994) Development 120:3339-3353), Hedgehog precursor proteins undergo an internal autoproteolytic cleavage which depends on conserved sequences in the C-terminal portion (Lee et al. (1994) Science 266:1528-1537; Porter et al. (1995) Nature 374:363-366). This autocleavage leads to a 19 kD N-terminal peptide and a C-terminal peptide of 26-28 kD (Lee et al. (1992) supra; Tabata et al. (1992) supra; Chang et al. (1994) supra; Lee et al. (1994) supra; Bumcrot, D.A., et al. (1995) Mol. Cell. Biol. 15:2294-2303; Porter et al. (1995) supra; Ekker, S.C. et al. (1995) Curr. Biol. 5:944-955; Lai, C.J. et al. (1995) Development 121:2349-2360). The N-terminal peptide stays tightly associated with the surface of cells in which it was synthesized, while the C-terminal peptide is freely diffusible both in vitro and in vivo (Lee et al. (1994) supra; Bumcrot et al. (1995) supra; Mart', E. et al. (1995) Development 121:2537-2547; Roelink, H. et al. (1995) Cell 81:445-455). Interestingly, cell surface retention of the N-terminal

peptide is dependent on autocleavage, as a truncated form of HH encoded by an RNA which terminates precisely at the normal position of internal cleavage is diffusible in vitro (Porter et al. (1995) supra) and in vivo (Porter, J.A. et al. (1996) Cell 86, 21-34). Biochemical studies have shown that the autoproteolytic cleavage of the HH precursor protein proceeds through an internal thioester intermediate which subsequently is cleaved in a nucleophilic substitution. It is likely that the nucleophile is a small lipophilic molecule which becomes covalently bound to the C-terminal end of the N-peptide (Porter et al. (1996) supra), tethering it to the cell surface. The biological implications are profound. As a result of the tethering, a high local concentration of N-terminal Hedgehog peptide is generated on the surface of the Hedgehog producing cells. It is this N-terminal peptide which is both necessary and sufficient for short and long range Hedgehog signaling activities in Drosophila and vertebrates (Porter et al. (1995) supra; Ekker et al. (1995) supra; Lai et al. (1995) supra; Roelink, H. et al. (1995) Cell 81:445-455; Porter et al. (1996) supra; Fietz, M.J. et al. (1995) Curr. Biol. 5:643-651; Fan, C.-M. et al. (1995) Cell 81:457-465; Mart', E., et al. (1995) Nature 375:322-325; Lopez-Martinez et al. (1995) Curr. Biol 5:791-795; Ekker, S.C. et al. (1995) Development 121:2337-2347; Forbes, A.J. et al. (1996) Development 122:1125-1135).

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HH has been implicated in short- and longe range patterning processes at various sites during *Drosophila* development. In the establishment of segment polarity in early embryos, it has short range effects which appear to be directly mediated, while in the patterning of the imaginal discs, it induces long range effects via the induction of secondary signals.

In vertebrates, several *hedgehog* genes have been cloned in the past few years (see Table 1). Of these genes, *Shh* has received most of the experimental attention, as it is expressed in different organizing centers which are the sources of signals that pattern neighbouring tissues. Recent evidence indicates that *Shh* is involved in these interactions.

The interaction of a *hedgehog* protein with one of its cognate receptor, *patched*, sets in motion a cascade involving the activation and inhibition of downstream effectors, the ultimate consequence of which is, in some instances, a detectable change in the transcription or translation of a gene. Transcriptional targets of *hedgehog* signaling are the *patched* gene itself (Hidalgo and Ingham, 1990 *Development* 110, 291-301; Marigo et al., 1996) and the vertebrate homologs of the drosophila cubitus interruptus (Ci) gene, the *GLI* genes (Hui et al.

(1994) Dev Biol 162:402-413). Patched gene expression has been shown to be induced in cells of the limb bud and the neural plate that are responsive to Shh. (Marigo et al. (1996) Development 122:1225-1233). The GLI genes encode putative transcription factors having zinc finger DNA binding domains (Orenic et al. (1990) Genes & Dev 4:1053-1067; Kinzler et al. (1990) Mol Cell Biol 10:634-642). Transcription of the GLI gene has been reported to be upregulated in response to hedgehog in limb buds, while transcription of the GLI3 gene is downregulated in response to hedgehog induction (Marigo et al. (1996) Development 122:1225-1233). Moreover, it has been demonstrated that elevated levels of Ci are sufficient to activate patched (ptc) and other hedgehog target genes, even in the absence of hedgehog activity.

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#### Summary of the Invention

One aspect of the present application relates to a method for modulating the growth state of an lung tissue, or a cell thereof, e.g., by ectopically contacting the tissue, in vitro or in vivo, with a hedgehog therapeutic, a ptc therapeutic, or an FGF-10 therapeutic (described infra) in an amount effective to alter the rate (promote or inhibit) of proliferation of cells in the lung tissue, e.g., relative to the absence of administeration of the hedgehog therapeutic or ptc therapeutic. The subject method can be used, for example, to modulate the growth state of epithelial and/or mesenchymal cells of a lung tissue, such as may be useful as part of a regimen for prevention of a disease state, or in the treatment of an existing disease state or other damage to the lung tissue.

Wherein the subject method is carried out using a hedgehog therapeutic, the hedgehog therapeutic preferably a polypeptide including a hedgehog portion comprising at least a bioactive extracellular portion of a hedgehog protein, e.g., the hedgehog portion includes at least 50, 100 or 150 (contiguous) amino acid residues of an N-terminal half of a hedgehog protein. In preferred embodiments, the hedgehog portion includes at least a portion of the hedgehog protein corresponding to a 19kd fragment of the extracellular domain of a hedgehog protein.

In certain preferred embodiments, the *hedgehog* portion has an amino acid sequence at least 60, 75, 85, or 95 percent identical with a *hedgehog* protein of any of SEQ ID Nos. 10-18 or 20, though sequences identical to those sequence listing entries are also contemplated as useful in the present method. The *hedgehog* portion can be encoded by a nucleic acid which hybridizes under stringent conditions to a nucleic acid sequence of any of SEQ ID Nos. 1-9 or 19, e.g., the *hedgehog* portion can be encoded by a vertebrate *hedgehog* gene, especially a human *hedgehog* gene.

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In certain embodiments, the *hedgehog* polypeptide is modified with one or more sterol moieties, e.g., cholesterol or a derivative thereof.

In certain embodiments, the *hedgehog* polypeptide is modified with one or more fatty acid moieties, such as a fatty acid moiety selected from the group consisting of myristoyl, palmitoyl, stearoyl, and arachidoyl.

In other embodiments, the subject method can be carried out by administering a gene activation construct, wherein the gene activation construct is deigned to recombine with a genomic *hedgehog* gene of the patient to provide a heterologous transcriptional regulatory sequence operatively linked to a coding sequence of the *hedgehog* gene.

In still other embodiments, the subject method can be practiced with the administration of a gene therapy construct encoding a *hedgehog* polypeptide. For instance, the gene therapy construct can be provided in a composition selected from a group consisting of a recombinant viral particle, a liposome, and a poly-cationic nucleic acid binding agent,

In yet other embodiments, the subject method can be carried out using a ptc therapeutic. An exemplary ptc therapeutic is a small organic molecule which binds to a patched protein and derepresses patched-mediated inhibition of mitosis, e.g., a molecule which binds to patched and mimics hedgehog-mediated patched signal transduction, which binds to patched and regulates patched-dependent gene expression. For instance, the binding of the ptc therapeutic to patched may result in upregulation of patched and/or gli expression.

In a more generic sense, the ptc therapeutic can be a small organic molecule which induces hedgehog-mediated patched signal transduction, such as by altering the localization, protein-protein binding and/or enzymatic activity of an intracellular protein involved in a

patched signal pathway. For instance, the ptc therapeutic may alter the level of expression of a hedgehog protein, a patched protein or a protein involved in the intracellular signal transduction pathway of patched.

In certain embodiments, the *ptc* therapeutic is an antisense construct which inhibits the expression of a protein which is involved in the signal transduction pathway of *patched* and the expression of which antagonizes *hedgehog*-mediated signals. The antisense construct is perferably an oligonucleotide of about 20-30 nucleotides in length and having a GC content of at least 50 percent.

In other embodiments, the *ptc* therapeutic is an inhibitor of protein kinase A (PKA), such as a 5-isoquinolinesulfonamide. The PKA inhibitor can be a cyclic AMP analog. Exemplary PKA inhibitors include N-[2-((p-bromocinnamyl)amino)ethyl]-5-isoquinolinesulfonamide, 1-(5-isoquinoline-sulfonyl)-2-methylpiperazine, KT5720, 8-bromocAMP, dibutyryl-cAMP and PKA Heat Stable Inhibitor isoform α. Another exemplary PKA inhibitor is represented in the general formula:

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wherein,

 $R_1$  and  $R_2$  each can independently represent hydrogen, and as valence and stability permit a lower alkyl, a lower alkenyl, a lower alkynyl, a carbonyl (such as a carboxyl, an ester, a formate, or a ketone), a thiocarbonyl (such as a thioester, a thioacetate, or a thioformate), an amino, an acylamino, an amido, a cyano, a nitro, an azido, a sulfate, a sulfonate, a sulfonamido,  $-(CH_2)_m-R_8$ ,  $-(CH_2)_m-OH$ ,  $-(CH_2)_m-OH$  over alkyl,  $-(CH_2)_m-OH$  over alkenyl,  $-(CH_2)_m-OH$  alkenyl,  $-(CH_2)_m-CH$  over alkenyl,  $-(CH_2)_m-CH$  or

R<sub>1</sub> and R<sub>2</sub> taken together with N form a heterocycle (substituted or unsubstituted);

 $R_3$  is absent or represents one or more substitutions to the isoquinoline ring such as a lower alkyl, a lower alkenyl, a lower alkynyl, a carbonyl (such as a carboxyl, an ester, a formate, or a ketone), a thiocarbonyl (such as a thioester, a thioacetate, or a thioformate), an amino, an acylamino, an amido, a cyano, a nitro, an azido, a sulfate, a sulfonate, a sulfonamido,  $-(CH_2)_m-R_8$ ,  $-(CH_2)_m-OH$ ,  $-(CH_2)_m-OH$  over alkyl,  $-(CH_2)_m-OH$  over alkenyl,  $-(CH_2)_m-CH$ ,  $-(CH_2)_m-$ 

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R<sub>8</sub> represents a substituted or unsubstituted aryl, aralkyl, cycloalkyl, cycloalkenyl, or heterocycle; and

n and m are independently for each occurrence zero or an integer in the range of 1 to 6.

The subject method can be used to prevent or treat various lung diseases, to control wound healing or other reformation processes in lung, and to augment lung transplantation.

Wherein the subject method is carried out using an fgf-10 therapeutic, the fgf-10 therapeutic preferably a polypeptide including a fgf-10 portion comprising at least a bioactive extracellular portion of a fgf-10 protein, e.g., the fgf-10 portion includes at least 50, 100 or 150 (contiguous) amino acid residues of a fgf-10 protein, preferably a human fgf-10 protein such as shown in SEQ ID No. 24.

In certain preferred embodiments, the fgf-10 portion has an amino acid sequence at least 60, 75, 85, or 95 percent identical with the fgf-10 protein of SEQ ID No. 24, though a sequence identical with SEQ ID No. 24 is also contemplated as useful in the present method. The fgf-10 portion can be encoded by a nucleic acid which hybridizes under stringent conditions to a nucleic acid sequence of SEQ ID No. 23, e.g., the fgf-10 portion can be encoded by a vertebrate fgf-10 gene, especially a human fgf-10 gene.

In other embodiments, the subject method can be carried out by administering a gene activation construct, wherein the gene activation construct is deigned to recombine with a genomic fgf-10 gene of the patient to provide a heterologous transcriptional regulatory sequence operatively linked to a coding sequence of the fgf-10 gene.

In still other embodiments, the subject method can be practiced with the administration of a gene therapy construct encoding a fgf-10 polypeptide. For instance, the gene therapy construct can be provided in a composition selected from a group consisting of a recombinant viral particle, a liposome, and a poly-cationic nucleic acid binding agent,

Yet another aspect of the present invention concerns preparations of a hedgehog, ptc or fgf-10 therapeutic formulated for application to lung tissue, e.g., by aerosol. For example, such formulations may include a polypeptide comprising a hedgehog polypeptide sequence including a bioactive fragment of a hedgehog protein, which polypeptide is formulated for application to lung tissue by inhalation.

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#### **Detailed Description of the Invention**

Figure 1. Morphology and epithelial phenotype of Shh -/- mouse lungs. (a) At 12.5 dpc, the wt mouse lung has branched several times to give rise to distinct lobes (arrows). (b) Trachea and esophagus are separate tubes. (c) Cross-section at the level of the lung shows branching and lobation. (d) At 12.5 dpc, Shh-deficient lungs have failed to undergo lobation or subsequent extensive branching. (e) Trachea and esophagus remain fused at the tracheoesophageal septum. (f) Mutant lungs have branched only once. (g) At 18.5 dpc, airsac formation is in progress in the wt and the respiratory surface is in tight association with blood vessels. (h) There is little branching or growth of the poorly vascularized mutant lungs, but airsac formation at the distal epithelial tips is apparent (arrows). (i) By 18.5 dpc, wild-type lungs have established the conducting airways and respiratory bronchioles, alveolar formation is in progress. (i) In contrast, in a mutant lung of the same stage, branching is dramatically decreased. Only a few primary branches (arrows) and air sacs (arrowheads) are present. (k) In the wild-type, trachea and esophagus are separated. The trachea is lined by columnar cells, the esophagus by stratified epithelium. (1) Air sacs are made of cuboidal cells. (m) In the mutant, trachea and esophagus are fused to form a fistula. Differentiation into columnar and stratified epithelium is apparent, (n) as is the characteristic cuboidal epithelium of the air sacs. Demarcation lines between terminal bronchioles and respiratory surface are indicated. (o) Proximal lung epithelium of the 18.5 dpc wt lung expresses CCSP in Clara cells, and (p) SP-C in type II pneumocytes of the distal epithelium. (q) CCSP and (p) SP-C

are expressed in the correct proximo-distal domain in the mutant. Bars denote 1 mm (g,h only) or 10  $\mu$ m. (a,d,g,h) are ventral views, all others transverse sections. Abbreviations: t - trachea, e - esophagus, l - lung, h - heart, s - stomach, mb - mainstem bronchus, b - bronchus, tb - terminal bronchiole, a - air sac.

Figure 2. In situ analysis of gene expression in the lungs of *Shh* mutants. Expression of the genes indicated was investigated in whole mount vibratome sections through lungs removed from wt 11.5 and 12.5 dpc, and Shh-mutant 12.5 dpc embryos.

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Figure 3. Mesenchyme differentiation at 18.5 dpc. (a) Both wt and mutant lungs display cartilaginous rings around the trachea as indicated by alcian-blue staining. (b) While in the wild-type lung a layer of smooth muscle surrounds the conducting epithelium, the mutant lung mesenchyme does not differentiate into muscle (right panel). Bars denote 10 mm

#### **Detailed Description of the Invention**

Development of the lung, through a process known as branching morphogenesis, is strictly dependent on interactions between endodermally derived epithelial cells and the splanchnic mesenchyme. Cell-cell interactions form the functional basis for branching morphogenesis and occur through the activity of a number of mediators, including the extracellular matrix, cellular receptors, and morphogenetic signaling molecules such as peptide growth factors. The molecular regulatory signals and in particular the role of transcriptional factors in branching morphogenesis and lung injury/repair are an important source of information for the treatment of injury. Furthermore, because the lungs continue to undergo development after birth, untimely activation of alternative morphogenetic signals released by tissue injury or repair or both may potentially derail normal morphogenesis and result in structural and functional aberrations characteristic of neonatal lung disease.

It is demonstrated herein that *hedgehog* proteins, such as *Shh*, is essential for development of the respiratory system. In *Shh* null mutants, for example, the trachea and esophagus do not separate properly and the lungs form a rudimentary sac due to failure of branching and growth after formation of the primary lung buds. Interestingly, normal

proximo-distal differentiation of the airway epithelium occurs, indicating that Shh is not needed for differentiation events. In addition, the transcription of several mesenchymally expressed downstream targets of Shh is abolished. These results highlight the importance of epithelially derived Shh in regulating branching morphogenesis of the lung, and establish a role for *hedgehog* in lung morphogenesis, disease and repair, and suggest that SHH normally regulates lung mesenchymal cell proliferation *in vivo*.

#### I. Overview

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The present application is directed to the discovery that preparations of hedgehog polypeptides can be used to control the formation and/or maintenance of lung tissue. As described in the appended examples, hedgehog proteins are implicated in the proliferation and differentiation of lung mesenchymal and epithelial cells and provide early signals that regulate the formation and maintenance of lung tissues. The present invention provides a method for regulating the growth state of lung tissue, e.g., either in in vitro or in vivo. In general, the method of the present invention comprises contacting lung tissue, or cells derived therefrom, with an amount of a hedgehog therapeutic (defined infra) which produces a nontoxic response by the cell of induction or inhibition of the formation of lung tissue microarchetecture, e.g., depending on the whether the hedgehog therapeutic is a sufficient hedgehog agonist or hedgehog antagonist. The subject method can be carried out on lung cells which may be either dispersed in culture or a part of an intact tissue or organ. Moreover, the method can be performed on cells which are provided in culture (in vitro), or on cells in a whole animal (in vivo).

Without wishing to be bound by any particular theory, the ability of hedgehog proteins to regulate the growth state of lung tissue may be due at least in part to the ability of these proteins to antagonize (directly or indirectly) patched-mediated regulation of gene expression and other physiological effects mediated by that protein. The patched gene product, a cell surface protein, is understood to signal through a pathway which causes transcriptional repression of members of the Wnt and Dpp/BMP families of morphogens, proteins which impart positional information. In development of the CNS and patterning of

limbs in vertebrates, the introduction of *hedgehog* relieves (derepresses) this inhibition conferred by *patched*, allowing expression of particular gene programs.

Recently, it has been reported that mutations in the human version of patched, a gene first identified in a fruit fly developmental pathway, cause a hereditary skin cancer and may contribute to sporadic skin cancers. See, for example, Hahn et al. (1996) Cell 86:841-851; and Johnson et al. (1996) Science 272:1668-1671. The demonstraction that nevoid basal-cell carcinoma (NBCC) results from mutations in the human patched gene provided an example of the roles patched plays in post-embryonic development. These observations have led the art to understand one activity of patched to be a tumor suppressor gene, which may act by inhibiting proliferative signals from hedgehog. Our observations set forth below reveal potential new roles for the hedgehog/patched pathway in maintenance of proliferation and differentiation of lung tissue. Accordingly, the present invention contemplates the use of other agents which are capable of mimicking the effect of the hedgehog protein on patched signalling, e.g., as may be identified from the drug screening assays described below.

Moreover, we demonstrate that fgf-10 is an important component of the hedgehog regulatory network present in the embryonic lung, controlling proliferation, differentiation and pattern formation. Accordingly, Applicants contemplate that agonists and antagonist of fgf-10 activity.

#### II. Definitions

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For convience, certain terms employed in the specification, examples, and appended claims are collected here.

The term "hedgehog therapeutic" refers to various forms of hedgehog polypeptides, as well as peptidomimetics, which can modulate the proliferation/differentiation state of lung cells by, as will be clear from the context of individual examples, mimicing or potentiating (agonizing) or inhibiting (antagonizing) the effects of a naturally-occurring hedgehog protein. A hedgehog therapeutic which mimics or potentiates the activity of a wild-type hedgehog protein is a "hedgehog agonist". Conversely, a hedgehog therapeutic which inhibits the activity of a wild-type hedgehog protein is a "hedgehog antagonist".

In particular, the term "hedgehog polypeptide" encompasses preparations of hedgehog proteins and peptidyl fragments thereof, both agonist and antagonist forms as the specific context will make clear.

As used herein the term "bioactive fragment of a hedgehog protein" refers to a fragment of a full-length hedgehog polypeptide, wherein the fragment specifically agonizes or antagonizes inductive events mediated by wild-type hedgehog proteins. The hedgehog biactive fragment preferably is a soluble extracellular portion of a hedgehog protein, where solubility is with reference to physiologically compatible solutions. Exemplary bioactive fragments are described in PCT publications WO 95/18856 and WO 96/17924.

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The term "patched" or "ptc" refers to a family of related transmembrane proteins which have been implicated in the signal transduction induced by contacting a cell with a hedgehog protein. For example, the mammalian ptc family includes ptc1 and ptc2. In addition to references set out below, see also Takabatake et al. (1997) FEBS Lett 410:485 and GenBank AB000847 for examples of ptc2. Unless otherwise evident from the context, it will be understood that embodiments described in the context of ptc1 (or just ptc) also refer to equivalent embodiments involving other ptc homologs like ptc2.

The term "ptc therapeutic" refers to agents which either (i) mimic the effect of hedgehog proteins on patched signalling, e.g., which antagonize the cell-cycle inhibitory activity of patched, or (ii) activate or potentiate patched signalling. In other embodiments, the ptc therapeutic can be a hedgehog antagonist. The ptc therapeutic can be, e.g., a peptide, a nucleic acid, a carbohydrate, a small organic molecule, or natural product extract (or fraction thereof).

The term "fgf-10 therapeutic" refers to agents which mimic or antagonize, as appropriate, the effect of fgf-10 proteins on proliferation and differentiation of lung tissue. Such agents also include small organic molecules which bind to the fgf-10 receptor and either inhibit or agonize fgf-10 signalling.

A "proliferative" form of a ptc, hedgehog or fgf-10 therapeutic is one which induces proliferation of lung cells, e.g., directly or indirectly, mesenchymal or epithelial cells. Conversely, an "antiproliferative" form of a ptc, hedgehog or fgf-10 therapeutic is one which

inhibits proliferation of lung cells, preferably in a non-toxic manner, e.g., by promoting or maintaining a differentiated phenotype or otherwise promoting quiescence.

By way of example, though not wishing to be bound by a particular theory, proliferative *hedgehog* polypeptide will generally be a form of the protein which derepresses *patched*-mediated cell-cycle arrest, e.g., the polypeptide mimics the effect of a naturally occurring *hedgehog* protein effect on lung tissues. A proliferative *ptc* therapeutic includes other agents which depress *patched*-mediated cell-cycle arrest, and may act extracellularly or intracellularly.

An illustrative antiproliferative *pic* therapeutic agent may potentiate *patched*-mediated cell-cycle arrest. Such agents can be small molecules that inhibit, e.g., *hedgehog* binding to *patched*, as well as agents which stimulate and/or potentiate a signal transduction pathway of the *patched* protein.

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As used herein, "proliferating" and "proliferation" refer to cells undergoing mitosis.

As used herein, "transformed cells" refers to cells which have spontaneously converted to a state of unrestrained growth, i.e., they have acquired the ability to grow through an indefinite number of divisions in culture. Transformed cells may be characterized by such terms as neoplastic, anaplastic and/or hyperplastic, with respect to their loss of growth control.

As used herein, "immortalized cells" refers to cells which have been altered via chemical and/or recombinant means such that the cells have the ability to grow through an indefinite number of divisions in culture.

A "patient" or "subject" to be treated by the subject method can mean either a human or non-human animal.

An "effective amount" of, e.g., a hedgehog therapeutic, with respect to the subject method of treatment, refers to an amount of, e.g., a hedgehog polypeptide in a preparation which, when applied as part of a desired dosage regimen brings about a change in the rate of cell proliferation and/or the state of differentiation of a cell so as to produce (or inhibit as the case may be) proliferation of lung cells in an amount according to clinically acceptable standards for the disorder to be treated or the cosmetic purpose.

The "growth state" of a cell refers to the rate of proliferation of the cell and the state of differentiation of the cell.

"Homology" and "identity" each refer to sequence similarity between two polypeptide sequences, with identity being a more strict comparison. Homology and identity can each be determined by comparing a position in each sequence which may be aligned for purposes-of comparison. When a position in the compared sequence is occupied by the same amino acid residue, then the polypeptides can be referred to as identical at that position; when the equivalent site is occupied by the same amino acid (e.g., identical) or a similar amino acid (e.g., similar in steric and/or electronic nature), then the molecules can be referred to as homologous at that position. A percentage of homology or identity between sequences is a function of the number of matching or homologous positions shared by the sequences. An "unrelated" or "non-homologous" sequence shares less than 40 percent identity, though preferably less than 25 percent identity, with a hedgehog sequence disclosed herein.

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The term "corresponds to", when referring to a particular polypeptide or nucleic acid sequence is meant to indicate that the sequence of interest is identical or homologous to the reference sequence to which it is said to correspond.

The terms "recombinant protein", "heterologous protein" and "exogenous protein" are used interchangeably throughout the specification and refer to a polypeptide which is produced by recombinant DNA techniques, wherein generally, DNA encoding the polypeptide is inserted into a suitable expression construct which is in turn used to transform a host cell to produce the heterologous protein. That is, the polypeptide is expressed from a heterologous nucleic acid.

A "chimeric protein" or "fusion protein" is a fusion of a first amino acid sequence encoding a *hedgehog* polypeptide with a second amino acid sequence defining a domain foreign to and not substantially homologous with any domain of hh protein. A chimeric protein may present a foreign domain which is found (albeit in a different protein) in an organism which also expresses the first protein, or it may be an "interspecies", "intergenic", etc. fusion of protein structures expressed by different kinds of organisms. In general, a fusion protein can be represented by the general formula  $(X)_n$ - $(hh)_m$ - $(Y)_n$ , wherein hh represents all or a portion of the *hedgehog* protein, X and Y each independently represent an

amino acid sequences which are not naturally found as a polypeptide chain contiguous with the *hedgehog* sequence, m is an integer greater than or equal to 1, and each occurrence of n is, independently, 0 or an integer greater than or equal to 1 (n and m are preferably no greater than 5 or 10).

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## III. Exemplary Applications of Method and Compositions

The subject method has wide applicability to the treatment or prophylaxis of disorders afflicting lung tissue, as well as in *in vitro* cultures. In general, the method can be characterized as including a step of administering to an animal an amount of a *ptc. hedgehog* or *fgf-10* therapeutic effective to alter the growth state of a treated lung tissue. The mode of administration and dosage regimens will vary depending on the phenotype of, and desired effect on the target lung tissue. Likewise, as described in further detail below, the use of a particular *ptc, hedgehog* or *fgf-10* therapeutic, e.g., an agonist or antagonist, will depend on whether proliferation of cells in the treated lung tissue is desired or intended to be prevented.

In one aspect, the present invention provides pharmaceutical preparations and methods for controlling the proliferation of lung tissue utilizing, as an active ingredient, a hedgehog polypeptide or a mimetic thereof. The invention also relates to methods of controlling proliferation of mesenchymal and epithelial cells of the tissue by use of the pharmaceutical preparations of the invention.

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The formulations of the present invention may be used as part of regimens in the treatment of disorders of, surgical repair of, or transplantation of lung tissues and whole organs. The methods and compositions disclosed herein also provide for the treatment of a variety of proliferative cancerous disorders effecting lung tissue. For instance, the subject method can be used to control wound healing processes, as for example may be desirable in connection with any surgery involving lung tissue.

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In certain embodiments, the subject compositions can be used to inhibit, rather than promote, growth of lung-derived tissue. For instance, certain of the compositions disclosed herein may be applied to the treatment or prevention of a variety hyperplastic or neoplastic conditions. The method can find application for the treatment or prophylaxis of, e.g., used to

inhibit the growth and metastasis of lung cancer cells. For instance, inhibitory forms of the the subject ptc, hedgehog and fgf-10 therapeutics may be used as part of a treatment program for small cell lung cancer (SCLC), as well as non-small cell lung cancer (NSCLC), such as adenocarcinoma, lung cell carcinoma and squamus cell carcinoma.

In other embodiments, the subject method can be used to treat rheumatoid—lung disease, which may be marked by pleural thickening, adhesions, and pleural effusions. Such lung (pulmonary) manifestations can occur in both adult and juvenile forms of rheumatoid arthritis.

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In other embodiments, the subject method can be used to treat, or lessen the severity of, damage to lung tissue as a complication of respiratory diseases such as bronchopneumonia, chronic bronchitis, cystic fibrosis and asthma, and bronchospasm, or other apical interstitial lung diseases, such as cystic fibrosis, ankylosing spondylitis, sarcoidosis, silicosis, eosinophlic granuloma, tuberculosis, and lung infections.

In certain embodiments, the subject method can be used to treat or prevent damage to lung tissue resulting from allergic rhinitis, asthma, emphysema, chronic bronchitis, pneumoconiosis, respiratory distress syndrome, idiopathic pulmonary fibrosis and primary pulmonary hypertension

The subject method can be used in the treatment or prevention of occupational lung disease such as asbestos-related diseases, silicosis, occupational asthma, coal worker's pneumoconiosis, berylliosis, and industrial bronchitis.

In still other embodiments, the subject method can be used to treat certain health consequences of smoking which may result in degeneration of lung tissue.

The subject hedgehog treatments are effective on both human and animal subjects afflicted with these conditions. Animal subjects to which the invention is applicable extend to both domestic animals and livestock, raised either as pets or for commercial purposes. Examples are dogs, cats, cattle, horses, sheep, hogs and goats.

Still another aspect of the present invention provides a method of stimulating the growth and regulating the differentiation of epithelial tissue in tissue culture.

In one embodiment, the subject method can be used to regulate the proliferation and/or differentiation of lung mesenchymal progenitor cells.

The maintenance of lung tissues and whole organs ex vivo is also highly desirable. Lung and heart-lung transplantation therapy is well established in the treatment of certain human disease. The subject method can be used to maintain the tissue structure of lung tissue ex vivo, and in certain embodiments to accelerate the growth of certain lung tissue in vitro. The present method can also be used for improving the "take rate" of a lung transplants in vivo.

# IV. Exemplary hedgehog therapeutic compounds.

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The hedgehog therapeutic compositions of the subject method can be generated by any of a variety of techniques, including purification of naturally occurring proteins, recombinantly produced proteins and synthetic chemistry. Polypeptide forms of the hedgehog therapeutics are preferably derived from vertebrate hedgehog proteins, e.g., have sequences corresponding to naturally occurring hedgehog proteins, or fragments thereof, from vertebrate organisms. However, it will be appreciated that the hedgehog polypeptide can correspond to a hedgehog protein (or fragment thereof) which occurs in any metazoan organism.

The various naturally-occurring hedgehog proteins from which the subject therapeutics can be derived are characterized by a signal peptide, a highly conserved N-terminal region, and a more divergent C-terminal domain. In addition to signal sequence cleavage in the secretory pathway (Lee, J.J. et al. (1992) Cell 71:33-50; Tabata, T. et al. (1992) Genes Dev. 2635-2645; Chang, D.E. et al. (1994) Development 120:3339-3353), hedgehog precursor proteins naturally undergo an internal autoproteolytic cleavage which depends on conserved sequences in the C-terminal portion (Lee et al. (1994) Science 266:1528-1537; Porter et al. (1995) Nature 374:363-366). This autocleavage leads to a 19 kD N-terminal peptide and a C-terminal peptide of 26-28 kD (Lee et al. (1992) supra; Tabata et al. (1992) supra; Chang et al. (1994) supra; Lee et al. (1994) supra; Bumcrot, D.A., et al. (1995) Mol. Cell. Biol. 15:2294-2303; Porter et al. (1995) supra; Ekker, S.C. et al. (1995)

Curr. Biol. 5:944-955; Lai, C.J. et al. (1995) Development 121:2349-2360). The N-terminal peptide stays tightly associated with the surface of cells in which it was synthesized, while the C-terminal peptide is freely diffusible both in vitro and in vivo (Lee et al. (1994) supra; Bumcrot et al. (1995) supra; Mart., E. et al. (1995) Development 121:2537-2547; Roelink, H. et al. (1995) Cell 81:445-455). Cell surface retention of the N-terminal peptide is dependent on autocleavage, as a truncated form of hedgehog encoded by an RNA which terminates precisely at the normal position of internal cleavage is diffusible in vitro (Porter et al. (1995) supra) and in vivo (Porter, J.A. et al. (1996) Cell 86, 21-34). Biochemical studies have shown that the autoproteolytic cleavage of the hedgehog precursor protein proceeds through an internal thioester intermediate which subsequently is cleaved in a nucleophilic substitution. It is suggested that the nucleophile is a small lipophilic molecule, more particularly cholesterol, which becomes covalently bound to the C-terminal end of the N-peptide (Porter et al. (1996) supra), tethering it to the cell surface.

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The vertebrate family of hedgehog genes includes at least four members, e.g., paralogs of the single drosophila hedgehog gene (SEQ ID No. 19). Three of these members, herein referred to as Desert hedgehog (Dhh), Sonic hedgehog (Shh) and Indian hedgehog (Ihh), apparently exist in all vertebrates, including fish, birds, and mammals. A fourth member, herein referred to as tiggie-winkle hedgehog (Thh), appears specific to fish. According to the appended sequence listing, (see also Table 1) a chicken Shh polypeptide is encoded by SEQ ID No:1; a mouse Dhh polypeptide is encoded by SEQ ID No:2; a mouse Ihh polypeptide is encoded by SEQ ID No:3; a mouse Shh polypeptide is encoded by SEQ ID No:6; a human Shh polypeptide is encoded by SEQ ID No:7; a human Dhh polypeptide is encoded by SEQ ID No:7; a human Dhh polypeptide is encoded by SEQ ID No:7; a human Dhh polypeptide is encoded by SEQ ID No:8; and a zebrafish Thh is encoded by SEQ ID No.9.

Table 1

Guide to hedgehog sequences in Sequence Listing

	Nucleotide	Amino Acid
Chicken Shh	SEQ ID No. 1	SEQ ID No. 10
Mouse Dhh	SEQ ID No. 2	SEQ ID No. 11
Mouse Ihh	SEQ ID No. 3	SEQ ID No. 12
Mouse Shh	SEQ ID No. 4	SEQ ID No. 13
Zebrafish Shh	SEQ ID No. 5	SEQ ID No. 14
Human Shh	SEQ ID No. 6	SEQ ID No. 15
Human Ihh	SEQ ID No. 7	SEQ ID No. 16
Human Dhh	SEQ ID No. 8	SEQ ID No. 17
Zebrafish Thh	SEQ ID No. 9	SEQ ID No. 18
Drosophila HH	SEQ ID No. 19	SEQ ID No. 20

In addition to the sequence variation between the various *hedgehog* homologs, the *hedgehog* proteins are apparently present naturally in a number of different forms, including a pro-form, a full-length mature form, and several processed fragments thereof. The pro-form includes an N-terminal signal peptide for directed secretion of the extracellular domain, while the full-length mature form lacks this signal sequence.

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As described above, further processing of the mature form occurs in some instances to yield biologically active fragments of the protein. For instance, *sonic hedgehog* undergoes additional proteolytic processing to yield two peptides of approximately 19 kDa and 27 kDa, the 19kDa fragment corresponding to an proteolytic N-terminal portion of the mature protein.

In addition to proteolytic fragmentation, the vertebrate *hedgehog* proteins can also be modified post-translationally, such as by glycosylation and/or addition of lipophilic moieties,

such as stents, fatty acids, etc., though bacterially produced (e.g. unmodified) forms of the proteins still maintain certain of the bioactivities of the native protein. Bioactive fragments of *hedgehog* polypeptides of the present invention have been generated and are described in great detail in, e.g., PCT publications WO 95/18856 and WO 96/17924.

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There are a wide range of lipophilic moieties with which *hedgehog* polypeptides can be derivatived. The term "lipophilic group", in the context of being attached to a *hedgehog* polypeptide, refers to a group having high hydrocarbon content thereby giving the group high affinity to lipid phases. A lipophilic group can be, for example, a relatively long chain alkyl or cycloalkyl (preferably n-alkyl) group having approximately 7 to 30 carbons. The alkyl group may terminate with a hydroxy or primary amine "tail". To further illustrate, lipophilic molecules include naturally-occurring and synthetic aromatic and non-aromatic moieties such as fatty acids, sterols, esters and alcohols, other lipid molecules, cage structures such as adamantane and buckminsterfullerenes, and aromatic hydrocarbons such as benzene, perylene, phenanthrene, anthracene, naphthalene, pyrene, chrysene, and naphthacene.

In one embodiment, the *hedgehog* polypeptide is modified with one or more sterol moieties, such as cholesterol. See, for example, PCT publication WO 96/17924. In certain embodiments, the cholesterol is preferably added to the C-terminal glycine were the *hedgehog* polypeptide corresponds to the naturally-occurring N-terminal proteolytic fragment.

In another embodiment, the *hedgehog* polypeptide can be modified with a fatty acid moiety, such as a myrostoyl, palmitoyl, stearoyl, or arachidoyl moiety. See, e.g., Pepinsky et al. (1998) J Biol. Chem 273: 14037.

In addition to those effects seen by cholesterol-addition to the C-terminus or fatty acid addition to the N-terminus of extracellular fragments of the protein, at least certain of the biological activities of the *hedgehog* gene products can potentiated by derivativation of the protein with lipophilic moieties at other sites on the protein and/or by moieties other than cholesterol or fatty acids. Certain aspects of the invention are directed to the use of preparations of *hedgehog* polypeptides which are modified at sites other than N-terminal or C-terminal residues of the natural processed form of the protein, and/or which are modified at

such terminal residues with lipophilic moieties other than a sterol at the C-terminus or fatty acid at the N-terminus.

Particularly useful as lipophilic molecules are alicyclic hydrocarbons, saturated and unsaturated fatty acids and other lipid and phospholipid moieties, waxes, cholesterol, isoprenoids, terpenes and polyalicyclic hydrocarbons including adamantane and buckminsterfullerenes, vitamins, polyethylene glycol or oligoethylene glycol, (C1-C18)-alkyl phosphate diesters, -O-CH2-CH(OH)-O-(C12-C18)-alkyl, and in particular conjugates with pyrene derivatives. The lipophilic moiety can be a lipophilic dye suitable for use in the invention include, but are not limited to, diphenylhexatriene, Nile Red, N-phenyl-1-Laurodan, Pyrene, Perylene, rhodamine, rhodamine B, naphthylamine, Prodan, Red, 1,1'-didodecyltetramethylrhodamine, Texas sulforhodamine, 3,3,3',3'tetramethylindocarbocyanine perchlorate, octadecyl rhodamine B and the BODIPY dyes available from Molecular Probes Inc.

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Other exemplary lipophilic moietites include aliphatic carbonyl radical groups include 1- or 2-adamantylacetyl, 3-methyladamant-1-ylacetyl, 3-methyl-3-bromo-1-adamantylacetyl, 1-decalinacetyl, camphoracetyl, camphaneacetyl, noradamantylacetyl, norbornaneacetyl, bicyclo[2.2.2.]-oct-5-ene-2-carbonyl, cis-5-norbornene-endo-2,3-dicarbonyl, 5-norbornen-2-ylacetyl, (1R)-( - )-myrtentaneacetyl, 2-norbornaneacetyl, anti-3-oxo-tricyclo[2.2.1.0<2,6> ]-heptane-7-carbonyl, decanoyl, dodecanoyl, dodecanoyl, decynoyl or dodecynoyl.

The *hedgehog* polypeptide can be linked to the hydrophobic moiety in a number of ways including by chemical coupling means, or by genetic engineering.

Moreover, mutagenesis can be used to create modified *hh* polypeptides, e.g., for such purposes as enhancing therapeutic or prophylactic efficacy, or stability (e.g., *ex vivo* shelf life and resistance to proteolytic degradation *in vivo*). Such modified peptides can be produced, for instance, by amino acid substitution, deletion, or addition. Modified *hedgehog* polypeptides can also include those with altered post-translational processing relative to a naturally occurring *hedgehog* protein, e.g., altered glycosylation, cholesterolization, prenylation and the like.

In one embodiment, the *hedgehog* therapeutic is a polypeptide encodable by a nucleotide sequence that hybridizes under stringent conditions to a *hedgehog* coding sequence represented in one or more of SEQ ID Nos:1-7. Appropriate stringency conditions which promote DNA hybridization, for example, 6.0 x sodium chloride/sodium citrate (SSC) at about 45°C, followed by a wash of 2.0 x SSC at 50°C, are known to those skilled in the art or can be found in *Current Protocols in Molecular Biology*, John Wiley & Sons, N.Y. (1989), 6.3.1-6.3.6. For example, the salt concentration in the wash step can be selected from a low stringency of about 2.0 x SSC at 50°C to a high stringency of about 0.2 x SSC at 50°C. In addition, the temperature in the wash step can be increased from low stringency conditions at room temperature, about 22°C, to high stringency conditions at about 65°C.

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As described in the literature, genes for other *hedgehog* proteins, e.g., from other animals, can be obtained from mRNA or genomic DNA samples using techniques well known in the art. For example, a cDNA encoding a *hedgehog* protein can be obtained by isolating total mRNA from a cell, e.g. a mammalian cell, e.g. a human cell, including embryonic cells. Double stranded cDNAs can then be prepared from the total mRNA, and subsequently inserted into a suitable plasmid or bacteriophage vector using any one of a number of known techniques. The gene encoding a *hedgehog* protein can also be cloned using established polymerase chain reaction techniques.

Preferred nucleic acids encode a *hedgehog* polypeptide comprising an amino acid sequence at least 60% homologous or identical, more preferably 70% homologous or identical, and most preferably 80% homologous or identical with an amino acid sequence selected from the group consisting of SEQ ID Nos:8-14. Nucleic acids which encode polypeptides at least about 90%, more preferably at least about 95%, and most preferably at least about 98-99% homology or identity with an amino acid sequence represented in one of SEO ID Nos:8-14 are also within the scope of the invention.

In addition to native *hedgehog* proteins, hedgehog polypeptides preferred by the present invention are at least 60% homologous or identical, more preferably 70% homologous or identical and most preferably 80% homologous or identical with an amino acid sequence represented by any of SEQ ID Nos:8-14. Polypeptides which are at least 90%, more preferably at least 95%, and most preferably at least about 98-99% homologous or

identical with a sequence selected from the group consisting of SEQ ID Nos:8-14 are also within the scope of the invention. The only prerequisite is that the *hedgehog* polypeptide is capable of modulating the growth of lung cells.

The term "recombinant protein" refers to a polypeptide of the present invention which is produced by recombinant DNA techniques, wherein generally, DNA encoding a hedgehog polypeptide is inserted into a suitable expression vector which is in turn used to transform a host cell to produce the heterologous protein. Moreover, the phrase "derived from", with respect to a recombinant hedgehog gene, is meant to include within the meaning of "recombinant protein" those proteins having an amino acid sequence of a native hedgehog protein, or an amino acid sequence similar thereto which is generated by mutations including substitutions and deletions (including truncation) of a naturally occurring form of the protein.

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The method of the present invention can also be carried out using variant forms of the naturally occurring *hedgehog* polypeptides, e.g., mutational variants.

As is known in the art, hedgehog polypeptides can be produced by standard biological techniques or by chemical synthesis. For example, a host cell transfected with a nucleic acid vector directing expression of a nucleotide sequence encoding the subject polypeptides can be cultured under appropriate conditions to allow expression of the peptide to occur. The polypeptide hedgehog may be secreted and isolated from a mixture of cells and medium containing the recombinant hedgehog polypeptide. Alternatively, the peptide may be retained cytoplasmically by removing the signal peptide sequence from the recombinant hedgehog gene and the cells harvested, lysed and the protein isolated. A cell culture includes host cells, media and other byproducts. Suitable media for cell culture are well known in the art. The recombinant hedgehog polypeptide can be isolated from cell culture medium, host cells, or both using techniques known in the art for purifying proteins including ion-exchange chromatography, gel filtration chromatography, ultrafiltration, electrophoresis, and immunoaffinity purification with antibodies specific for such peptide. In a preferred embodiment, the recombinant hedgehog polypeptide is a fusion protein containing a domain which facilitates its purification, such as an hedgehog/GST fusion protein. The host cell may be any prokaryotic or eukaryotic cell.

Recombinant hedgehog genes can be produced by ligating nucleic acid encoding an hedgehog protein, or a portion thereof, into a vector suitable for expression in either prokaryotic cells, eukaryotic cells, or both. Expression vectors for production of recombinant forms of the subject hedgehog polypeptides include plasmids and other vectors. For instance, suitable vectors for the expression of a hedgehog polypeptide include plasmids of the types: pBR322-derived plasmids, pEMBL-derived plasmids, pEX-derived plasmids, pBTac-derived plasmids and pUC-derived plasmids for expression in prokaryotic cells, such as E. coli.

A number of vectors exist for the expression of recombinant proteins in yeast. For instance, YEP24, YIP5, YEP51, YEP52, pYES2, and YRP17 are cloning and expression vehicles useful in the introduction of genetic constructs into *S. cerevisiae* (see, for example, Broach et al. (1983) in Experimental Manipulation of Gene Expression, ed. M. Inouye Academic Press, p. 83, incorporated by reference herein). These vectors can replicate in *E. coli* due to the presence of the pBR322 ori, and in *S. cerevisiae* due to the replication determinant of the yeast 2 micron plasmid. In addition, drug resistance markers such as ampicillin can be used. In an illustrative embodiment, an hedgehog polypeptide is produced recombinantly utilizing an expression vector generated by sub-cloning the coding sequence of one of the hedgehog genes represented in SEQ ID Nos:1-7.

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The preferred mammalian expression vectors contain both prokaryotic sequences, to facilitate the propagation of the vector in bacteria, and one or more eukaryotic transcription units that are expressed in eukaryotic cells. The pcDNAI/amp, pcDNAI/neo, pRc/CMV, pSV2gpt, pSV2neo, pSV2-dhfr, pTk2, pRSVneo, pMSG, pSVT7, pko-neo and pHyg derived vectors are examples of mammalian expression vectors suitable for transfection of eukaryotic cells. Some of these vectors are modified with sequences from bacterial plasmids, such as pBR322, to facilitate replication and drug resistance selection in both prokaryotic and eukaryotic cells. Alternatively, derivatives of viruses such as the bovine papillomavirus (BPV-1), or Epstein-Barr virus (pHEBo, pREP-derived and p205) can be used for transient expression of proteins in eukaryotic cells. The various methods employed in the preparation of the plasmids and transformation of host organisms are well known in the art. For other suitable expression systems for both prokaryotic and eukaryotic cells, as well as general recombinant procedures, see *Molecular Cloning A Laboratory Manual*, 2nd Ed., ed. by

Sambrook, Fritsch and Maniatis (Cold Spring Harbor Laboratory Press: 1989) Chapters 16 and 17.

In some instances, it may be desirable to express the recombinant *hedgehog* polypeptide by the use of a baculovirus expression system. Examples of such baculovirus expression systems include pVL-derived vectors (such as pVL1392, pVL1393 and pVL941), pAcUW-derived vectors (such as pAcUW1), and pBlueBac-derived vectors (such as the \(\beta\)-gal containing pBlueBac III).

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When it is desirable to express only a portion of an *hedgehog* protein, such as a form lacking a portion of the N-terminus, i.e. a truncation mutant which lacks the signal peptide, it may be necessary to add a start codon (ATG) to the oligonucleotide fragment containing the desired sequence to be expressed. It is well known in the art that a methionine at the N-terminal position can be enzymatically cleaved by the use of the enzyme methionine aminopeptidase (MAP). MAP has been cloned from *E. coli* (Ben-Bassat et al. (1987) *J. Bacteriol.* 169:751-757) and *Salmonella typhimurium* and its *in vitro* activity has been demonstrated on recombinant proteins (Miller et al. (1987) *PNAS* 84:2718-1722). Therefore, removal of an N-terminal methionine, if desired, can be achieved either *in vivo* by expressing *hedgehog*-derived polypeptides in a host which produces MAP (e.g., *E. coli* or CM89 or *S. cerevisiae*), or *in vitro* by use of purified MAP (e.g., procedure of Miller et al., *supra*).

Alternatively, the coding sequences for the polypeptide can be incorporated as a part of a fusion gene including a nucleotide sequence encoding a different polypeptide. It is widely appreciated that fusion proteins can also facilitate the expression of proteins, and accordingly, can be used in the expression of the *hedgehog* polypeptides of the present invention. For example, *hedgehog* polypeptides can be generated as glutathione-S-transferase (GST-fusion) proteins. Such GST-fusion proteins can enable easy purification of the *hedgehog* polypeptide, as for example by the use of glutathione-derivatized matrices (see, for example, *Current Protocols in Molecular Biology*, eds. Ausubel et al. (N.Y.: John Wiley & Sons, 1991)). In another embodiment, a fusion gene coding for a purification leader sequence, such as a poly-(His)/enterokinase cleavage site sequence, can be used to replace the signal sequence which naturally occurs at the N-terminus of the *hedgehog* protein (e.g.of the pro-form, in order to permit purification of the poly(His)-*hedgehog* protein by affinity

chromatography using a Ni<sup>2+</sup> metal resin. The purification leader sequence can then be subsequently removed by treatment with enterokinase (e.g., see Hochuli et al. (1987) *J. Chromatography* 411:177; and Janknecht et al. *PNAS* 88:8972).

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Techniques for making fusion genes are known to those skilled in the art. Essentially, the joining of various DNA fragments coding for different polypeptide sequences is performed in accordance with conventional techniques, employing blunt-ended or stagger-ended termini for ligation, restriction enzyme digestion to provide for appropriate termini, filling-in of cohesive ends as appropriate, alkaline phosphatase treatment to avoid undesirable joining, and enzymatic ligation. In another embodiment, the fusion gene can be synthesized by conventional techniques including automated DNA synthesizers. Alternatively, PCR amplification of gene fragments can be carried out using anchor primers which give rise to complementary overhangs between two consecutive gene fragments which can subsequently be annealed to generate a chimeric gene sequence (see, for example, Current Protocols in Molecular Biology, eds. Ausubel et al. John Wiley & Sons: 1992).

Hedgehog polypeptides may also be chemically modified to create hedgehog derivatives by forming covalent or aggregate conjugates with other chemical moieties, such as glycosyl groups, cholesterol, isoprenoids, lipids, phosphate, acetyl groups and the like. Covalent derivatives of hedgehog proteins can be prepared by linking the chemical moieties to functional groups on amino acid sidechains of the protein or at the N-terminus or at the C-terminus of the polypeptide.

For instance, *hedgehog* proteins can be generated to include a moiety, other than sequence naturally associated with the protein, that binds a component of the extracellular matrix and enhances localization of the analog to cell surfaces. For example, sequences derived from the fibronectin "type-III repeat", such as a tetrapeptide sequence R-G-D-S (Pierschbacher et al. (1984) *Nature* 309:30-3; and Kornblihtt et al. (1985) *EMBO* 4:1755-9) can be added to the *hedgehog* polypeptide to support attachment of the chimeric molecule to a cell through binding ECM components (Ruoslahti et al. (1987) *Science* 238:491-497; Pierschbacheret al. (1987) *J. Biol. Chem.* 262:17294-8.; Hynes (1987) *Cell* 48:549-54; and Hynes (1992) *Cell* 69:11-25).

In a preferred embodiment, the *hedgehog* polypeptide is isolated from, or is otherwise substantially free of, other cellular proteins, especially other extracellular or cell surface associated proteins which may normally be associated with the hedgehog polypeptide, unless provided in the form of fusion protein with the hedgehog polypeptide. The term "substantially free of other cellular or extracellular proteins" (also referred to herein as "contaminating proteins") or "substantially pure preparations" or "purified preparations" are defined as encompassing preparations of hedgehog polypeptides having less than 20% (by dry weight) contaminating protein, and preferably having less than 5% contaminating protein. By "purified", it is meant that the indicated molecule is present in the substantial absence of other biological macromolecules, such as other proteins. The term "purified" as used herein preferably means at least 80% by dry weight, more preferably in the range of 95-99% by weight, and most preferably at least 99.8% by weight, of biological macromolecules of the same type present (but water, buffers, and other small molecules, especially molecules having a molecular weight of less than 5000, can be present). The term "pure" as used herein preferably has the same numerical limits as "purified" immediately above.

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As described above for recombinant polypeptides, isolated *hedgehog* polypeptides can include all or a portion of the amino acid sequences represented in any of SEQ ID Nos:10-18 or 20, or a homologous sequence thereto. Preferred fragments of the subject *hedgehog* proteins correspond to the N-terminal and C-terminal proteolytic fragments of the mature protein. Bioactive fragments of *hedgehog* polypeptides are described in great detail in PCT publications WO 95/18856 and WO 96/17924.

With respect to bioctive fragments of *hedgehog* polypeptide, preferred *hedgehog* therapeutics include at least 50 (contiguous) amino acid residues of a *hedgehog* polypeptide, more preferably at least 100 (contiguous), and even more preferably at least 150 (contiguous) residues.

Another preferred *hedgehog* polypeptide which can be included in the *hedgehog* therapeutic is an N-terminal fragment of the mature protein having a molecular weight of approximately 19 kDa.

Preferred human *hedgehog* proteins include N-terminal fragments corresponding approximately to residues 24-197 of SEQ ID No. 15, 28-202 of SEQ ID No. 16, and 23-198

of SEQ ID No. 17. By "corresponding approximately" it is meant that the sequence of interest is at most 20 amino acid residues different in length to the reference sequence, though more preferably at most 5, 10 or 15 amino acid different in length.

As described above for recombinant polypeptides, isolated *hedgehog* polypeptides can include all or a portion of the amino acid sequences represented in SEQ ID No:8, SEQ ID No:9, SEQ ID No:10, SEQ ID No:11, SEQ ID No:12, SEQ ID No:13 or SEQ ID No:14, or a homologous sequence thereto. Preferred fragments of the subject *hedgehog* proteins correspond to the N-terminal and C-terminal proteolytic fragments of the mature protein. Bioactive fragments of *hedgehog* polypeptides are described in great detail in PCT publications WO 95/18856 and WO 96/17924.

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Still other preferred hedgehog polypeptides includes an amino acid sequence represented by the formula A-B wherein: (i) A represents all or the portion of the amino acid sequence designated by residues 1-168 of SEQ ID No:21; and B represents at least one amino acid residue of the amino acid sequence designated by residues 169-221 of SEQ ID No:21; (ii) A represents all or the portion of the amino acid sequence designated by residues 24-193 of SEQ 1D No:15; and B represents at least one amino acid residue of the amino acid sequence designated by residues 194-250 of SEQ ID No:15; (iii) A represents all or the portion of the amino acid sequence designated by residues 25-193 of SEQ ID No:13; and B represents at least one amino acid residue of the amino acid sequence designated by residues 194-250 of SEQ ID No:13; (iv) A represents all or the portion of the amino acid sequence designated by residues 23-193 of SEQ ID No.11; and B represents at least one amino acid residue of the amino acid sequence designated by residues 194-250 of SEQ ID No:11; (v) A represents all or the portion of the amino acid sequence designated by residues 28-197 of SEQ ID No:12; and B represents at least one amino acid residue of the amino acid sequence designated by residues 198-250 of SEQ ID No:12; (vi) A represents all or the portion of the amino acid sequence designated by residues 29-197 of SEQ ID No:16; and B represents at least one amino acid residue of the amino acid sequence designated by residues 198-250 of SEQ ID No:16; or (vii) A represents all or the portion of the amino acid sequence designated by residues 23-193 of SEQ ID No. 17, and B represents at least one amino acid residue of the amino acid sequence designated by residues 194-250 of SEQ ID No. 17. In certain preferred

embodiments, A and B together represent a contiguous polypeptide sequence designated sequence, A represents at least 25, 50, 75, 100, 125 or 150 (contiguous) amino acids of the designated sequence, and B represents at least 5, 10, or 20 (contiguous) amino acid residues of the amino acid sequence designated by corresponding entry in the sequence listing, and A and B together preferably represent a contiguous sequence corresponding to the sequence listing entry. Similar fragments from other *hedgehog* also contemplated, e.g., fragments which correspond to the preferred fragments from the sequence listing entries which are enumerated above. In preferred embodiments, the *hedgehog* polypeptide includes a C-terminal glycine (or other appropriate residue) which is derivatized with a cholesterol.

Isolated peptidyl portions of *hedgehog* proteins can be obtained by screening peptides recombinantly produced from the corresponding fragment of the nucleic acid encoding such peptides. In addition, fragments can be chemically synthesized using techniques known in the art such as conventional Merrifield solid phase f-Moc or t-Boc chemistry. For example, a *hedgehog* polypeptide of the present invention may be arbitrarily divided into fragments of desired length with no overlap of the fragments, or preferably divided into overlapping fragments of a desired length. The fragments can be produced (recombinantly or by chemical synthesis) and tested to identify those peptidyl fragments which can function as either agonists or antagonists of a wild-type (e.g., "authentic") *hedgehog* protein. For example, Román et al. (1994) *Eur J Biochem* 222:65-73 describe the use of competitive-binding assays using short, overlapping synthetic peptides from larger proteins to identify binding domains.

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The recombinant *hedgehog* polypeptides of the present invention also include homologs of the authentic *hedgehog* proteins, such as versions of those protein which are resistant to proteolytic cleavage, as for example, due to mutations which alter potential cleavage sequences or which inactivate an enzymatic activity associated with the protein. *Hedgehog* homologs of the present invention also include proteins which have been post-translationally modified in a manner different than the authentic protein. Exemplary derivatives of *hedgehog* proteins include polypeptides which lack N-glycosylation sites (e.g. to produce an unglycosylated protein), which lack sites for cholesterolization, and/or which lack N-terminal and/or C-terminal sequences.

Modification of the structure of the subject hedgehog polypeptides can also be for such purposes as enhancing therapeutic or prophylactic efficacy, or stability (e.g., ex vivo shelf life and resistance to proteolytic degradation in vivo). Such modified peptides, when designed to retain at least one activity of the naturally-occurring form of the protein, are considered functional equivalents of the hedgehog polypeptides described in more detail herein. Such modified peptides can be produced, for instance, by amino acid substitution, deletion, or addition.

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It is well known in the art that one could reasonably expect that certain isolated replacements of amino acids, e.g., replacement of an amino acid residue with another related amino acid (i.e. isosteric and/or isoelectric mutations), can be carried out without major effect on the biological activity of the resulting molecule. Conservative replacements are those that take place within a family of amino acids that are related in their side chains. Genetically encoded amino acids are can be divided into four families: (1) acidic = aspartate, glutamate; (2) basic = lysine, arginine, histidine; (3) nonpolar = alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan; and (4) uncharged polar = glycine, asparagine, glutamine, cysteine, serine, threonine, tyresine. Phenylalanine, tryptophan, and tyrosine are sometimes classified jointly as aromatic amino acids. In similar fashion, the amino acid repertoire can be grouped as (1) acidic = aspartate, glutamate; (2) basic = lysine, arginine histidine, (3) aliphatic = glycine, alanine, valine, leucine, isoleucine, serine, threonine, with serine and threonine optionally be grouped separately as aliphatic-hydroxyl; (4) aromatic = phenylalanine, tyrosine, tryptophan: (5) amide = asparagine, glutamine; and (6) sulfur -containing = cysteine and methionine. (see, for example, Biochemistry, 2nd ed., Ed. by L. Stryer, WH Freeman and Co.: 1981). Whether a change in the amino acid sequence of a peptide results in a functional hedgehog homolog (e.g. functional in the sense that it acts to mimic or antagonize the wild-type form) can be readily determined by assessing the ability of the variant peptide to produce a response in cells in a fashion similar to the wild-type protein, or competitively inhibit such a response. Polypeptides in which more than one replacement has taken place can readily be tested in the same manner.

It is specifically contemplated that the methods of the present invention can be carried using homologs of naturally occurring *hedgehog* proteins. In one embodiment, the invention

contemplates using hedgehog polypeptides generated by combinatorial mutagenesis. Such methods, as are known in the art, are convenient for generating both point and truncation mutants, and can be especially useful for identifying potential variant sequences (e.g. homologs) that are functional in binding to a receptor for hedgehog proteins. The purpose of screening such combinatorial libraries is to generate, for example, novel hedgehog homologs which can act as either agonists or antagonist. To illustrate, hedgehog homologs can be engineered by the present method to provide more efficient binding to a cognate receptor, such as patched, yet still retain at least a portion of an activity associated with hedgehog. Thus, combinatorially-derived homologs can be generated to have an increased potency relative to a naturally occurring form of the protein. Likewise, hedgehog homologs can be generated by the present combinatorial approach to act as antagonists, in that they are able to mimic, for example, binding to other extracellular matrix components (such as receptors), yet not induce any biological response, thereby inhibiting the action of authentic hedgehog or hedgehog agonists. Moreover, manipulation of certain domains of hedgehog by the present method can provide domains more suitable for use in fusion proteins, such as one that incorporates portions of other proteins which are derived from the extracellular matrix and/or which bind extracellular matrix components.

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To further illustrate the state of the art of combinatorial mutagenesis, it is noted that the review article of Gallop et al. (1994) *J Med Chem* 37:1233 describes the general state of the art of combinatorial libraries as of the earlier 1990's. In particular, Gallop et al state at page 1239 "[s]creening the analog libraries aids in determining the minimum size of the active sequence and in identifying those residues critical for binding and intolerant of substitution". In addition, the Ladner et al. PCT publication WO90/02809, the Goeddel et al. U.S. Patent 5,223,408, and the Markland et al. PCT publication WO92/15679 illustrate specific techniques which one skilled in the art could utilize to generate libraries of *hedgehog* variants which can be rapidly screened to identify variants/fragments which retained a particular activity of the *hedgehog* polypeptides. These techniques are exemplary of the art and demonstrate that large libraries of related variants/truncants can be generated and assayed to isolate particular variants without undue experimentation. Gustin et al. (1993) *Virology* 193:653, and Bass et al. (1990) *Proteins: Structure, Function and Genetics* 8:309-314

also describe other exemplary techniques from the art which can be adapted as means for generating mutagenic variants of *hedgehog* polypeptides.

Indeed, it is plain from the combinatorial mutagenesis art that large scale mutagenesis of *hedgehog* proteins, without any preconceived ideas of which residues were critical to the biological function, and generate wide arrays of variants having equivalent biological activity. Indeed, it is the ability of combinatorial techniques to screen billions of different variants by high throughout analysis that removes any requirement of *a priori* understanding or knowledge of critical residues.

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To illustrate, the amino acid sequences for a population of hedgehog homologs or other related proteins are aligned, preferably to promote the highest homology possible. Such a population of variants can include, for example, hedgehog homologs from one or more species. Amino acids which appear at each position of the aligned sequences are selected to create a degenerate set of combinatorial sequences. In a preferred embodiment, the variegated library of hedgehog variants is generated by combinatorial mutagenesis at the nucleic acid level, and is encoded by a variegated gene library. For instance, a mixture of synthetic oligonucleotides can be enzymatically ligated into gene sequences such that the degenerate set of potential hedgehog sequences are expressible as individual polypeptides, or alternatively, as a set of larger fusion proteins (e.g. for phage display) containing the set of hedgehog sequences therein.

As illustrated in PCT publication WO 95/18856, to analyze the sequences of a population of variants, the amino acid sequences of interest can be aligned relative to sequence homology. The presence or absence of amino acids from an aligned sequence of a particular variant is relative to a chosen consensus length of a reference sequence, which can be real or artificial.

In an illustrative embodiment, alignment of exons 1, 2 and a portion of exon 3 encoded sequences (e.g. the N-terminal approximately 221 residues of the mature protein) of each of the *Shh* clones produces a degenerate set of *Shh* polypeptides represented by the general formula:

C-G-P-G-R-G-X(1)-G-X(2)-R-R-H-P-K-K-L-T-P-L-A-Y-K-Q-F-I-P-N-V-A-E-K-T-L-G-A-S-G-R-Y-E-G-K-I-X(3)-R-N-S-E-R-F-K-E-L-T-P-N-Y-N-P-D-I-I-F-K-D-E-E-N-T-G-A-D-R-L-M-T-Q-R-C-K-D-K-L-N-X(4)-L-A-I-S-V-M-N-X(5)-W-P-G-V-X(6)-I.-R-V-T-E-G-W-D-E-D-G-H-H-X(7)-E-E-S-L-H-Y-E-G-R-A-V-D-I-T-T-S-D-R-D-X(8)-S-K-Y-G-X(9)-L-X(10)-R-L-A-V-E-A-G-F-D-W-V-Y-Y-E-S-K-A-H-I-H-C-S-V-K-A-E-N-S-V-A-A-K-S-G-G-C-F-P-G-S-A-X(11)-V-X(12)-L-X(13)-X(14)-G-G-X(15)-K-X-(16)-V-K-D-L-X(17)-P-G-D-X(18)-V-L-A-A-D-X(19)-X(20)-G-X(21)-L-X(22)-X(23)-S-D-F-X(24)-X(25)-F-X(26)-D-R (SEQ ID No: 21

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wherein each of the degenerate positions "X" can be an amino acid which occurs in that position in one of the human, mouse, chicken or zebrafish Shh clones, or, to expand the library, each X can also be selected from amongst amino acid residue which would be conservative substitutions for the amino acids which appear naturally in each of those positions. For instance, Xaa(1) represents Gly, Ala, Val, Leu, Ile. Phe, Tyr or Trp; Xaa(2) represents Arg, His or Lys; Xaa(3) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(4) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(5) represents Lys, Arg, His, Asn or Gln; Xaa(6) represents Lys, Arg or His; Xaa(7) represents Ser, Thr, Tyr, Trp or Phe; Xaa(8) represents Lys, Arg or His; Xaa(9) represents Met, Cys, Ser or Thr; Xaa(10) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(11) represents Leu, Val, Met, Thr or Ser; Xaa(12) represents His, Phe, Tyr, Ser, Thr, Met or Cys; Xaa(13) represents Gln, Asn, Glu, or Asp; Xaa(14) represents His, Phe, Tyr, Thr, Gln, Asn, Glu or Asp; Xaa(15) represents Gln, Asn, Glu, Asp, Thr, Ser, Met or Cys; Xaa(16) represents Ala, Gly, Cys, Leu, Val or Met; Xaa(17) represents Arg, Lys, Met, Ile, Asn, Asp, Glu, Gln, Ser, Thr or Cys; Xaa(18) represents Arg, Lys, Met or Ile; Xaa(19) represents Ala, Gly, Cys, Asp, Glu, Gln, Asn, Ser, Thr or Met; Xaa(20) represents Ala, Gly, Cys, Asp, Asn, Glu or Gln; Xaa(21) represents Arg, Lys, Met, Ile, Asn, Asp, Glu or Gln; Xaa(22) represent Leu, Val, Met or Ile; Xaa(23) represents Phe, Tyr, Thr, His or Trp; Xaa(24) represents Ile, Val, Leu or Met; .Xaa(25) represents Met, Cys, Ile, Leu, Val, Thr or Ser; Xaa(26) represents Leu, Val, Met, Thr or Ser. In an even more expansive library, each X can be selected from any amino acid.

In similar fashion, alignment of each of the human, mouse, chicken and zebrafish *hedgehog* clones, can provide a degenerate polypeptide sequence represented by the general formula:

C-G-P-G-R-G-X(1)-X(2)-X(3)-R-R-X(4)-X(5)-X(6)-P-K-X(7)-L-X(8)-P-L-X(9)-Y-K-Q-F-X(10)-P-X(11)-X(12)-X(13)-E-X(14)-T-L-G-A-S-G-X(15)-X(16)-E-G-X(17)-X(18)-X(19)-R-X(20)-S-E-R-F-X(21)-X(22)-L-T-P-N-Y-N-P-D-I-I-F-K-D-E-E-N-X(23)-G-A-D-R-L-M-T-X(24)-R-C-K-X(25)-X(26)-X(27)-N-X(28)-L-A-I-S-V-M-N-X(29)-W-P-G-V-X(30)-L-R-V-T-E-G-X(31)-D-E-D-G-H-H-X(32)-X(33)-X(34)-S-L-H-Y-E-G-R-A-X(35)-D-I-T-T-S-D-R-D-X(36)-X(37)-K-Y-G-X(38)-L-X(39)-R-L-A-V-E-A-G-F-D-W-V-Y-Y-E-S-X(40)-X(41)-H-X(42)-H-X(43)-S-V-K-X(44)-X(45) (SEQ IDNo:22

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wherein, as above, each of the degenerate positions "X" can be an amino acid which occurs in a corresponding position in one of the wild-type clones, and may also include amino acid residue which would be conservative substitutions, or each X can be any amino acid residue. In an exemplary embodiment, Xaa(1) represents Gly, Ala, Val, Leu, Ile, Pro, Phe or Tyr; Xaa(2) represents Gly, Ala, Val, Leu or Ile; Xaa(3) represents Gly, Ala, Val, Leu, Ile, Lys, His or Arg; Xaa(4) represents Lys, Arg or His; Xaa(5) represents Phe. Trp, Tyr or an amino acid gap; Xaa(6) represents Gly, Ala, Val, Leu, Ile or an amino acid gap; Xaa(7) represents Asn. Gln, His, Arg or Lys; Xaa(8) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(9) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(10) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(11) represents Ser, Thr, Gln or Asn; Xaa(12) represents Met, Cys, Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(13) represents Gly, Ala, Val, Leu, Ile or Pro; Xaa(14) represents Arg, His or Lys; Xaa(15) represents Gly, Ala, Val, Leu, Ile, Pro, Arg, His or Lys; Xaa(16) represents Gly, Ala, Val, Leu, Ile, Phe or Tyr; Xaa(17) represents Arg, His or Lys; Xaa(18) represents Gly, Ala, Val, Leu, Ile, Ser or Thr; Xaa(19) represents Thr or Ser; Xaa(20) represents Gly, Ala, Val, Leu, Ile, Asn or Gln; Xaa(21) represents Arg, His or Lys; Xaa(22) represents Asp or Glu; Xaa(23) represents Ser or Thr; Xaa(24) represents Glu, Asp, Gln or Asn; Xaa(25) represents Glu or Asp; Xaa(26) represents Arg, His or Lys; Xaa(27) represents Gly, Ala, Val, Leu or Ile; Xaa(28) represents Gly, Ala, Val, Leu, Ile, Thr or Ser; Xaa(29) represents Met, Cys, Gln, Asn, Arg, Lys or His; Xaa(30) represents Arg, His or Lys; Xaa(31)

represents Trp, Phe, Tyr, Arg, His or Lys; Xaa(32) represents Gly, Ala, Val, Leu, Ile, Ser, Thr, Tyr or Phe; Xaa(33) represents Gln, Asn, Asp or Glu; Xaa(34) represents Asp or Glu; Xaa(35) represents Gly, Ala, Val, Leu, or Ile; Xaa(36) represents Arg, His or Lys; Xaa(37) represents Asn, Gln, Thr or Ser; Xaa(38) represents Gly, Ala, Val, Leu, Ile, Ser, Thr, Met or Cys; Xaa(39) represents Gly, Ala, Val, Leu, Ile, Thr or Ser; Xaa(40) represents Arg, His or Lys; Xaa(41) represents Asn, Gln, Gly, Ala, Val, Leu or Ile; Xaa(42) represents Gly, Ala, Val, Leu or Ile; Xaa(43) represents Gly, Ala, Val, Leu, Ile, Ser, Thr or Cys; Xaa(44) represents Gly, Ala, Val, Leu, Ile, Thr or Ser; and Xaa(45) represents Asp or Glu.

There are many ways by which the library of potential hedgehog homologs can be generated from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be carried out in an automatic DNA synthesizer, and the synthetic genes then ligated into an appropriate expression vector. The purpose of a degenerate set of genes is to provide, in one mixture, all of the sequences encoding the desired set of potential hedgehog sequences. The synthesis of degenerate oligonucleotides is well known in the art (see for example, Narang, SA (1983) Tetrahedron 39:3; Itakura et al. (1981) Recombinant DNA, Proc 3rd Cleveland Sympos. Macromolecules, ed. AG Walton, Amsterdam: Elsevier pp273-289; Itakura et al. (1984) Annu. Rev. Biochem. 53:323; Itakura et al. (1984) Science 198:1056; Ike et al. (1983) Nucleic Acid Res. 11:477. Such techniques have been employed in the directed evolution of other proteins (see, for example, Scott et al. (1990) Science 249:386-390; Roberts et al. (1992) PNAS 89:2429-2433; Devlin et al. (1990) Science 249: 404-406; Cwirla et al. (1990) PNAS 87: 6378-6382; as well as U.S. Patents Nos. 5,223,409, 5,198,346, and 5,096,815).

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A wide range of techniques are known in the art for screening gene products of combinatorial libraries made by point mutations, and for screening cDNA libraries for gene products having a certain property. Such techniques will be generally adaptable for rapid screening of the gene libraries generated by the combinatorial mutagenesis of *hedgehog* homologs. The most widely used techniques for screening large gene libraries typically comprises cloning the gene library into replicable expression vectors, transforming appropriate cells with the resulting library of vectors, and expressing the combinatorial genes under conditions in which detection of a desired activity facilitates relatively easy isolation of

the vector encoding the gene whose product was detected. Each of the illustrative assays described below are amenable to high through-put analysis as necessary to screen large numbers of degenerate *hedgehog* sequences created by combinatorial mutagenesis techniques.

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In one embodiment, the combinatorial library is designed to be secreted (e.g. the polypeptides of the library all include a signal sequence but no transmembrane or cytoplasmic domains), and is used to transfect a eukaryotic cell that can be co-cultured with lung cells, e.g., lung mesenchymal or epithelial cells. A functional *hedgehog* protein secreted by the cells expressing the combinatorial library will diffuse to the neighboring lung cells and induce a particular biological response, such as proliferation. The pattern of detection of proliferation will resemble a gradient function, and will allow the isolation (generally after several repetitive rounds of selection) of cells producing *hedgehog* homologs active as proliferative agents with respect to the lung cells. Likewise, *hedgehog* antagonists can be selected in similar fashion by the ability of the cell producing a functional antagonist to protect neighboring cells (e.g., to inhibit proliferation) from the effect of wild-type *hedgehog* added to the culture media.

To illustrate, target lung cells are cultured in 24-well microtitre plates. Other eukaryotic cells are transfected with the combinatorial hedgehog gene library and cultured in cell culture inserts (e.g. Collaborative Biomedical Products, Catalog #40446) that are able to fit into the wells of the microtitre plate. The cell culture inserts are placed in the wells such that recombinant hedgehog homologs secreted by the cells in the insert can diffuse through the porous bottom of the insert and contact the target cells in the microtitre plate wells. After a period of time sufficient for functional forms of a hedgehog protein to produce a measurable response in the target cells, such as proliferation, the inserts are removed and the effect of the variant hedgehog proteins on the target cells determined. Cells from the inserts corresponding to wells which score positive for activity can be split and re-cultured on several inserts, the process being repeated until the active clones are identified.

In yet another screening assay, the candidate *hedgehog* gene products are displayed on the surface of a cell or viral particle, and the ability of particular cells or viral particles to associate with a *hedgehog*-binding moiety (such as the *patched* protein or other *hedgehog* 

receptor) via this gene product is detected in a "panning assay". Such panning steps can be carried out on cells cultured from embryos. For instance, the gene library can be cloned into the gene for a surface membrane protein of a bacterial cell, and the resulting fusion protein detected by panning (Ladner et al., WO 88/06630; Fuchs et al. (1991) Bio/Technology 9:1370-1371; and Goward et al. (1992) TIBS 18:136-140). In a similar fashion, fluorescently labeled molecules which bind hedgehog can be used to score for potentially functional hedgehog homologs. Cells can be visually inspected and separated under a fluorescence microscope, or, where the morphology of the cell permits, separated by a fluorescence-activated cell sorter.

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In an alternate embodiment, the gene library is expressed as a fusion protein on the surface of a viral particle. For instance, in the filamentous phage system, foreign peptide sequences can be expressed on the surface of infectious phage, thereby conferring two significant benefits. First, since these phage can be applied to affinity matrices at very high concentrations, large number of phage can be screened at one time. Second, since each infectious phage displays the combinatorial gene product on its surface, if a particular phage is recovered from an affinity matrix in low yield, the phage can be amplified by another round of infection. The group of almost identical *E.coli* filamentous phages M13, fd, and fl are most often used in phage display libraries, as either of the phage gIII or gVIII coat proteins can be used to generate fusion proteins without disrupting the ultimate packaging of the viral particle (Ladner et al. PCT publication WO 90/02909; Garrard et al., PCT publication WO 92/09690; Marks et al. (1992) J. Biol. Chem. 267:16007-16010; Griffths et al. (1993) EMBO J 12:725-734; Clackson et al. (1991) Nature 352:624-628; and Barbas et al. (1992) PNAS 89:4457-4461).

In an illustrative embodiment, the recombinant phage antibody system (RPAS, Pharamacia Catalog number 27-9400-01) can be easily modified for use in expressing and screening *hedgehog* combinatorial libraries. For instance, the pCANTAB 5 phagemid of the RPAS kit contains the gene which encodes the phage gIII coat protein. The *hedgehog* combinatorial gene library can be cloned into the phagemid adjacent to the gIII signal sequence such that it will be expressed as a gIII fusion protein. After ligation, the phagemid is used to transform competent *E. coli* TG1 cells. Transformed cells are subsequently

infected with M13KO7 helper phage to rescue the phagemid and its candidate *hedgehog* gene insert. The resulting recombinant phage contain phagemid DNA encoding a specific candidate *hedgehog*, and display one or more copies of the corresponding fusion coat protein. The phage-displayed candidate *hedgehog* proteins which are capable of binding an *hedgehog* receptor are selected or enriched by panning. For instance, the phage library can be applied to cells which express the *patched* protein and unbound phage washed away from the cells. The bound phage is then isolated, and if the recombinant phage express at least one copy of the wild type gIII coat protein, they will retain their ability to infect *E. coli*. Thus, successive rounds of reinfection of *E. coli*, and panning will greatly enrich for *hedgehog* homologs, which can then be screened for further biological activities in order to differentiate agonists and antagonists.

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Combinatorial mutagenesis has a potential to generate very large libraries of mutant proteins, e.g., in the order of 10<sup>26</sup> molecules. Combinatorial libraries of this size may be technically challenging to screen even with high throughput screening assays such as phage display. To overcome this problem, a new technique has been developed recently, recursive ensemble mutagenesis (REM), which allows one to avoid the very high proportion of nonfunctional proteins in a random library and simply enhances the frequency of functional proteins, thus decreasing the complexity required to achieve a useful sampling of sequence space. REM is an algorithm which enhances the frequency of functional mutants in a library when an appropriate selection or screening method is employed (Arkin and Yourvan, 1992, *PNAS USA* 89:7811-7815; Yourvan et al., 1992, *Parallel Problem Solving from Nature*, 2., In Maenner and Manderick, eds., Elsevir Publishing Co., Amsterdam, pp. 401-410; Delgrave et al., 1993, *Protein Engineering* 6(3):327-331).

The invention also provides for reduction of the *hedgehog* protein to generate mimetics, e.g. peptide or non-peptide agents, which are able to disrupt binding of a *hedgehog* polypeptide of the present invention with an *hedgehog* receptor. Thus, such mutagenic techniques as described above are also useful to map the determinants of the *hedgehog* proteins which participate in protein-protein interactions involved in, for example, binding of the subject *hedgehog* polypeptide to other extracellular matrix components. To illustrate, the critical residues of a subject *hedgehog* polypeptide which are involved in molecular

recognition of an hedgehog receptor such as patched can be determined and used to generate hedgehog-derived peptidomimetics which competitively inhibit binding of the authentic hedgehog protein with that moiety. By employing, for example, scanning mutagenesis to map the amino acid residues of each of the subject hedgehog proteins which are involved in binding other extracellular proteins, peptidomimetic compounds can be generated which mimic those residues of the hedgehog protein which facilitate the interaction. Such mimetics may then be used to interfere with the normal function of a hedgehog protein. For instance, non-hydrolyzable peptide analogs of such residues can be generated using benzodiazepine (e.g., see Freidinger et al. in Peptides: Chemistry and Biology, G.R. Marshall ed., ESCOM Publisher: Leiden, Netherlands, 1988), azepine (e.g., see Huffman et al. in Peptides: Chemistry and Biology, G.R. Marshall ed., ESCOM Publisher: Leiden, Netherlands, 1988), substituted gama lactam rings (Garvey et al. in Peptides: Chemistry and Biology, G.R. Marshall ed., ESCOM Publisher: Leiden, Netherlands, 1988), keto-methylene pseudopeptides (Ewenson et al. (1986) J Med Chem 29:295; and Ewenson et al. in Peptides: Structure and Function (Proceedings of the 9th American Peptide Symposium) Pierce Chemical Co. Rockland, IL, 1985), β-turn dipeptide cores (Nagai et al. (1985) Tetrahedron Lett 26:647; and Sato et al. (1986) J Chem Soc Perkin Trans 1:1231), and β-aminoalcohols (Gordon et al. (1985) Biochem Biophys Res Commun126:419; and Dann et al. (1986) Biochem Biophys Res Commun 134:71).

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Recombinantly produced forms of the *hedgehog* proteins can be produced using, e.g, expression vectors containing a nucleic acid encoding a *hedgehog* polypeptide, operably linked to at least one transcriptional regulatory sequence. Operably linked is intended to mean that the nucleotide sequence is linked to a regulatory sequence in a manner which allows expression of the nucleotide sequence. Regulatory sequences are art-recognized and are selected to direct expression of a *hedgehog* polypeptide. Accordingly, the term transcriptional regulatory sequence includes promoters, enhancers and other expression control elements. Such regulatory sequences are described in Goeddel; *Gene Expression Technology: Methods in Enzymology* 185, Academic Press, San Diego, CA (1990). For instance, any of a wide variety of expression control sequences, sequences that control the expression of a DNA sequence when operatively linked to it, may be used in these vectors to express DNA sequences encoding *hedgehog* polypeptide. Such useful expression control

sequences, include, for example, a viral LTR, such as the LTR of the Moloney murine leukemia virus, the early and late promoters of SV40, adenovirus or cytomegalovirus immediate early promoter, the lac system, the trp system, the TAC or TRC system, T7 promoter whose expression is directed by T7 RNA polymerase, the major operator and promoter regions of phage  $\lambda$ , the control regions for fd coat protein, the promoter for-3-phosphoglycerate kinase or other glycolytic enzymes, the promoters of acid phosphatase, e.g., Pho5, the promoters of the yeast  $\alpha$ -mating factors, the polyhedron promoter of the baculovirus system and other sequences known to control the expression of genes of prokaryotic or eukaryotic cells or their viruses, and various combinations thereof. It should be understood that the design of the expression vector may depend on such factors as the choice of the host cell to be transformed and/or the type of protein desired to be expressed. Moreover, the vector's copy number, the ability to control that copy number and the expression of any other proteins encoded by the vector, such as antibiotic markers, should also be considered.

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In addition to providing a ready source of *hedgehog* polypeptides for purification, the gene constructs of the present invention can also be used as a part of a gene therapy protocol to deliver nucleic acids encoding either an agonistic or antagonistic form of a *hedgehog* polypeptide. Thus, another aspect of the invention features expression vectors for *in vivo* transfection of a *hedgehog* polypeptide in particular cell types so as cause ectopic expression of a *hedgehog* polypeptide in lung tissue.

Formulations of such expression constructs may be administered in any biologically effective carrier, e.g. any formulation or composition capable of effectively delivering the recombinant gene to cells *in vivo*. Approaches include insertion of the *hedgehog* coding sequence in viral vectors including recombinant retroviruses, adenovirus, adeno-associated virus, and herpes simplex virus-1, or recombinant bacterial or eukaryotic plasmids. Viral vectors transfect cells directly; plasmid DNA can be delivered with the help of, for example, cationic liposomes (lipofectin) or derivatized (e.g. antibody conjugated), polylysine conjugates, gramacidin S, artificial viral envelopes or other such intracellular carriers, as well as direct injection of the gene construct or CaPO<sub>4</sub> precipitation carried out *in vivo*. It will be appreciated that because transduction of appropriate target cells represents the critical first

step in gene therapy, choice of the particular gene delivery system will depend on such factors as the phenotype of the intended target and the route of administration, e.g. locally or systemically. Furthermore, it will be recognized that the particular gene construct provided for *in vivo* transduction of *hedgehog* expression are also useful for *in vitro* transduction of cells, such as for use in the *ex vivo* tissue culture systems described below.

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A preferred approach for *in vivo* introduction of nucleic acid into a cell is by use of a viral vector containing nucleic acid, e.g. a cDNA, encoding the particular form of the *hedgehog* polypeptide desired. Infection of cells with a viral vector has the advantage that a large proportion of the targeted cells can receive the nucleic acid. Additionally, molecules encoded within the viral vector, e.g., by a cDNA contained in the viral vector, are expressed efficiently in cells which have taken up viral vector nucleic acid.

Retrovirus vectors and adeno-associated virus vectors are generally understood to be the recombinant gene delivery system of choice for the transfer of exogenous genes in vivo, particularly into humans. These vectors provide efficient delivery of genes into cells, and the transferred nucleic acids are stably integrated into the chromosomal DNA of the host. A major prerequisite for the use of retroviruses is to ensure the safety of their use, particularly with regard to the possibility of the spread of wild-type virus in the cell population. The development of specialized cell lines (termed "packaging cells") which produce only replication-defective retroviruses has increased the utility of retroviruses for gene therapy, and defective retroviruses are well characterized for use in gene transfer for gene therapy purposes (for a review see Miller, A.D. (1990) Blood 76:271). Thus, recombinant retrovirus can be constructed in which part of the retroviral coding sequence (gag, pol, env) has been replaced by nucleic acid encoding a hedgehog polypeptide and renders the retrovirus replication defective. The replication defective retrovirus is then packaged into virions which can be used to infect a target cell through the use of a helper virus by standard techniques. Protocols for producing recombinant retroviruses and for infecting cells in vitro or in vivo with such viruses can be found in Current Protocols in Molecular Biology, Ausubel, F.M. et al. (eds.) Greene Publishing Associates, (1989), Sections 9.10-9.14 and other standard laboratory manuals. Examples of suitable retroviruses include pLJ, pZIP, pWE and pEM which are well known to those skilled in the art. Examples of suitable packaging virus lines

for preparing both ecotropic and amphotropic retroviral systems include Crip, Cre, 2 and Am. Retroviruses have been used to introduce a variety of genes into many different cell types, including lung cells, in vitro and/or in vivo (see for example Eglitis, et al. (1985) Science 230:1395-1398; Danos and Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:6460-6464; Wilson et al. (1988) Proc. Natl. Acad. Sci. USA 85:3014-3018; Armentano et al. (1990) Proc. Natl. Acad. Sci. USA 87:6141-6145; Huber et al. (1991) Proc. Natl. Acad. Sci. USA 88:8039-8043; Ferry et al. (1991) Proc. Natl. Acad. Sci. USA 88:8377-8381; Chowdhury et al. (1991) Science 254:1802-1805; van Beusechem et al. (1992) Proc. Natl. Acad. Sci. USA 89:7640-7644; Kay et al. (1992) Human Gene Therapy 3:641-647; Dai et al. (1992) Proc. Natl. Acad. Sci. USA 89:10892-10895; Hwu et al. (1993) J. Immunol. 150:4104-4115; U.S. Patent No. 4,868,116; U.S. Patent No. 4,980,286; PCT Application WO 89/07136; PCT Application WO 89/07573).

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Furthermore, it has been shown that it is possible to limit the infection spectrum of retroviruses and consequently of retroviral-based vectors, by modifying the viral packaging proteins on the surface of the viral particle (see, for example PCT publications WO93/25234 and WO94/06920). For instance, strategies for the modification of the infection spectrum of retroviral vectors include: coupling antibodies specific for cell surface antigens to the viral env protein (Roux et al. (1989) PNAS 86:9079-9083; Julan et al. (1992) J. Gen Virol 73:3251-3255; and Goud et al. (1983) Virology 163:251-254); or coupling cell surface receptor ligands to the viral env proteins (Neda et al. (1991) J Biol Chem 266:14143-14146). Coupling can be in the form of the chemical cross-linking with a protein or other variety (e.g. lactose to convert the env protein to an asialoglycoprotein), as well as by generating fusion proteins (e.g. single-chain antibody/env fusion proteins). This technique, while useful to limit or otherwise direct the infection to certain tissue types, can also be used to convert an ecotropic vector in to an amphotropic vector.

Moreover, use of retroviral gene delivery can be further enhanced by the use of tissueor cell-specific transcriptional regulatory sequences which control expression of the hedgehog gene of the retroviral vector.

Another viral gene delivery system useful in the present method utilizes adenovirusderived vectors. The genome of an adenovirus can be manipulated such that it encodes and

expresses a gene product of interest but is inactivated in terms of its ability to replicate in a normal lytic viral life cycle. See for example Berkner et al. (1988) BioTechniques 6:616; Rosenfeld et al. (1991) Science 252:431-434; and Rosenfeld et al. (1992) Cell 68:143-155. Suitable adenoviral vectors derived from the adenovirus strain Ad type 5 dl324 or other strains of adenovirus (e.g., Ad2, Ad3, Ad7 etc.) are well known to those skilled in the-art. Recombinant adenoviruses can be advantageous in certain circumstances in that they can be used to infect a wide variety of cell types, including lung cells (Rosenfeld et al. (1992) cited supra). Furthermore, the virus particle is relatively stable and amenable to purification and concentration, and as above, can be modified so as to affect the spectrum of infectivity. Additionally, introduced adenoviral DNA (and foreign DNA contained therein) is not integrated into the genome of a host cell but remains episomal, thereby avoiding potential problems that can occur as a result of insertional mutagenesis in situations where introduced DNA becomes integrated into the host genome (e.g., retroviral DNA). Moreover, the carrying capacity of the adenoviral genome for foreign DNA is large (up to 8 kilobases) relative to other gene delivery vectors (Berkner et al. cited supra; Haj-Ahmand and Graham (1986) J. Virol. 57:267). Most replication-defective adenoviral vectors currently in use and therefore favored by the present invention are deleted for all or parts of the viral E1 and E3 genes but retain as much as 80% of the adenoviral genetic material (see, e.g., Jones et al. (1979) Cell 16:683; Berkner et al., supra; and Graham et al. in Methods in Molecular Biology, E.J. Murray, Ed. (Humana, Clifton, NJ, 1991) vol. 7. pp. 109-127). Expression of the inserted hedgehog gene can be under control of, for example, the E1A promoter, the major late promoter (MLP) and associated leader sequences, the E3 promoter, or exogenously added promoter sequences.

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In addition to viral transfer methods, such as those illustrated above, non-viral methods can also be employed to cause expression of a *hedgehog* polypeptide in the tissue of an animal. Most nonviral methods of gene transfer rely on normal mechanisms used by mammalian cells for the uptake and intracellular transport of macromolecules. In preferred embodiments, non-viral gene delivery systems of the present invention rely on endocytic pathways for the uptake of the *hedgehog* polypeptide gene by the targeted cell. Exemplary gene delivery systems of this type include liposomal derived systems, poly-lysine conjugates, and artificial viral envelopes.

In clinical settings, the gene delivery systems for the therapeutic *hedgehog* gene can be introduced into a patient by any of a number of methods, each of which is familiar in the art. For instance, a pharmaceutical preparation of the gene delivery system can be introduced systemically, e.g. by intravenous injection, and specific transduction of the protein in the target cells occurs predominantly from specificity of transfection provided by the gene delivery vehicle, cell-type or tissue-type expression due to the transcriptional regulatory sequences controlling expression of the receptor gene, or a combination thereof. In other embodiments, initial delivery of the recombinant gene is more limited with introduction into the animal being quite localized. For example, the gene delivery vehicle can be introduced by catheter (see U.S. Patent 5,328,470) or by stereotactic injection (e.g. Chen et al. (1994) *PNAS* 91: 3054-3057). A *hedgehog* expression construct can be delivered in a gene therapy construct to dermal cells by, e.g., electroporation using techniques described, for example, by Dev et al. ((1994) *Cancer Treat Rev* 20:105-115).

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The pharmaceutical preparation of the gene therapy construct can consist essentially of the gene delivery system in an acceptable diluent, or can comprise a slow release matrix in which the gene delivery vehicle is imbedded. Alternatively, where the complete gene delivery system can be produced intact from recombinant cells, e.g. retroviral vectors, the pharmaceutical preparation can comprise one or more cells which produce the gene delivery system.

In yet another embodiment, the ptc, hedgehog or fgf-10 therapeutic can be a "gene activation" construct which, by homologous recombination with a genomic DNA, alters the transcriptional regulatory sequences of an endogenous gene. For instance, the gene activation construct can replace the endogenous promoter of a hedgehog gene with a heterologous promoter, e.g., one which causes consitutive expression of the hedgehog gene or which causes inducible expression of the gene under conditions different from the normal expression pattern of the gene. Other genes in the patched signaling pathway can be similarly targeted. A vareity of different formats for the gene activation constructs are available. See, for example, the Transkaryotic Therapies, Inc PCT publications WO93/09222, WO95/31560, WO96/29411, WO95/31560 and WO94/12650.

In preferred embodiments, the nucleotide sequence used as the gene activation construct can be comprised of (1) DNA from some portion of the endogenous *hedgehog* gene (exon sequence, intron sequence, promoter sequences, etc.) which direct recombination and (2) heterologous transcriptional regulatory sequence(s) which is to be operably linked to the coding sequence for the genomic *hedgehog* gene upon recombination of the gene activation construct. For use in generating cultures of *hedgehog* producing cells, the construct may further include a reporter gene to detect the presence of the knockout construct in the cell.

The gene activation construct is inserted into a cell, and integrates with the genomic DNA of the cell in such a position so as to provide the heterologous regulatory sequences in operative association with the native *hedgehog* gene. Such insertion occurs by homologous recombination, i.e., recombination regions of the activation construct that are homologous to the endogenous *hedgehog* gene sequence hybridize to the genomic DNA and recombine with the genomic sequences so that the construct is incorporated into the corresponding position of the genomic DNA.

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The terms "recombination region" or "targeting sequence" refer to a segment (i.e., a portion) of a gene activation construct having a sequence that is substantially identical to or substantially complementary to a genomic gene sequence, e.g., including 5' flanking sequences of the genomic gene, and can facilitate homologous recombination between the genomic sequence and the targeting transgene construct.

As used herein, the term "replacement region" refers to a portion of a activation construct which becomes integrated into an endogenous chromosomal location following homologous recombination between a recombination region and a genomic sequence.

The heterologous regulatory sequences, e.g., which are provided in the replacement region, can include one or more of a variety elements, including: promoters (such as constitutive or inducible promoters), enhancers, negative regulatory elements, locus control regions, transcription factor binding sites, or combinations thereof. Promoters/enhancers which may be used to control the expression of the targeted gene *in vivo* include, but are not limited to, the cytomegalovirus (CMV) promoter/enhancer (Karasuyama et al., 1989, *J. Exp. Med.*, 169:13), the human β-actin promoter (Gunning et al. (1987) *PNAS* 84:4831-4835), the glucocorticoid-inducible promoter present in the mouse mammary tumor virus long terminal

repeat (MMTV LTR) (Klessig et al. (1984) *Mol. Cell Biol.* 4:1354-1362), the long terminal repeat sequences of Moloney murine leukemia virus (MuLV LTR) (Weiss et al. (1985) *RNA Tumor Viruses*, Cold Spring Harbor Laboratory, Cold Spring Harbor, New York), the SV40 early or late region promoter (Bernoist et al. (1981) *Nature* 290:304-310; Templeton et al. (1984) *Mol. Cell Biol.*, 4:817; and Sprague et al. (1983) *J. Virol.*, 45:773), the promoter contained in the 3' long terminal repeat of Rous sarcoma virus (RSV) (Yamamoto et al., 1980, *Cell*, 22:787-797), the herpes simplex virus (HSV) thymidine kinase promoter/enhancer (Wagner et al. (1981) *PNAS* 82:3567-71), and the herpes simplex virus LAT promoter (Wolfe et al. (1992) *Nature Genetics*, 1:379-384).

In an exemplary embodiment, portions of the 5' flanking region of the human Shh gene are amplified using primers which add restriction sites, to generate the following fragments

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As illustrated, primer 1 includes a 5' non-coding region of the human Shh gene and is flanked by an AsuII and ClaI restriction sites. Primer 2 includes a portion of the 5' non-coding region immediately 3' to that present in primer 1. The *hedgehog* gene sequence is flanked by XhoII and BamHI restriction sites. The purified amplimers are cut with each of the enzymes as appropriate.

The vector pCDNA1.1 (Invitrogen) includes a CMV promoter. The plasmid is cut with with AsuII, which cleaves just 3' to the CMV promoter sequence. The AsuII/ClaI fragment of primer 1 is ligated to the AsuII cleavage site of the pcDNA vector. The ClaI/AsuII ligation destroys the AsuII site at the 3' end of a properly inserted primer 1.

The vector is then cut with BamHI, and an XhoII/BamHI fragment of primer 2 is ligated to the BamHI cleavage site. As above, the BamHI/XhoII ligation destroys the BamHI site at the 5' end of a properly inserted primer 2.

Individual colonies are selected, cut with AsuII and BamHI, and the size of the AsuII/BamHI fragment determined. Colonies in which both the primer 1 and primer 2 sequences are correctly inserted are further amplified, an cut with AsuII and BamHI to produce the gene activation construct

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cqaagcqaggcagccagcgagggagagagcgagcggggcgagcggagcgaggaaATCGAAGG TTCGAATCCTTCCCCCACCACCATCACTTTCAAAAGTCCGAAAGAATCTGCTCCCTGCTTGT GTGTTGGAGGTCGCTGAGTAGTGCGCGAGTAAAATTTAAGCTACAACAAGGCAAGGCTTGAC CGACAATTGCATGAAGAATCTGCTTAGGGTTAGGCGTTTTGCGCTGCTTCGCGATGTACGGG CCAGATATACGCGTTGACATTGATTATTGACTAGTTATTAATAGTAATCAATTACGGGGTCA TTAGTTCATAGCCCATATATGGAGTTCCGCGTTACATAACTTACGGTAAATGGCCCGCCTGG CTGACCGCCCAACGACCCCCGCCCATTGACGTCAATAATGACGTATGTTCCCATAGTAACGC CAATAGGGACTTTCCATTGACGTCAATGGGTGGACTATTTACGGTAAACTGCCCACTTGGCA GTACATCAAGTGTATCATATGCCAAGTACGCCCCCTATTGACGTCAATGACGGTAAATGGCC CGCCTGGCATTATGCCCAGTACATGACCTTATGGGACTTTCCTACTTGGCAGTACATCTACG TATTAGTCATCGCTATTACCATGGTGATGCGGTTTTTGGCAGTACATCAATGGGCGTGGATAG GCACCAAAATCAACGGGACTTTCCAAAATGTCGTAACAACTCCGCCCCATTGACGCAAATGG GCGGTAGGCGTGTACGGTGGGAGGTCTATATAAGCAGAGCTCTCTGGCTAACTAGAGAACCC ACTGCTTACTGGCTTATCGAAATTAATACGACTCACTATAGGGAGACCCAAGCTTGGTACCG AGCTCGGATCgatctgggaaagcgcaagagagcgcacacgcacaccccgccgcgcgcac tcgg

In this construct, the flanking primer 1 and primer 2 sequences provide the recombination region which permits the insertion of the CMV promoter in front of the coding sequence for the human *Shh* gene. Other heterologous promoters (or other transcriptional regulatory sequences) can be inserted in a genomic *hedgehog* gene by a similar method.

In still other embodiments, the replacement region merely deletes a negative transcriptional control element of the native gene, e.g., to activate expression, or ablates a positive control element, e.g., to inhibit expression of the targeted gene.

## V. Exemplary ptc therapeutic compounds.

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In another embodiment, the subject method is carried out using a ptc therapeutic composition. Such compositions can be generated with, for example, compounds which bind to patched and alter its signal transduction activity, compounds which alter the binding and/or enzymatic activity of a protein (e.g., intracellular) involved in patched signal pathway, and compounds which alter the level of expression of a hedgehog protein, a patched protein or a protein involved in the intracellular signal transduction pathway of patched.

The availability of purified and recombinant hedgehog polypeptides facilitates the generation of assay systems which can be used to screen for drugs, such as small organic molecules, which are either agonists or antagonists of the normal cellular function of a hedgehog and/or patched protein, particularly their role in the pathogenesis of proliferation and/or differentiation of various lung cells and maintenance of lung tissue. In one embodiment, the assay evaluates the ability of a compound to modulate binding between a hedgehog polypeptide and a hedgehog receptor such as patched. In other embodiments, the assay merely scores for the ability of a test compound to alter the signal transduction actitity of the patched protein. In this manner, a variety of hedgehog and/or ptc therapeutics, both proliferative and anti-proliferative in activity, can be identified. A variety of assay formats will suffice and, in light of the present disclosure, will be comprehended by skilled artisan.

In many drug screening programs which test libraries of compounds and natural extracts, high throughput assays are desirable in order to maximize the number of compounds surveyed in a given period of time. Assays which are performed in cell-free systems, such as may be derived with purified or semi-purified proteins, are often preferred as "primary" screens in that they can be generated to permit rapid development and relatively easy detection of an alteration in a molecular target which is mediated by a test compound. Moreover, the effects of cellular toxicity and/or bioavailability of the test compound can be generally ignored in the *in vitro* system, the assay instead being focused primarily on the

effect of the drug on the molecular target as may be manifest in an alteration of binding affinity with receptor proteins.

Acordingly, in an exemplary screening assay for ptc therapeutics, the compound of interest is contacted with a mixture including a hedgehog receptor protein (e.g., a cell expressing the patched receptor) and a hedgehog protein under conditions in which it is ordinarily capable of binding the hedgehog protein. To the mixture is then added a composition containing a test compound. Detection and quantification of receptor/hedgehog complexes provides a means for determining the test compound's efficacy at inhibiting (or potentiating) complex formation between the receptor protein and the hedgehog polypeptide. The efficacy of the compound can be assessed by generating dose response curves from data obtained using various concentrations of the test compound. Moreover, a control assay can also be performed to provide a baseline for comparison. In the control assay, isolated and purified hedgehog polypeptide is added to the receptor protein, and the formation of receptor/hedgehog complex is quantitated in the absence of the test compound.

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In other embodiments, a ptc therapeutic of the present invention is one which disrupts the association of patched with smoothened.

Agonist and antagonists of cell growth can be distinguished, and the efficacy of the compound can be assessed, by subsequent testing with certain lung cells, e.g., in culture.

In an illustrative embodiment, the polypeptide utilized as a hedgehog receptor can be generated from the patched protein. Accordingly, an exemplary screening assay includes all or a suitable portion of the patched protein which can be obtained from, for example, the human patched gene (GenBank U43148) or other vertebrate sources (see GenBank Accession numbers U40074 for chicken patched and U46155 for mouse patched), as well as from drosophila (GenBank Accession number M28999) or other invertebrate sources. The patched protein can be provided in the screening assay as a whole protein (preferably expressed on the surface of a cell), or alternatively as a fragment of the full length protein which binds to hedgehog polypeptides, e.g., as one or both of the substantial extracellular domains (e.g. corresponding to residues Asn120-Ser438 and/or Arg770-Trp1027 of the human patched protein - which are also potential antagonists of hedgehog-dependent signal transduction). For instance, the patched protein can be provided in soluble form, as for

example a preparation of one of the extracellular domains, or a preparation of both of the extracellular domains which are covalently connected by an unstructured linker (see, for example, Huston et al. (1988) PNAS 85:4879; and U.S. Patent No. 5,091,513). In other embodiments, the protein can be provided as part of a liposomal preparation or expressed on the surface of a cell. The *patched* protein can derived from a recombinant gene, e.g., being ectopically expressed in a heterologous cell. For instance, the protein can be expressed on oocytes, mammalian cells (e.g., COS, CHO, 3T3 or the like), or yeast cell by standard recombinant DNA techniques. These recombinant cells can be used for receptor binding, signal transduction or gene expression assays. Marigo et al. (1996) *Development* 122:1225-1233 illustrates a binding assay of human *hedgehog* to chick *patched* protein ectopically expressed in *Xenopus laevis* oocytes. The assay system of Marigo et al. can be adapted to the present drug screening assays. As illustrated in that reference, *Shh* binds to the *patched* protein in a selective, saturable, dose-dependent manner, thus demonstrating that *patched* is a receptor for *Shh*.

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Complex formation between the *hedgehog* polypeptide and a *hedgehog* receptor may be detected by a variety of techniques. For instance, modulation of the formation of complexes can be quantitated using, for example, detectably labelled proteins such as radiolabelled, fluorescently labelled, or enzymatically labelled *hedgehog* polypeptides, by immunoassay, or by chromatographic detection.

Typically, for cell-free assays, it will be desirable to immobilize either the *hedgehog* receptor or the *hedgehog* polypeptide to facilitate separation of receptor/*hedgehog* complexes from uncomplexed forms of one of the proteins, as well as to accommodate automation of the assay. In one embodiment, a fusion protein can be provided which adds a domain that allows the protein to be bound to a matrix. For example, glutathione-S-transferase/receptor (GST/receptor) fusion proteins can be adsorbed onto glutathione sepharose beads (Sigma Chemical, St. Louis, MO) or glutathione derivatized microtitre plates, which are then combined with the *hedgehog* polypeptide, e.g. an <sup>35</sup>S-labeled *hedgehog* polypeptide, and the test compound and incubated under conditions conducive to complex formation, e.g. at physiological conditions for salt and pH, though slightly more stringent conditions may be desired. Following incubation, the beads are washed to remove any unbound *hedgehog* 

polypeptide, and the matrix bead-bound radiolabel determined directly (e.g. beads placed in scintillant), or in the supernatant after the receptor/hedgehog complexes are dissociated. Alternatively, the complexes can be dissociated from the bead, separated by SDS-PAGE gel, and the level of hedgehog polypeptide found in the bead fraction quantitated from the gel using standard electrophoretic techniques.

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Other techniques for immobilizing proteins on matrices are also available for use in the subject assay. For instance, soluble portions of the hedgehog receptor protein can be immobilized utilizing conjugation of biotin and streptavidin. For instance, biotinylated receptor molecules can be prepared from biotin-NHS (N-hydroxy-succinimide) using techniques well known in the art (e.g., biotinylation kit, Pierce Chemicals, Rockford, IL), and immobilized in the wells of streptavidin-coated 96 well plates (Pierce Chemical). Alternatively, antibodies reactive with the *hedgehog* receptor but which do not interfere with hedgehog binding can be derivatized to the wells of the plate, and the receptor trapped in the weils by antibody conjugation. As above, preparations of a hedgehog polypeptide and a test compound are incubated in the receptor-presenting wells of the plate, and the amount of receptor/hedgehog complex trapped in the well can be quantitated. Exemplary methods for detecting such complexes, in addition to those described above for the GST-immobilized complexes, include immunodetection of complexes using antibodies reactive with the hedgehog polypeptide, or which are reactive with the receptor protein and compete for binding with the hedgehog polypeptide; as well as enzyme-linked assays which rely on detecting an enzymatic activity associated with the hedgehog polypeptide. In the instance of the latter, the enzyme can be chemically conjugated or provided as a fusion protein with the hedgehog polypeptide. To illustrate, the hedgehog polypeptide can be chemically crosslinked or genetically fused with alkaline phosphatase, and the amount of hedgehog polypeptide trapped in the complex can be assessed with a chromogenic substrate of the enzyme, e.g. paranitrophenylphosphate. Likewise, a fusion protein comprising the hedgehog polypeptide and glutathione-S-transferase can be provided, and complex formation quantitated by detecting the GST activity using 1-chloro-2,4-dinitrobenzene (Habig et al (1974) J Biol Chem 249:7130).

For processes which rely on immunodetection for quantitating one of the proteins trapped in the complex, antibodies against the protein, such as the anti-hedgehog antibodies described herein, can be used. Alternatively, the protein to be detected in the complex can be "epitope tagged" in the form of a fusion protein which includes, in addition to the hedgehog polypeptide or hedgehog receptor sequence, a second polypeptide for which antibodies are readily available (e.g. from commercial sources). For instance, the GST fusion proteins described above can also be used for quantification of binding using antibodies against the GST moiety. Other useful epitope tags include myc-epitopes (e.g., see Ellison et al. (1991) J Biol Chem 266:21150-21157) which includes a 10-residue sequence from c-myc, as well as the pFLAG system (International Biotechnologies, Inc.) or the pEZZ-protein A system (Pharamacia, NJ).

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Where the desired portion of the *hedgehog* receptor (or other *hedgehog* binding molecule) cannot be provided in soluble form, liposomal vesicles can be used to provide manipulatable and isolatable sources of the receptor. For example, both authentic and recombinant forms of the *patched* protein can be reconstituted in artificial lipid vesicles (e.g. phosphatidylcholine liposomes) or in cell membrane-derived vesicles (see, for example, Bear et al. (1992) *Cell* 68:809-818, Newton et al. (1983) *Biochemistry* 22:6110-6117; and Reber et al. (1987) *J Biol Chem* 262:11369-11374).

In addition to cell-free assays, such as described above, the readily available source of hedgehog proteins provided by the art also facilitates the generation of cell-based assays for identifying small molecule agonists/antagonists and the like. Analogous to the cell-based assays described above for screening combinatorial libraries, cells which are sensitive to hedgehog induction, e.g. patched-expressing cells or other lung-derived cells sensitive to hedgehog induction, can be contacted with a hedgehog protein and a test agent of interest, with the assay scoring for anything from simple binding to the cell to modulation in hedgehog inductive responses by the target cell in the presence and absence of the test agent. As with the cell-free assays, agents which produce a statistically significant change in hedgehog activities (either inhibition or potentiation) can be identified.

In other emdodiments, the cell-based assay scores for agents which disrupt association of *patched* and *smoothened* proteins, e.g., in the cell surface membrane or liposomal preparation.

In addition to characterizing cells that naturally express the *patched* protein, cells which have been genetically engineered to ectopically express *patched* can be utilized for drug screening assays. As an example, cells which either express low levels or lack expression of the *patched* protein, e.g. *Xenopus laevis* oocytes, COS cells or yeast cells, can be genetically modified using standard techniques to ectopically express the *patched* protein. (see Marigo et al., *supra*).

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The resulting recombinant cells, e.g., which express a functional patched receptor, can be utilized in receptor binding assays to identify agonist or anatagonsts of hedgehog binding. Binding assays can be performed using whole cells. Furthermore, the recombinant cells of the present invention can be engineered to include other heterolgous genes encoding proteins involved in hedgehog-dependent siganl pathways. For example, the gene products of one or more of smoothened, costal-2 and/or fused can be co-expressed with patched in the reagent cell, with assays being sensitive to the functional reconstitution of the hedgehog signal transduction cascade.

Alternatively, liposomal preparations using reconstituted patched protein can be utilized. Patched protein purified from detergent extracts from both authentic and recombinant origins can be reconstituted in in artificial lipid vesicles (e.g. phosphatidylcholine liposomes) or in cell membrane-derived vesicles (see, for example, Bear et al. (1992) Cell 68:809-818; Newton et al. (1983) Biochemistry 22:6110-6117; and Reber et al. (1987) J Biol Chem 262:11369-11374). The lamellar structure and size of the resulting liposomes can be characterized using electron microscopy. External orientation of the patched protein in the reconstituted membranes can be demonstrated, for example, by immunoelectron microscopy. The hedgehog protein binding activity of liposomes containing patched and liposomes without the protein in the presence of candidate agents can be compared in order to identify potential modulators of the hedgehog-patched interaction.

The *hedgehog* protein used in these cell-based assays can be provided as a purified source (natural or recombinant in origin), or in the form of cells/tissue which express the

protein and which are co-cultured with the target cells. As in the cell-free assays, where simple binding (rather than induction) is the *hedgehog* activity scored for in the assay, the protein can be labelled by any of the above-mentioned techniques, e.g., fluorescently, enzymatically or radioactively, or detected by immunoassay.

In addition to binding studies, functional assays can be used to identified modulators, i.e., agonists or antagonists, of *hedgehog* or *patched* activities. By detecting changes in intracellular signals, such as alterations in second messengers or gene expression, in *patched* expressing cells contacted with a test agent, candidate agonists and antagonists to *patched* signaling can be identified.

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A number of gene products have been implicated in *patched*-mediated signal transduction, including *patched*, the transcription factor *cubitus interruptus* (ci), the serine/threonine kinase *fused* (fu) and the gene products of *costal-2*, *smoothened* and *suppressor of fused*.

The interaction of a hedgehog protein with patched sets in motion a cascade involving the activation and inhibition of downstream effectors, the ultimate consequence of which is, in some instances, a detectable change in the transcription or translation of a gene. Potential transcriptional targets of patched signaling are the patched gene itself (Hidalgo and Ingham, 1990 Development 110, 291-301; Marigo et al., 1996) and the vertebrate homologs of the drosophila cubitus interruptus gene, the GLI genes (Hui et al. (1994) Dev Biol 162:402-413). Putched gene expression has been shown to be induced in cells of the limb bud and the neural plate that are responsive to Shh. (Marigo et al. (1996) PNAS; Marigo et al. (1996) Development 122:1225-1233). The GLI genes encode putative transcription factors having zinc finger DNA binding domains (Orenic et al. (1990) Genes & Dev 4:1053-1067; Kinzler et al. (1990) Mol Cell Biol 10:634-642). Transcription of the GLI gene has been reported to be upregulated in response to hedgehog in limb buds, while transcription of the GL13 gene is downregulated in response to hedgehog induction (Marigo et al. (1996) Development 122:1225-1233). By selecting transcriptional regulatory sequences from such target genes, e.g. from patched or GLI genes, that are responsible for the up- or down regulation of these genes in response to patched signalling, and operatively linking such promoters to a reporter gene, one can derive a transcription based assay which is sensitive to the ability of a specific

test compound to modify *patched* signalling pathways. Expression of the reporter gene, thus, provides a valuable screening tool for the development of compounds that act as agonists or antagonists of *ptc* induction of differentiation/quiescence.

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Reporter gene based assays of this invention measure the end stage of the above described cascade of events, e.g., transcriptional modulation. Accordingly, in practicing one embodiment of the assay, a reporter gene construct is inserted into the reagent cell in order to generate a detection signal dependent on *ptc* signaling. To identify potential regulatory elements responsive to *ptc* signaling present in the transcriptional regulatory sequence of a target gene, nested deletions of genomic clones of the target gene can be constructed using standard techniques. See, for example, <u>Current Protocols in Molecular Biology</u>, Ausubel, F.M. et al. (eds.) Greene Publishing Associates, (1989); U.S. Patent 5,266,488; Sato et al. (1995) *J Biol Chem* 270:10314-10322; and Kube et al. (1995) *Cytokine* 7:1-7. A nested set of DNA fragments from the gene's 5'-flanking region are placed upstream of a reporter gene, such as the luciferase gene, and assayed for their ability to direct reporter gene expression in *patched* expressing cells. Host cells transiently transfected with reporter gene constructs can be scored for the induction of expression of the reporter gene in the presence and absence of *hedgehog* to determine regulatory sequences which are responsice to *patched*-dependent signalling.

In practicing one embodiment of the assay, a reporter gene construct is inserted into the reagent cell in order to generate a detection signal dependent on second messengers generated by induction with *hedgehog* protein. Typically, the reporter gene construct will include a reporter gene in operative linkage with one or more transcriptional regulatory elements responsive to the *hedgehog* activity, with the level of expression of the reporter gene providing the *hedgehog*-dependent detection signal. The amount of transcription from the reporter gene may be measured using any method known to those of skill in the art to be suitable. For example, mRNA expression from the reporter gene may be detected using RNAse protection or RNA-based PCR, or the protein product of the reporter gene may be identified by a characteristic stain or an intrinsic activity. The amount of expression from the reporter gene is then compared to the amount of expression in either the same cell in the absence of the test compound (or *hedgehog*) or it may be compared with the amount of

transcription in a substantially identical cell that lacks the target receptor protein. Any statistically or otherwise significant difference in the amount of transcription indicates that the test compound has in some manner altered the signal transduction of the *patched* protein, e.g., the test compound is a potential *ptc* therapeutic.

As described in further detail below, in preferred embodiments the gene product of the reporter is detected by an intrinsic activity associated with that product. For instance, the reporter gene may encode a gene product that, by enzymatic activity, gives rise to a detection signal based on color, fluorescence, or luminescence. In other preferred embodiments, the reporter or marker gene provides a selective growth advantage, e.g., the reporter gene may enhance cell viability, relieve a cell nutritional requirement, and/or provide resistance to a drug.

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Preferred reporter genes are those that are readily detectable. The reporter gene may also be included in the construct in the form of a fusion gene with a gene that includes desired transcriptional regulatory sequences or exhibits other desirable properties. Examples of reporter genes include, but are not limited to CAT (chloramphenicol acetyl transferase) (Alton and Vapnek (1979), Nature 282: 864-869) luciferase, and other enzyme detection systems, such as beta-galactosidase; firefly luciferase (deWet et al. (1987), Mol. Cell. Biol. 7:725-737); bacterial luciferase (Engebrecht and Silverman (1984), PNAS 1: 4154-4158; Baldwin et al. (1984), Biochemistry 23: 3663-3667); alkaline phosphatase (Toh et al. (1989) Eur. J. Biochem. 182: 231-238, Hall et al. (1983) J. Mol. Appl. Gen. 2: 101), human placental secreted alkaline phosphatase (Cullen and Malim (1992) Methods in Enzymol. 216:362-368).

Transcriptional control elements which may be included in a reporter gene construct include, but are not limited to, promoters, enhancers, and repressor and activator binding sites. Suitable transcriptional regulatory elements may be derived from the transcriptional regulatory regions of genes whose expression is induced after modulation of a *patched* signal transduction pathway. The characteristics of preferred genes from which the transcriptional control elements are derived include, but are not limited to, low or undetectable expression in quiescent cells, rapid induction at the transcriptional level within minutes of extracellular simulation, induction that is transient and independent of new protein synthesis, subsequent

shut-off of transcription requires new protein synthesis, and mRNAs transcribed from these genes have a short half-life. It is not necessary for all of these properties to be present.

In yet other embodiments, second messenger generation can be measured directly in the detection step, such as mobilization of intracellular calcium, phospholipid metabolism or adenylate cyclase activity are quantitated, for instance, the products of phospholipid hydrolysis IP<sub>3</sub>, DAG or cAMP could be measured. For example, recent studies have implicated protein kinase A (PKA) as a possible component of hedgehog/patched signaling (Hammerschmidt et al. (1996) Genes & Dev 10:647). High PKA activity has been shown to antagonize hedgehog signaling in these systems. Although it is unclear whether PKA acts directly downstream or in parallel with hedgehog signaling, it is possible that hedgehog signalling occurs via inhibition of PKA activity. Thus, detection of PKA activity provides a potential readout for the instant assays.

In a preferred embodiment, the *ptc* therapeutic is a PKA inhibitor. A variety of PKA inhibitors are known in the art, including both peptidyl and organic compounds. For instance, the *ptc* therapeutic can be a 5-isoquinolinesulfonamide, such as represented in the general formula:

wherein.

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 $R_1$  and  $R_2$  each can independently represent hydrogen, and as valence and stability permit a lower alkyl, a lower alkenyl, a lower alkynyl, a carbonyl (such as a carboxyl, an ester, a formate, or a ketone), a thiocarbonyl (such as a thioester, a thioacetate, or a thioformate), an amino, an acylamino, an amido, a cyano, a nitro, an azido, a sulfate, a sulfonate, a sulfonamido,  $-(CH_2)_m-R_8$ ,  $-(CH_2)_m-OH$ ,  $-(CH_2)_m-OH$ ,  $-(CH_2)_m-OH$ 

lower alkenyl,  $-(CH_2)_n$ -O- $(CH_2)_m$ -R<sub>8</sub>,  $-(CH_2)_m$ -SH,  $-(CH_2)_m$ -S-lower alkyl,  $-(CH_2)_m$ -S-lower alkenyl,  $-(CH_2)_n$ -S- $-(CH_2)_m$ -R<sub>8</sub>, or

 $R_1$  and  $R_2$  taken together with N form a heterocycle (substituted or unsubstituted);

 $R_3$  is absent or represents one or more substitutions to the isoquinoline ring such as a lower alkyl, a lower alkynyl, a carbonyl (such as a carboxyl, an ester, a formate, or a ketone), a thiocarbonyl (such as a thioester, a thioacetate, or a thioformate), an amino, an acylamino, an amido, a cyano, a nitro, an azido, a sulfate, a sulfonate, a sulfonamido,  $-(CH_2)_m-R_8$ ,  $-(CH_2)_m-OH$ ,  $-(CH_2)_m-OH$  over alkyl,  $-(CH_2)_m-OH$  over alkenyl,  $-(CH_2)_m-OH$  alkenyl,  $-(CH_2)_m-CH$  alkenyl,  $-(CH_2)_m-C$ 

R<sub>8</sub> represents a substituted or unsubstituted aryl, aralkyl, cycloalkyl, cycloalkenyl, or heterocycle; and

n and m are independently for each occurrence zero or an integer in the range of 1 to 6.

In a preferred embodiment, the PKA inhibitor is N-[2-((p-bromocinnamyl)amino)ethyl]-5-isoquinolinesulfonamide (H-89; Calbiochem Cat. No. 371963), e.g., having the formula:

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In another embodiment, the PKA inhibitor is 1-(5-isoquinolinesulfonyl)-2-methylpiperazine (H-7; Calbiochem Cat. No. 371955), e.g., having the formula:

In still other embodiments, the PKA inhibitor is KT5720 (Calbiochem Cat. No. 420315), having the structure

A variety of nucleoside analogs are also useful as PKA inhibitors. For example, the subject method can be carried out cyclic AMP analogs which inhibit the kinase activity of PKA, as for example, 8-bromo-cAMP or dibutyryl-cAMP

$$\begin{array}{c} \mathsf{NH}_2 \\ \mathsf{N} \\$$

Exemplary peptidyl inhibitors of PKA activity include the PKA Heat Stable Inhibitor (isoform  $\alpha$ ; see, for example, Calbiochem Cat. No. 539488, and Wen et al. (1995) *J Biol Chem* 270:2041).

Certain *hedehog* receptors may stimulate the activity of phospholipases. Inositol lipids can be extracted and analyzed using standard lipid extraction techniques. Water soluble derivatives of all three inositol lipids (IP<sub>1</sub>, IP<sub>2</sub>, IP<sub>3</sub>) can also be quantitated using radiolabelling techniques or HPLC.

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The mobilization of intracellular calcium or the influx of calcium from outside the cell may be a response to *hedgehog* stimulation or lack there of. Calcium flux in the reagent cell can be measured using standard techniques. The choice of the appropriate calcium indicator, fluorescent, bioluminescent, metallochromic, or Ca<sup>++</sup>-sensitive microelectrodes depends on the cell type and the magnitude and time constant of the event under study (Borle (1990) *Environ Health Perspect* 84:45-56). As an exemplary method of Ca<sup>++</sup> detection, cells could be loaded with the Ca<sup>++</sup> sensitive fluorescent dye fura-2 or indo-1, using standard methods, and any change in Ca<sup>++</sup> measured using a fluorometer.

In certain embodiments of the assay, it may be desirable to screen for changes in cellular phosphorylation. As an example, the drosophila gene *fused* (fu) which encodes a serine/threonine kinase has been identified as a potential downstream target in *hedgehog* signaling. (Preat et al., 1990 *Nature* 347, 87-89; Therond et al. 1993, *Mech. Dev.* 44. 65-80). The ability of compounds to modulate serine/threonine kinase activation could be screened using colony immunoblotting (Lyons and Nelson (1984) *Proc. Natl. Acad. Sci. USA* 81:7426-7430) using antibodies against phosphorylated serine or threonine residues. Reagents for performing such assays are commercially available, for example, phosphoserine and phosphothreonine specific antibodies which measure increases in phosphorylation of those residues can be purchased from comercial sources.

In yet another embodiment, the *ptc* therapeutic is an antisense molecule which inhibits expression of a protein involved in a *patched*-mediated signal transduction pathway. To illustrate, by inhibiting the expression of a protein which are involved in *patched* signals,

such as fused, costal-2, smoothened and/or Gli genes, the ability of the *patched* signal pathway(s) to inhibit proliferation of a cell can be altered, e.g., potentiated or repressed.

As used herein, "antisense" therapy refers to administration or in situ generation of oligonucleotide probes or their derivatives which specifically hybridize (e.g. bind) under cellular conditions with cellular mRNA and/or genomic DNA encoding a hedgehog protein, patched, or a protein involved in patched-mediated signal transduction. The hybridization should inhibit expression of that protein, e.g. by inhibiting transcription and/or translation. The binding may be by conventional base pair complementarity, or, for example, in the case of binding to DNA duplexes, through specific interactions in the major groove of the double helix. In general, "antisense" therapy refers to the range of techniques generally employed in the art, and includes any therapy which relies on specific binding to oligonucleotide sequences.

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An antisense construct of the present invention can be delivered, for example, as an expression plasmid which, when transcribed in the cell, produces RNA which is complementary to at least a unique portion of the target cellular mRNA. Alternatively, the antisense construct is an oligonucleotide probe which is generated *ex vivo* and which, when introduced into the cell causes inhibition of expression by hybridizing with the mRNA and/or genomic sequences of a target gene. Such oligonucleotide probes are preferably modified oligonucleotide which are resistant to endogenous nucleases, e.g. exonucleases and/or endonucleases, and is therefore stable *in vivo*. Exemplary nucleic acid molecules for use as antisense oligonucleotides are phosphoramidate, phosphothioate and methylphosphonate analogs of DNA (see also U.S. Patents 5,176,996; 5,264,564; and 5,256,775). Additionally, general approaches to constructing oligomers useful in antisense therapy have been reviewed, for example, by Van der Krol et al. (1988) *Biotechniques* 6:958-976; and Stein et al. (1988) *Cancer Res* 48:2659-2668.

Several considerations should be taken into account when constructing antisense oligonucleotides for the use in the methods of the invention: (1) oligos should have a GC content of 50% or more; (2) avoid sequences with stretches of 3 or more G's; and (3) oligonucleotides should not be longer than 25-26 mers. When testing an antisense oligonucleotide, a mismatched control can be constructed. The controls can be generated by

reversing the sequence order of the corresponding antisense oligonucleotide in order to conserve the same ratio of bases.

In an illustrative embodiment, the *ptc* therapeutic can be an antisense construct for inhibiting the expression of *patched*, e.g., to mimic the inhibition of *patched* by *hedgehog*. Exemplary antisense constructs include:

5'-GTCCTGGCGCCGCCGCCGTCGCC

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- 5'-TTCCGATGACCGGCCTTTCGCGGTGA
- 5'-GTGCACGGAAAGGTGCAGGCCACACT

## VI. Exemplary pharmaceutical preparations of hedgehog and ptc therapeutics

The source of the *hedgehog* and *ptc* therapeutics to be formulated will depend on the particular form of the agent. Small organic molecules and peptidyl fragments can be chemically synthesized and provided in a pure form suitable for pharmaceutical/cosmetic usage. Products of natural extracts can be purified according to techniques known in the art. For example, the Cox et al. U.S. Patent 5,286,654 describes a method for purifying naturally occurring forms of a secreted protein and can be adapted for purification of *hedgehog* polypeptides. Recombinant sources of *hedgehog* polypeptides are also available. For example, the gene encoding *hedgehog* polypeptides, are known, *inter alia*, from PCT publications WO 95/18856 and WO 96/17924.

Those of skill in treating lung tissues can determine the effective amount of an ptc, hedgehog or fgf-10 therapeutic to be formulated in a pharmaceutical or cosmetic preparation.

The ptc, hedgehog or fgf-10 therapeutic formulations used in the method of the invention are most preferably applied in the form of appropriate compositions. As appropriate compositions there may be cited all compositions usually employed for systemically or topically administering drugs. The pharmaceutically acceptable carrier should be substantially inert, so as not to act with the active component. Suitable inert carriers include water, alcohol polyethylene glycol, mineral oil or petroleum gel, propylene glycol and the like.

To prepare the pharmaceutical compositions of this invention, an effective amount of the particular ptc, hedgehog or fgf-10 therapeutic as the active ingredient is combined in intimate admixture with a pharmaceutically acceptable carrier, which carrier may take a wide variety of forms depending on the form of preparation desired for administration. These pharmaceutical compositions are desirable in unitary dosage form suitable, particularly for administration orally, rectally, percutaneously, or by parenteral injection. For example, in preparing the compositions in oral dosage form, any of the usual pharmaceutical media may be employed such as, for example, water, glycols, oils, alcohols and the like in the case of oral liquid preparations such as suspensions, syrups, elixirs and solutions; or solid carriers such as starches, sugars, kaolin, lubricants, binders, disintegrating agents and the like in the case of powders, pills, capsules, and tablets. Because of their ease in administration, tablets and capsules represents the most advantageous oral dosage unit form, in which case solid pharmaceutical carriers are obviously employed. For parenteral compositions, the carrier will usually comprise sterile water, at least in large part, though other ingredients, for example, to aid solubility, may be included. Injectable solutions, for example, may be prepared in which the carrier comprises saline solution, glucose solution or a mixture of saline and glucose solution. Injectable suspensions may also be prepared in which case appropriate liquid carriers, suspending agents and the like may be employed. Also included are solid form preparations which are intended to be converted, shortly before use, to liquid form preparations. In the compositons suitable for percutaneous administration, the carrier optionally comprises a penetration enhancing agent and/or a suitable wetting agent, optionally combined with suitable additives of any nature in minor proportions, which additives do not introduce a significant deleterious effect on the skin.

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In addition to the direct topical application of the preparations they can be topically administered by other methods, for example, encapsulated in a temperature and/or pressure sensitive matrix or in film or solid carrier which is soluble in body fluids and the like for subsequent release, preferably sustained-release of the active component.

As appropriate compositions for topical application there may be cited all compositions usually employed for topically administering therapeuites, e.g., creams, gellies, dressings, shampoos, tinctures, pastes, ointments, salves, powders, liquid or semiliquid

formulation and the like. Application of said compositions may be by aerosol e.g. with a propellent such as nitrogen carbon dioxide, a freon, or without a propellent such as a pump spray, drops, lotions, or a semisolid such as a thickened composition which can be applied by a swab. In particular compositions, semisolid compositions such as salves, creams, pastes, gellies, ointments and the like will conveniently be used.

It is especially advantageous to formulate the subject compositions in dosage unit form for ease of administration and uniformity of dosage. Dosage unit form as used in the specification and claims herein refers to physically discreate units suitable as unitary dosages, each unit containing a predetermined quantity of active ingredient calculated to produce the desired therapeutic effect in association with the required pharmaceutical carrier. Examples of such dosage unit forms are tablets (including scored or coated tablets), capsules, pills, powders packets, wafers, injectable solutions or suspensions, teaspoonfuls, tablespoonfuls and the like, and segregated multiples thereof.

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The pharmaceutical preparations of the present invention can be used, as stated above, for the many applications which can be considered cosmetic uses. Cosmetic compositions known in the art, preferably hypoallergic and pH controlled are especially preferred, and include toilet waters, packs, lotions, skin milks or milky lotions. The preparations contain, besides the *ptc*, *hedgehog* or *fgf-10* therapeutic, components usually employed in such preparations. Examples of such components are oils, fats, waxes, surfactants, humectants, thickening agents, antioxidants, viscosity stabilizers, chelating agents, buffers, preservatives, perfumes, dyestuffs, lower alkanols, and the like. If desired, further ingredients may be incorporated in the compositions, e.g. antiinflammatory agents, antibacterials, antifungals, disinfectants, vitamins, sunscreens, antibiotics, or other anti-acne agents.

Examples of oils comprise fats and oils such as olive oil and hydrogenated oils; waxes such as beeswax and lanolin; hydrocarbons such as liquid paraffin, ceresin, and squalane; fatty acids such as stearic acid and oleic acid; alcohols such as cetyl alcohol, stearyl alcohol, lanolin alcohol, and hexadecanol; and esters such as isopropyl myristate, isopropyl palmitate and butyl stearate. As examples of surfactants there may be cited anionic surfactants such as sodium stearate, sodium cetylsulfate, polyoxyethylene laurylether phosphate, sodium N-acyl glutamate; cationic surfactants such as stearyldimethylbenzylammonium chloride and

stearyltrimethylammonium chloride; ampholytic surfactants such as alkylaminoethylglycine hydrocloride solutions and lecithin; and nonionic surfactants such as glycerin monostearate, sorbitan monostearate, sucrose fatty acid esters, propylene glycol monostearate, polyoxyethylene oleylether, polyethylene glycol monostearate, polyoxyethylene sorbitan monopalmitate, polyoxyethylene coconut fatty acid monoethanolamide, polyoxypropylene glycol (e.g. the materials sold under the trademark "Pluronic"), polyoxyethylene castor oil, and polyoxyethylene lanolin. Examples of humectants include glycerin, 1,3-butylene glycol, and propylene glycol; examples of lower alcohols include ethanol and isopropanol; examples of thickening agents include xanthan gum, hydroxypropyl cellulose, hydroxypropyl methyl cellulose, polyethylene glycol and sodium carboxymethyl cellulose; examples of antioxidants comprise butylated hydroxytoluene, butylated hydroxyanisole, propyl gallate, citric acid and ethoxyquin; examples of chelating agents include disodium edetate and ethanehydroxy diphosphate; examples of buffers comprise citric acid, sodium citrate, boric acid, borax, and preservatives examples of disodium hvdrogen phosphate; and parahydroxybenzoate, ethyl parahydroxybenzoate, dehydroacetic acid, salicylic acid aud benzoic acid.

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For preparing ointments, creams, toilet waters, skin milks, and the like, typically from 0.01 to 10% in particular from 0.1 to 5% and more in particular from 0.2 to 2.5% of the active ingredient, e.g., of the ptc, hedgehog or fgf-10 therapeutic, will be incorporated in the compositions. In ointments or creams, the carrier for example consists of 1 to 20%, in particular 5 to 15% of a humectant, 0.1 to 10% in particular from 0.5 to 5% of a thickener and water; or said carrier may consist of 70 to 99%, in particular 20 to 95% of a surfactant, and 0 to 20%, in particular 2.5 to 15% of a fat; or 80 to 99.9% in particular 90 to 99% of a thickener; or 5 to 15% of a surfactant, 2-15% of a humectant, 0 to 80% of an oil, very small (<2%) amounts of preservative, coloring agent and/or perfume, and water. In a toilet water, the carrier for example consists of 2 to 10% of a lower alcohol, 0.1 to 10% or in particular 0.5 to 1% of a surfactant, 1 to 20%, in particular 3 to 7% of a humectant, 0 to 5% of a buffer, water and small amounts (<2%) of preservative, dyestuff and/or perfume. In a skin milk, the carrier typically consists of 10-50% of oil, 1 to 10% of surfactant, 50-80% of water and 0 to 3% of preservative and/or perfume. In the aforementioned preparations, all % symbols refer to weight by weight percentage.

Particular compositions for use in the method of the present invention are those wherein the ptc, hedgehog or fgf-10 therapeutic is formulated in liposome-containing compositions. Liposomes are artificial vesicles formed by amphiphatic molecules such as polar lipids, for example, phosphatidyl cholines, ethanolamines and serines, sphingomyelins, cardiolipins, plasmalogens, phosphatidic acids and cerebiosides. Liposomes are formed when suitable amphiphathic molecules are allowed to swell in water or aqueous solutions to form liquid crystals usually of multilayer structure comprised of many bilayers separated from each other by aqueous material (also referred to as coarse liposomes). Another type of liposome known to be consisting of a single bilayer encapsulating aqueous material is referred to as a unilamellar vesicle. If water-soluble materials are included in the aqueous phase during the swelling of the lipids they become entrapped in the aqueous layer between the lipid bilayers.

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Water-soluble active ingredients such as, for example, various salt forms of a hedgehog polypeptide, are encapsulated in the aqueous spaces between the molecular layers. The lipid soluble active ingredient of ptc, hedgehog or fgf-10 therapentic, such as an organic mimetic, is predominantly incorporated into the lipid layers, although polar head groups may protude from the layer into the aqueous space. The encapsulation of these compounds can be achieved by a number of methods. The method most commonly used involves casting a thin film of phospholipid onto the walls of a flask by evaporation from an organic solvent. When this film is dispersed in a suitable aqueous medium, multilamellar liposomes are formed. Upon suitable sonication, the coarse liposomes form smaller similarly closed vesicles.

Water-soluble active ingredients are usually incorporated by dispersing the cast film with an aqueous solution of the compound. The unencapsulated compound is then removed by centrifugation, chromatography, dialysis or other art-known suitable procedures. The lipid-soluble active ingredient is usually incorporated by dissolving it in the organic solvent with the phospholipid prior to casting the film. If the solubility of the material in the lipid phase is not exceeded or the amount present is not in excess of that which can be bound to the lipid, liposomes prepared by the above method usually contain most of the material bound in the lipid bilayers; separation of the liposomes from unencapsulated material is not required.

A particularly convenient method for preparing liposome formulated forms of hedgehog and ptc therapeutics is the method described in EP-A-253,619, incorporated herein by reference. In this method, single bilayered liposomes containing encapsulated active ingredients are prepared by dissolving the lipid component in an organic medium, injecting the organic solution of the lipid component under pressure into an aqueous component while simultaneously mixing the organic and aqueous components with a high speed homogenizer or mixing means, whereupon the liposomes are formed spontaneously.

The single bilayered liposomes containing the encapsulated ptc, hedgehog or fgf-10 therapeutic can be employed directly or they can be employed in a suitable pharmaceutically acceptable carrier for topical administration. The viscosity of the liposomes can be increased by the addition of one or more suitable thickening agents such as, for example xanthan gum, hydroxypropyl cellulose, hydroxypropyl methylcellulose and mixtures thereof. The aqueous component may consist of water alone or it may contain electrolytes, buffered systems and other ingredients, such as, for example, preservatives. Suitable electrolytes which can be employed include metal salts such as alkali metal and alkaline earth metal salts. The preferred metal salts are calcium chloride, sodium chloride and potassium chloride. The concentration of the electrolyte may vary from zero to 260 mM, preferably from 5 mM to 160 mM. The aqueous component is placed in a suitable vessel which can be adapted to effect homogenization by effecting great turbulence during the injection of the organic component. Homogenization of the two components can be accomplished within the vessel, or, alternatively, the aqueous and organic components may be injected separately into a mixing means which is located outside the vessel. In the latter case, the liposomes are formed in the mixing means and then transferred to another vessel for collection purpose.

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The organic component consists of a suitable non-toxic, pharmaceutically acceptable solvent such as, for example ethanol, glycerol, propylene glycol and polyethylene glycol, and a suitable phospholipid which is soluble in the solvent. Suitable phospholipids which can be employed include lecithin, phosphatidylcholine, phosphatydylserine, phosphatidylethanolamine, phosphatidylinositol, lysophosphatidylcholine and phospha-tidyl glycerol, for example. Other lipophilic additives may be employed in order to selectively modify the

characteristics of the liposomes. Examples of such other additives include stearylamine, phosphatidic acid, tocopherol, cholesterol and lanolin extracts.

In addition, other ingredients which can prevent oxidation of the phospholipids may be added to the organic component. Examples of such other ingredients include tocopherol, butylated hydroxyanisole, butylated hydroxytoluene, ascorbyl palmitate and ascorbyl oleate. Preservatives such a benzoic acid, methyl paraben and propyl paraben may also be added.

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Apart from the above-described compositions, use may be made of covers, e.g. plasters, bandages, dressings, gauze pads and the like, containing an appropriate amount of a ptc, hedgehog or fgf-10 therapeutic. In some cases use may be made of plasters, bandages, dressings, gauze pads and the like which have been impregnated with a topical formulation containing the therapeutic formulation.

## Exemplification

The invention now being generally described, it will be more readily understood by reference to the following examples which are included merely for purposes of illustration of certain aspects and embodiments of the present invention, and are not intended to limit the invention.

The mammalian lung, like many other organs, develops by branching morphogenesis of an epithelium [see ref. 1]. Development initiates with evagination of two ventral buds of foregut endoderm into the underlying splanchnic mesoderm. As they extend, they send out lateral branches at precise, invariant positions establishing the primary airways and the lobes of each lung. Dichotomous branching leads to further extension of the airways. Grafting studies have demonstrated the importance of bronchial mesenchyme in inducing epithelial branching, but the significance of epithelial signaling is largely unstudied. The morphogen Sonic hedgehog (Shh) is widely expressed in the foregut endoderm and is specifically upregulated in the distal epithelium of the lung where branching is occurring [see ref. 2]. Ectopic expression of Shh disrupts branching and increases proliferation suggesting that local Shh signaling regulates lung development [see ref. 2]. We report here that Shh is essential for

development of the respiratory system. In *Shh* null mutants, the trachea and esophagus do not separate properly and the lungs form a rudimentary sac due to failure of branching and growth after formation of the primary lung buds. Interestingly, normal proximo-distal differentiation of the airway epithelium occurs, indicating that Shh is not needed for differentiation events. In addition, the transcription of several mesenchymally expressed downstream targets of Shh is abolished. These results highlight the importance of epithelially derived Shh in regulating branching morphogenesis of the lung.

## Results and discussion

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To address the role of *Shh* in respiratory tract development, we examined a null mutant of the gene (3). At 10.5 days *post coitum* (dpc) of embryonic mouse development, the lung of wild- type (wt) siblings consists of a left and right bud [see ref. 1]. By 12.5 dpc, the trachea epithelium has separated ventrally from the esophageal component of the foregut and the two lung buds have formed several lateral branches which will give rise to primary airways of the lung lobes (Figure 1a-c). In contrast, the esophageal and tracheal tubes remain closely associated in *Shh* mutants (Figure 1d,e) and although left and right buds form, they either have not branched or possess one abnormally positioned branch point (Figure 1f). Wild- type lungs undergo considerable growth and branching in organ culture. However, in explant culture of lungs from *Shh* mutants, bronchial mesenchyme cells detach from the endoderm and the epithelium fails to grow, or branch extensively (data not shown). We conclude that the defect in branching morphogenesis is independent of other *Shh*-expressing organs (i.e., the gut), and that the observed branching phenotype reflects an absence of Shh signaling which is normally associated with the branching process.

To determine if branching is merely delayed and whether *Shh* plays a role in differentiation, we examined lungs removed at 15.5 (data not shown) and 18.5 dpc (Figure 1g,h). At this time, five well-developed lobes are evident in the wild-type (four right, one left), and highly branched airways form a ramifying epithelial network, the respiratory tree (Figure 1i,k,l). To mediate gas exchange in the alveolar sacs, the respiratory surface is well vascularized (Figure 1g). In contrast, *Shh* mutants form only a rudimentary respiratory organ with a few large, poorly vascularized airways (Figure 1h). Trachea and esophagus are so

closely juxtaposed that their tubes share some common epithelium (Figure 1e) and a fistulalike fusion of the alimentary and respiratory tract is formed, mirroring a lethal anomaly well described in human pathology [see ref. 4,5] (Figure 1j,m).

Remarkably, despite the absence of branching, evidence of normal proximo-distal epithelial differentiation can be observed. Most proximally, the pulmonary epithelium forms a columnar epithelium typical of the mainstem bronchi (Figure 1m) and expresses *CCSP* [see ref. 6], a marker for terminally differentiated secretory Clara cells (Figure 1q). More distally, the epithelium consists of a mixture of columnar and cuboidal epithelium as observed in the bronchioles (Figure 1n), and alveolar air sacs are formed which correspondingly express *SP-C* [see ref. 7], a type II pneumocyte marker (Figure 1r).

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In summary, *Shh* is not required for proximo-distal differentiation of lung epithelium, but is essential for three different events of regional morphogenesis of the foregut endoderm, formation of the tracheoesophageal septum, lung lobation and generation of the respiratory tree, all of which are essential in forming a functional lung.

The exact role for *Shh* in branching processes remains to be determined. Grafting studies indicate that, whereas budding can be supported by mesenchyme from many different sources, only bronchial mesenchyme can induce organotypic branching morphogenesis [see ref. 8]. The requirement for *Shh* in the epithelium suggests that regulation of its expression may be a reciprocal epithelial response to mesenchymal signaling.

To examine in more detail how Shh might regulate early branching of the lung epithelium, we performed digoxigenin in situ hybridization with probes recognizing general targets of *Hedgehog* signaling (Figure 2a-e and data not shown), or genes specifically implicated in lung morphogenesis (Figure 2f-k). As *Shh* mutants are growth retarded and show a general delay in lung budding, we compared expression of these markers at 12.5 dpc with wild type embryos collected at 11.5 and 12.5 dpc.

Patched genes encode proteins thought to be Hedgehog receptors, while Gli-genes encode transcriptional mediators of Hedgehog signaling [see ref. 9]. Both Ptc-1 and Gli-1 are up-regulated when Shh is ectopically expressed in the lung indicating that here, as elsewhere in the embryo, they are transcriptional targets of Shh signaling [see ref. 9,10]. Consistent with

this model, Ptc-1 and Gli-1 are normally expressed in the mesenchyme of wild-type embryos with highest levels at the distal branch points mirroring epithelial Shh expression [see ref. 10] (Figure 2a,c). In Shh mutants, only basal levels of expression of both genes are detected (Figure 2a,c). Gli-3 which shows more wide-spread expression in the mesenchyme is also down-regulated (Figure 2e). In contrast, Ptc-2 which is expressed at higher levels in the epithelium and Gli-2, which is normally expressed more uniformly in the mesenchyme are not altered (Figure 2b,d). These data indicate that the lung mesenchyme, not the epithelium, is most likely the direct cellular target of Shh signaling. Further, they suggest that modulation of Gli-1 and Gli-3 transcription may be a critical aspect of lung morphogenesis. As Gli-1 mutants do not have a lung phenotype, the Shh phenotype cannot simply be ascribed to a loss of Gli-1 transcriptional activity [see ref. 10]. Given that post-transcriptional processing regulates Gli (Ci) activity in invertebrates [see ref. 11], we cannot rule out that Gli-2 is expressed, but posttranscriptionally inactivated. Gli genes are clearly involved in lung development, as evidenced by the relatively weak lobular hypoplasia observed in Gli-3 mutants [see ref. 10], but revealing the full extent of Gli action may require the generation of compound mutants.

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Several lines of evidence indicate that *hedgehog* signaling regulates the expression of *Bmp*, *Wnt* and *FGF* family members [see ref. 11]. In the lung, *Bmp-4* is strongly expressed in the distal-most tips of the epithelium. Ectopic expression results in decreased epithelial proliferation, disrupted branching and reduced differentiation of distal cell types in the airway [see ref. 12]. In *Shh* mutants, *Bmp 4* is expressed in the normal position but at higher levels (Figure 2f), suggesting that enhanced *Bmp 4* signaling could contribute to the block in branching. *Wnt-7b* is normally expressed in the lung epithelium and is required for normal branching (S. Lee, W. Cardoso, B. Parr & A. McMahon; unpublished), whereas *Wnt-2* is expressed in the underlying mesenchyme suggesting a role in epithelial maintenance [see ref. 2]. In *Shh* mutants, *Wnt-7b* expression is not altered (Figure 2g) but *Wnt-2* expression is down-regulated (Figure 2h). This observation lends further support to the model that the lung mesenchyme is the primary target of Shh signaling and indicates that mesenchymal signaling is abnormal in *Shh* mutants. However, no role for Wnt-2 in lung development has been reported in *Wnt-2* mutants [see ref. 13].

Ectopic expression of a dominant negative form of FGF-R2 in the lung epithelium arrests branching after formation of left and right buds which then grow caudally as tubes, differentiating into proximal epithelial structures only [see ref. 14]. An arrest in branching after initial budding is reminiscent of Shh mutants, but there are clearly differences in subsequent morphogenesis and differentiation which is largely unaffected in Shh mutants. The recent observation that Fgf10 is expressed in mesenchyme cells preceding branch formation and can induce branching of lung epithelium in culture, points to its role as a putative ligand [see ref. 15]. In Shh mutants, expression of FGF-R2 is unaltered (Figure 2I). In contrast, Fgf10 which in wild-type embryos is highly localized to small patches of mesenchyme at a distance from the lung epithelium (arrows in Figure 2j), is expressed broadly in mesenchyme immediately adjacent to the epithelium in the mutant lung. These results indicate that Shh is not required for Fgf10 expression. Further, they suggest that Shh signaling may spatially restrict Fgf10 expression to the distal mesenchyme. Such an inhibitory role for Shh in the local regulation of Fgf10 expression is supported by transgenic studies [see ref. 16]. The intriguing possibility that the altered position of Fgf10 expression then disrupts branching remains to be determined.

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 $HNF-3\beta$  and Nkx-2.1 are specific transcriptional effectors of Shh signaling in the neural tube. In the gut, HNF-3b is widely expressed in the epithelium, including the lung, whereas Nkx-2.1 expression is specific to the lung epithelium and a few other endodermal derivatives [see ref. 17]. Mice lacking Nkx 2.1 develop cystic unbranched lungs indicating that it is essential for lung morphogenesis [see ref. 17]. Expression of both genes is unaltered in the epithelium of Shh mutant lungs suggesting that in this organ their expression is independent of the Shh signaling pathway (Figure 2k and data not shown).

As loss of *Shh* activity predominantly affects the expression of mesenchyme markers, we analyzed late mesenchyme differentiation. Formation of cartilage rings, albeit disorganized, occurs in the mutant (Figure 3a), while the layer of smooth muscle typically lining the proximal epithelium is absent (Figure 3b). The observation that Shh is required for formation of smooth muscle is in agreement with previous studies [see ref. 18].

In summary, the results reported here establish Shh as a regulator of foregut development and more specifically as a key factor in the control of branching morphogenesis

in the mouse lung. They also indicate that the genetic control of growth and branching in the lung epithelium is most likely a complex process involving both epithelial and mesenchymal interactions at the branch points, and that the downstream targets of Shh signaling in this organ are primarily mesenchymally expressed genes.

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### Materials and methods

#### Shh mutants

Generation of the *Shh* mutants has been described elsewhere [see ref. 3]. Mice homozygous for the null allele appear phenotypically identical to those reported in [see ref. 19].

## Histological/in situ analysis

Tissue was processed for standard histology, or a modified in situ hybridization procedure [see ref. 20].

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## Antibody staining

Antibody staining with a monoclonal antibody against smooth muscle actin (Sigma) was carried out according to the manufacturer's instructions.

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- All of the above-cited references and publications are hereby incorporated by reference.

#### Equivalents

Those skilled in the art will recognize, or be able to ascertain using no more than routine experimentation, numerous equivalents to the specific polypeptides, nucleic acids, methods, assays and reagents described herein. Such equivalents are considered to be within the scope of this invention.

#### We claim:

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1. A method for modulating the growth state of lung tissue, or cells derived therefrom, comprising ectopically contacting the tissue with an amount of an agent effective to alter the rate of proliferation of the lung tissue, wherein the agent is selected from the group consisting of a hedgehog therapeutic, a ptc therapeutic and an fgf-10 therapeutic.

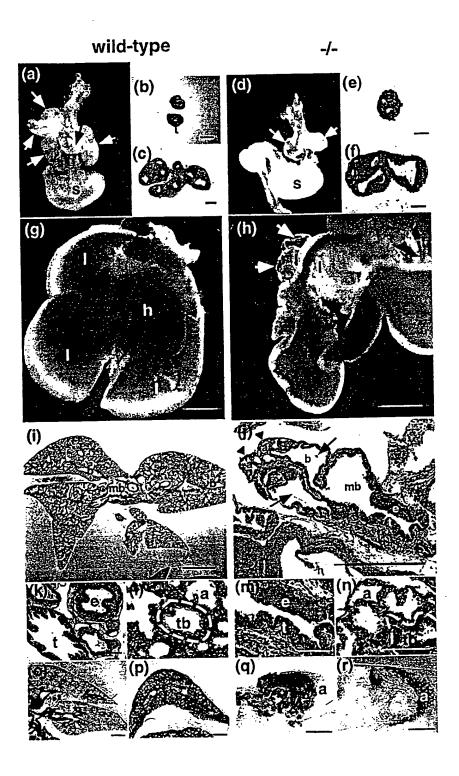
- 2. A method for inducing the formation of, or the maintenance or functiona performance of lung tissue, comprising contacting the lung tissue with an amount of an agent effective to induce the formation of new lung tissue, wherein the agent is selected from the group consisting of a hedgehog therapeutic, a pic therapeutic and an fgf-10 therapeutic.
- 3. The method of claim 1, wherein the lung tissue is in culture, and the agent is provided as a cell culture additive.
- 4. The method of claim 1, wherein the cell is treated in an animal and the agent is administered to the animal as a therapeutic composition.
  - 5. The method of claim 1, wherein the agent is a hedgehog therapeutic.
  - 6. The method of claim 5, wherein the *hedgehog* therapeutic is a polypeptide including a *hedgehog* polypeptide sequence of at least a bioactive extracellular portion of a *hedgehog* protein.
- 7. The method of claim 6, wherein the polypeptide includes at least 50 amino acids residues of an N-terminal half of the hedgehog protein
  - 8. The method of claim 6, wherein the polypeptide includes at least 100 amino acids of an extracellular domain of the *hedgehog* protein.
- The method of claim 6, wherein the polypeptide includes at least a portion of the
   hedgehog protein corresponding to a 19kd fragment of an extracellular domain of the
   hedgehog protein.
  - 10. The method of claim 6, wherein the *hedgehog* protein is encoded by a gene of a vertebrate organism.

11. The method of claim 6, wherein the polypeptide includes a hedgehog polypeptide sequence represented in the general formula of SEQ ID No. 21.

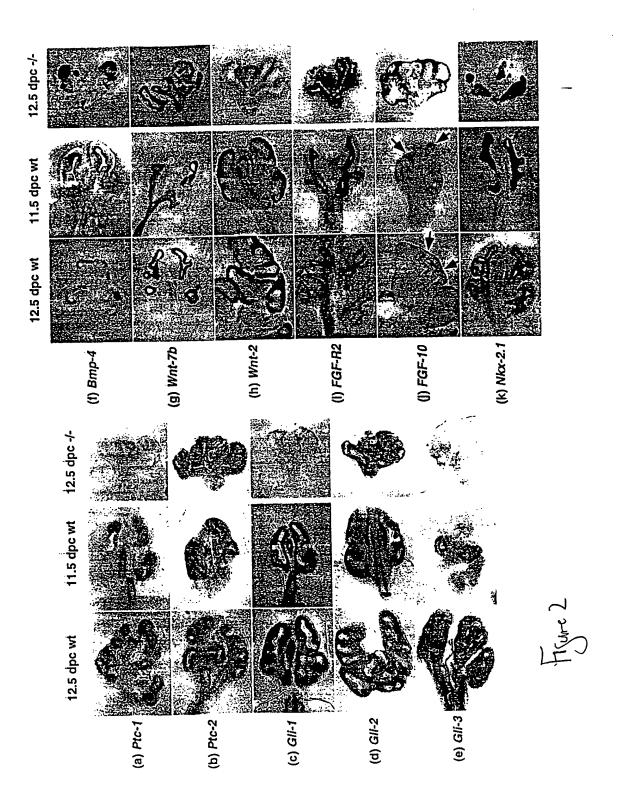
- 12. The method of claim 6, wherein the polypeptide includes a *hedgehog* polypeptide sequence represented in the general formula of SEQ ID No. 22.
- 5 13. The method of claim 6, wherein the hedgehog protein is encoded by a human hedgehog gene.
  - 14. The method of claim 6, wherein the *hedgehog* polypeptide sequence is at least 60 percent identical to an amino acid sequence of a *hedgehog* protein selected from the group consisting of SEQ ID No:9, SEQ ID No:10, SEQ ID No:11, SEQ ID No:12, SEQ ID No:13, SEQ ID No:14, SEQ ID No:15 and SEQ ID No:16.
  - 15. The method of claim 6, wherein the *hedgehog* polypeptide sequence is encodable by a nucleotide sequence which hybridizes under stringent conditions to a sequence selected from the group consisting of SEQ ID No:1, SEQ ID No:2, SEQ ID No:3, SEQ ID No:4, SEQ ID No:5, SEQ ID No:6, SEQ ID No:7 and SEQ ID No:8.
- 15 16. The method of claim 6, wherein the *hedgehog* polypeptide sequence is an amino acid sequence of a *hedgehog* protein selected from the group consisting of SEQ ID No:9, SEQ ID No:10, SEQ ID No:11, SEQ ID No:12, SEQ ID No:13, SEQ ID No:14, SEQ ID No:15 and SEQ ID No:16.
- 17. The method of claim 6, wherein the *hedgehog* polypeptide sequence is an amino acid sequence of a Sonic *hedgehog* protein.
  - 18. The method of claim 1, wherein the agent is a ptc therapeutic.

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- 19. The method of claim 18, wherein the ptc therapeutic is a small organic molecule which binds to a patched protein and derepresses patched-mediated inhibition of mitosis.
- 20. The method of claims 18, wherein the ptc therapeutic binds to patched and mimics hedgehog-mediated patched signal transduction.
  - 21. The method of claim 20, wherein the ptc therapeutic is a small organic molecule.



Frare 1



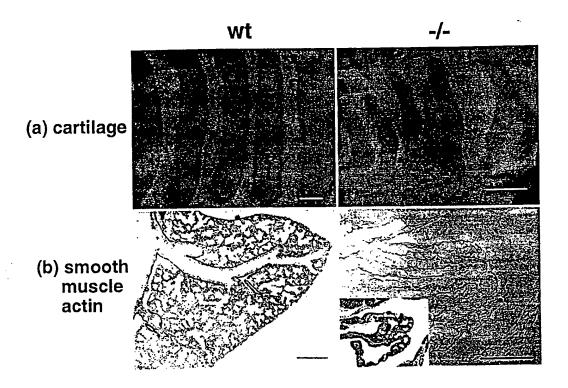


Figure 3

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	ССТ	GCC	<b>ም</b> ፐር	GAC	ATC	ACC	ACG	TCf	GAC	CGT	GAC	CGT	AAT	AAG	TAT	GGT	480
				Asp													
	145		200	1124		150			-		155					160	
25																	
	TTG	TTG	GCG	CGC	CTA	GCT	GTG	GAA	GCC	GGA	TTC	GAC	TGG	GTC	TAC	TAC	528
				Arg													
					165					170					175		
30	GAG	TCC	CGC	AAC	CAC	ATC	CAC	GTA	TCG	GTC	AAA	GCT	GAT	AAC	TCA	CTG	576
	Glu	Ser	Arg	Asn	His	Ile	His	Val	Ser	Val	Lys	Ala	Азр	Asn	Ser	Leu	
				180					185					190			
				GCC													624
35	Ala	Val	Arg	Ala	Gly	Gly	Cys	Phe	Pro	Gly	Asn	Ala	Thr	Vai	Arg	Leu	
			195					200					205				
				GAA													672
	Arg	Ser	Gly	Glu	Arg	Lys	Gly	Leu	Arg	Glu	Leu	His	Arg	Gly	Asp	Trp	

		210	-			•	215					220					
	GTA	CTG	GCC	GCT	GAT	GCA	GCG	GGC	CGA	GTG	GTA	ccc	ACG	CCA	GTG	CTG	720
									Arg								
5	225	Dea				230		,	5		235					240	-
	СТС	TTC	CTG	GAC	CGG	GAT	CTG	CAG	CGC	CGC	GCC	TCG	TTC	GTG	GCT	GTG	768
	Leu	Phe	Leu	Asp	Arg	Asp	Leu	Gln	Arg	Arg	Ala	Ser	Phe	Val	Ala	Val	
					245					250					255		
10																	
	GAG	ACC	GAG	CGG	CCT	CCG	CGC	AAA	CTG	TTG	CTC	ACA	CCC	TGG	CAT	CTG	816
	Glu	Thr	Glu	Arg	Pro	Pro	Arg	Lys	Leu	Leu	Leu	Thr	Pro	Trp	H.i.s	Leu	
				260					265					270			
15	GTG	TTC	GCT	GCT	CGC	GGG	CCA	GCG	CCT	GCT	CCA	GGT	GAC	TTT	GCA	CCG	864
	Val	Ph€	Ala	Ala	Arg	Gly	Pro	Ala	Pro	Ala	Pro	Gly	Asp	P'ne	Ala	Pro	
			275					280					285				
									GGC								912
20	Val	Phe	Ala	Arg	Arg	Leu	Arg	Ala	Gly	Asp	Ser	Val	Leu	Ala	Pro	Gly	
		290					295					300					
						•											
									GTA								960
	Gly	Asp	Ala	Leu	Gln	Pro	Ala	Arg	Val	Ala	Arg	Val	Ala	Arg	Glu		
25	305					310					315					320	
									ACT								1008
	Ala	Val	Gly	Val	Phe	Ala	Pro	Leu	Thr	Ala	His	СŢЪ	Thr	Leu		Val	
					325					330					335		
30																	
									GCG							TGG	1056
	Asn	Asp	Val	Leu	Ala	Ser	Cys	Tyr	Ala	Val	Leu	Glu	Ser	His	Gln	Trp	
				340					345					350			
35									CGG								1104
	Ala	His	Arg	Ala	Phe	Ala	Pro	Leu	Arg	Leu	Leu	His	Ala	Leu	Gly	Ala	
			355					360					365				
	CTG	CTC	CCT	GGG	GGT	GCA	GTC	CAG	CCG	ACT	GGC	ATG	CAT	TGG	TAC	TCT	1152

	Leu	Le12 370	Pro	Gly	Gly	Ala	Val 375	Gln	Pro	Thr	Ġly	Met 380	His	Trp	Tyr	Ser	
5								GAG Glu					ТG				1190
10	(2)							NO:3:									
		(i)	( <i>F</i>	A) LE B) TY	ENGT: YPE:	i: 12	281 N Leic	ISTICoase acic both	pai:	cs							
15		, , , , ,		) T(	OPOL	OGY:	line	ear									
						itu.	Colv	•									
20		(ix		A) NA	E: AME/I CCATI			1233									
25		(xi	) SE(	QUENC	CE DI	ESCR:	IPT I	ON:	SEQ :	ID N	0:3:						
													<b>T</b> GT Cys				48
30										Arg			GGG Gly				96
35	GTG Val	GTG Val	GGC Gly 35	AGC Ser	CGC Arg	CGG Arg	AGG Arg	CCG Pro	CCT Pro	CGC Arg	AAG Lys	CTC Leu	GTG Val 45	CCT Pro	CTT Leu	GCC Ala	144
	TAC Tyr	AAG Lys	CAG Gln	TTC Phe	AGC Ser	CCC Pro	AAC Asn	GTG Val	CCG Pro	GAG Glu	AAG Lys	ACC Thr	CTG Leu	GGC Gly	GCC Ala	AGC Ser	192

		50					55					60						-
	666	ccc	<b>ም</b> አር	GAA	GGC	AAG	ጋጥሬ	GCG	CGC	AGC	тст	GAG	CGC	TTC	AAA	GAG	240	
				Glu														
_	_	Arg	Tyr	GIU	СТУ	70	116	Ala	Arg	501	75	01-	5			80		
5	65					70					, 5							
	CTC	ACC	CCC	AAC	TAC	AAT	CCC	GAC	ATC	ATC	ттс	AAG	GAC	GAG	GAG	AAC	288	
	Leu	Thr	Pro	Asn	Tyr	Asn	Pro	Asp	Ile	Ile	Phe	Lys	Asp	Glu	Glu	Asn		
					85					90					95			
0																		
	ACG	GGT	GCC	GAC	CGC	CTC	ATG	ACC	CAG	CGC	TGC	AAG	GAC	CGT	CTG	AAC	336	
	Thr	Gly	Ala	Asp	Arg	Leu	Met	Thr	Gln	Arg	Cys	Lys	Asp	Arg	Leu	Asn		
				100					105					110				
15	TCA	CTG	GCC	ATC	TCT	GTC	ATG	AAC	CAG	TGG	CCT	GGT	GTG	AAA	CTG	CGG	384	
	Ser	Leu	Ala	Ile	Ser	Val	Met	Asn	Gln	Trp	Pro	Gly	Val	Lys	Leu	Arg		
			115					120					125					
	GTG	ACC	GAA	GGC	CGG	GAT	GAA.	GAT	GGC	CAT	CAC	TCA	GAG	GAG	TCT	TTA	432	
20	Val	Thr	Glu	Gly	Arg	Asp	Glu	Asp	Gly	His	His	Ser	Glu	Glu	Ser	Leu		
		130					135					140						
	CAC	ŢĄŢ	GAG	GGC	CGC	GCG	GTG	GAT	ATC	ACC	ACC	TCA	GAC	CGT	GAC	CGA	480	
	His	Tyr	Glu	Gly	Arg	Ala	Val	Asp	Ile	Thr	Thr	Ser	Asp	Arg	Asp	Arg		
25	145					150					155					160		
	AAT	AAG	TAT	GGA	CTG	CTG	GCG	CGC	TTA	GCA	GTG	GAG	GCC	GGC	TTC	GAC	528	
	Asn	Lys	Tyr	Gly	Leu	Leu	Ala	Arg	Leu	Ala	Val	Glu	Ala	Gly	Phe	Asp		
					165					170					175			
30																		
	TGG	GTG	TAT	TAC	GAG	TCC	AAG	GCC	CAC	GTG	CAT	TGC	TCT	GTC	AAG	TCT	576	
				Tyr												Ser		
				180					185					190				
35	GAG	CAT	TCG	GCC	GCT	GCC	AAG	ACA	GGT	GGC	TGC	TTT	CCT	GCC	GGA	GCC	624	
				Ala														
			195					200	-				205					
	CAG	GTG	CGC	CTA	GAG	AAC	GGG	GAG	CGT	GTG	GCC	CTG	TCA	GCT	GTA	AAG	672	

																_	
	Gln	Val	Arg	Leu	Glu	Asn		Glu	Arg	Val	Ala		Ser	Ala	Val	Lys	
		210					215					220					
				~~~	cmc	ome.	000	3.00	666	C 4 C	C M III	ccc	N.C.C	ccc	አ ርር	ጥጥር	720
_				CGG													720
5		GIŊ	Asp	Arg	vaı		АТА	мес	сту	GIU	235	сту	1111	FIO	1111	240	
	225					230					233					240	
	ΔίξΤ	СДТ	GTG.	CTT	ΑͲ <b>Ͳ</b>	TTC	CTG	GAC	CGC	GAG	CCA	AAC	CGG	CTG	AGA	GCT	768
				Leu													
10	502	7.00	, , ,		245				5	250			_		255		
•																	
	TTC	CAG	GTC	ATC	GAG	ACT	CAG	GAT	CCT	CCG	CGT	CGG	CTG	GCG	СТС	ACG	816
	Phe	Gln	Val	Ile	Glu	Thr	Gln	Asp	Pro	Prc	Arg	Arg	Leu	Ala	Leu	Thr	
				260					265					270			
15																	
	CCT	GCC	CAC	CTG	CTC	TTC	ATT	GCG	GAC	ŢAĄ.	CAT	ACA	GAA	CCA	GCA	GCC	864
	Pro	Ala	His	Leu	Leu	Phe	Ile	Ala	Asp	Asn	His	Thr	Glu	Pro	Ala	Ala	
			275					280					285				
20				GCC													912
	His		Arg	Ala	Thr	Phe		Ser	His	Val	Gln		Gly	GIn	Tyr	Val	
		290					295					300					
				GGG	cm s	CCD	ccc	CTC.	CNC	CCT	CCT	ccc	CTC	CCN	ССТ	GTC.	960
25				Gly													300
25	305	vaı	ser	GIY	Vai	310	G1. y	пец	GIII	110	315	n. g	Vu.	1114	1120	320	
	303	,				J10					515						
	TCC	ACC	CAC	GTG	GCC	СТТ	GGG	TCC	TAT	GCT	CCT	CTC	ACA	AGG	CAT	GGG	1008
				Val													
30					325		-		-	330					335		
	ACA	CTT	GTG	GTG	GAG	GAT	GTG	GTG	GCC	TCC	TGC	TTT	GCA	GCT	GTG	GCT	1056
	Thr	Leu	Val	Val	Glu	Asp	Val	Val	Ala	Ser	Cys	Phe	Ala	Ala	Val	Ala	
				340					345					350			
35																	
	GAC	CAC	CAT	CTG	GCT	CAG	TTG	GCC	TTC	TGG	CCC	CTG	CGA	CTG	TTT	CCC	1104
	Asp	His	His	Leu	Ala	Gln	Leu	Ala	Phe	Trp	Pro	Leu	Arg	Leu	Phe	Pro	
			355					360					365				

	AGT TTG GCA TGG GGC AGC TGG ACC CCA AGT GAG GGT GTT CAC TCC TAC	1152
	Ser Leu Ala Trp Gly Ser Trp Thr Pro Ser Glu Gly Val His Ser Tyr	
	370 375 380	
5	CCT CAG ATG CTC TAC CGC CTG GGG CGT CTC TTG CTA GAA GAG AGC ACC	12 <del>00</del>
	Pro Gln Met Leu Tyr Arg Leu Gly Arg Leu Leu Glu Glu Ser Thr	
	385 390 395 400	
	TTC CAT CCA CTG GGC ATG TCT GGG GCA GGA AGC TGAAGGGACT CTAACCACT	G 1253
10	Phe His Pro Leu Gly Met Ser Gly Ala Gly Ser	
	405 410	
	CCCTCCTGGA ACTGCTGTGC GTGGATCC	1281
15	AND THE PROPERTY OF THE AND	
	(2) INFORMATION FOR SEQ ID NO:4:	
	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 1313 base pairs	
20	(B) TYPE: nucleic acid	
20	(C) STRANDEDNESS: both	
	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: cDNA	
25		
	(ix) FEATURE:	
	(A) NAME/KEY: CDS	
	(B) LOCATION: 11314	
30		
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
	ATG CTG CTG CTG GCC AGA TGT TTT CTG GTG ATC CTT GCT TCC TCG	48
35	Met Leu Leu Leu Ala Arg Cys Phe Leu Val Ile Leu Ala Ser Ser	
	1 5 10 15	
		0.0
	CTG CTG GTG TGC CCC GGG CTG GCC TGT GGG CCC GGC AGG GGG TTT GGA	96
	Leu Leu Val Cys Pro Gly Leu Ala Cys Gly Pro Gly Arg Gly Phe Gly	
	10	

				20					25					30			
	AAG	AGG	CGG	CAC	CCC	AAA	AAG	CTG	ACC	CCT	TTA	GCC	TAC	AAG	CAG	TTT	144
						Lys											
5	-		35					40					45				
	ATT	CCC	AAC	GTA	GCC	GAG	AAG	ACC	CTA	GGG	GCC	AGC	GGC	AGA	TAT	GAA	192
	Ile	Pro	Asn	Val	Ala	Glu	Lys	Thr	Leu	Gly	Ala	Ser	Gly	Arg	Tyr	Glu	
		50					55					60					
10																	
	GGG	AAG	ATC	ACA	AGA	AAC	TCC	GAA	CGA	TTT	AAG	GAA	CTC	ACC	CCC	AAT	240
	Gly	Lys	Ile	Thr	Arg	Asn	Ser	Glu	Arg	Phe	Lys	Glu	Leu	Thr	Prc	Asn	
	<b>6</b> 5					70					75					80	
15	TAC	AAC	ccc	GAC	ATC	ATA	TTT	AAG	GAT	GAG	GAA	AAC	ACG	GGA	GCA	GAC	288
	Ίγτ	Asn	Pro	Asp	Ile	Ile	Phe	Lys	Asp	Glu	Glü	Asn	Thr	Gly	Ala	Asp	
					85					90					95		
	CGG	CTG	ATG	ACT	CAG	AGG	TGC	AAA	GAC	AAG	TTA	ААТ	GCC	TTG	GCC	ATC	336
20	Arg	Leu	Met	Thr	Gln	Arg	Cys	Lys	Asp	Lys	Leu	Asn	Ai.a	Leu	Ala	Ile	
				100					105					110			
	TCT	GTG	ATG	AAC	CAG	TGG	ССТ	GGA	GTG	AGG	CTG	CGA	GTG	VCC	GAG	GGC	384
	Ser	Val	Met	Asn	Gln	Trp	Pro	Gly	Val	Arg	Leu	Arg	Val	Thr	Glu	Gly	
25			115					120					125				
	TGG	GAT	GAG	GAC	GGC	CAT	CAT	TCA	GAG	GAG	TCT	СТА	CAC	TAT	GAG	GGT	432
	Trp	Asp	Glu	Asp	Gly	His	His	Ser	Glυ	Glu	Ser	Leu	His	Tyr	Glu	Gly	
		130					135					140					
30																	
	CGA	GCA	GTG	GAC	ATC	ACC	ACG	TCC	GAC	CGG	GAC	CGC	AGC	AAG	TAC	GGC	480
	Arg	Ala	Val	Asp	Ile	Thr	Thr	Ser	Asp	Arg	Asp	Arg	Ser	Lys	Tyr	Gly	
	145					150					155					160	
35	ATG	CTG	GCT	CGC	CTG	GCT	GTG	GAA	GCA	GGT	TTC	GAC	TGG	GTC	TAC	TAT	528
	Met	Leu	Ala	Arg	Leu	Ala	Val	Glu	Ala	Gly	Phe	Asp	Trp	Val	Tyr	Tyr	
					165					170					175		
	GAA	TCC	AAA	GCT	CAC	ATC	CAC	TGT	TCT	GTG	AAA	GCA	GAG	AAC	TCC	GTG	576

	Glu	Ser	Lys	Ala 180	His	Ile	His	Cys	Ser 185	Val	Lys	Ala	Glu	Asn 190	Ser	Val		
5			AAA Lys										Thr					624 —
	GAG	CAG	195 GGC	GGC	ACC	AAG	СТG	200 GTG	AAG	GAC	TTA	CGT	205 CCC	GGA	GAC	CGC		672
10	Glu	Gln 210	Gly	Gly	Thr	Lys	Leu 215	Val	Lys	Asp	Leu	Arg 220	Pro	Gly	Asp	Arg		
			GCG Ala															720
15			CTG Leu															768
20			CTG Leu															816
25			GTG Val 275	GCG					TCG									864
30			TTT Phe															912
			CGC Arg															960
35			CTG Leu														1	800

	CAC	GGC	ACC	ATT	ĊТС	ATC	AAC	CGG	GTG	CTC	GCC	TCG	TGC	TAC	GCT	GTC	1056
	His	Gly	Thr	Ile	Leu	Ile	Asn	Arg	Val	Leu	Ala	Ser	Cys	Tyr	Ala	Val	
				340					345					350			
5	ATC	GAG	GAG	CAC	AGC	TGG	GCA	CAC	CGG	GCC	TTC	GCG	CCT	TTC	CGC	CTG	1104
	Ile	Glu	Glu	His	Ser	Trp	Ala	His	Arg	Ala	Phe	Ala	Pro	Phe	Arg	Leu	
			355					360					365				
			GCG														1152
10	Ala	His	Ala	Leu	Leu	Ala	Ala	Leu	Ala	Pro	Ala	Arg	Thr	Asp	Gly	Gly	
		370					375					380					
			GGC														1200
	Gly	Gly	Gly	Ser	Ile	Pro	Ala	Ala	Gln	Ser		Thr	Glu	Ala	Arg		
15	385					390					395					400	
															<b></b>	07.0	7040
			CCG														1248
	Ala	Glu	Pro	Thr		G1 Å	Ile	His	Trp		Ser	Gin	⊥eu	Leu		HIS	
					405					410					415		
20				mcc.	ome	mmc.	CAC	7.00	C D C	7.00	A TO C	C N TP	CCZ	<b>ም</b> ምር	CCN	NTC.	1296
			ACC														1290
	11e	GIĀ	Thr		Leu	Leu	Asp	ser	425	Thr	мес	итг	PIO	430	GLY	Mec	
				420					423					4.0			
25	GCG	ርጥር	AAG	ጥርር	AGC	тG											1313
-5			Lys			10											
	13T CI	141	435														

30 (2) INFORMATION FOR SEQ ID NO:5:

35

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 1256 base pairs

(B) TYPE: nucleic acid

(C) STRANDEDNESS: both

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: cDNA

### (ix) FEATURE:

(A) NAME/KEY: CDS

(B) LOCATION: 1..1257

		(xi)	SEÇ	QUENC	CE DE	ESCR:	PTIC	ON: S	SEQ :	ID NO	0:5:						
	ATG	CGG	CTT	TTG	ACG	AGA	GTG	CTG	CTG	GTG	тст	CTT	CTC	ACT	CTG	TCC	48
10			Leu														
	1				5					10					15		
			GTG														96
	Leu	Val	Val		Gly	Leu	Ala	Cys		Pro	Gly	Arg	Gly		Gly	Arg	
15				20					25					30			
	AGA	AGA	CAT	CCG	AAG	AAG	CTG	ACA	ССТ	CTC	GCC	TAC	AAG	CAG	TTC	ATA	144
	Arg	Arg	His	Pro	Lys	Lys	Leu	Thr	Pro	Leu	Ala	Tyr	Lys	Gln	Phe	Ile	
			35					40					45				
20															an c	000	102
			GTC														192
	Fro		Val	Ala	GIu	Lys	Thr 55	Leu	сту	Ala	ser	60 60	Arg	ıyı	GIU	Gry	
		50					33										
25	AAG	ATA	ACG	CGC	AAT	TCG	GAG	AGA	TTT	AAA	GAA	CTT	ACT	CCA	AAT	TAC	240
	Lys	Ile	Thr	Arg	Asn	Ser	Glu	Arg	Phe	Lys	Glu	Leu	Thr	Pro	Asn	Tyr	
	65					70					75					80	
			GAC														288
30	Asn	Pro	Asp	Ile		Phe	Lys	Asp	Glu		Asn	Thr	Gly	Ala	Asp 95	Arg	
					85					90					90		
	CTC	ATG	ACA	CAG	AGA	TGC	AAA	GAC	AAG	CTG	AAC	TCG	CTG	GCC	ATC	TCT	336
	Leu	Met	Thr	Gln	Arg	Cys	Lys	Asp	Lys	Leu	Asn	Ser	Leu	Ala	Ile	Ser	
35				100					105					110			
			AAC														384
	Val	Met	Asn	His	Trp	Pro	Gly		Lys	Leu	Arg	Val		Glu	Gly	Trp	
			115					120					125				

	GAT	GAG	GAC	GGT	CAC	CAT	TTT	GAA	GAA	TCA	CTC	CAC	TAC	GAG	GGA	AGA	432
	Asp	Glu	Asp	Gly	His	His	Phe	Glu	Glu	Ser	Leu	His	Tyr	Glu	Gly	Arg	
		130					135					140					
<b>5</b> .																	
	GCT	GTT	GAT	ATT	ACC	ACC	TCT	GAC	CGA	GAC	AAG	AGC	AAA	TAC	GGG	ACA	480
	Ala	Val	Asp	Ile	Thr	Thr	Ser	Asp	Arg	Asp	Lys	Ser	Lys	Tyr	Gly	Thr	
	145					150					155					160	
10				CTA													528
	Leu	Ser	Arg	Leu	Ala	Val	Glu	Ala	Gly		Asp	Trp	Val	Tyr		Glu	
					165					170					175		
						03 T	mca	m c m	CMC	א א א א	CCA	C 7 7	ስ <b>ለ</b> ጥ	ጥርር	ርምሞ	CCT	576
				CAC													0.0
15	Ser	гуѕ	Ala	His	iie	пто	Cys	261	185	цуз	AIO	J. G	71511	190	•••		
				180					103								
	GCG	ααα	יייטיד	GGG	GGC	TGT	TTC	CCA	GGT	TCC	GCT	CTG	GTC	TCG	CTC	CAG	624
				Gly													
20		-,-	195		-	-		200					205				
	GAC	GGA	GGA	CAG	AAG	GCC	GTG	AAG	GAC	CTG	AAC	CCC	GGA	GAC	AAG	GTG	672
	Asp	Gly	Gly	Gln	Lys	Ala	Val	Lys	Asp	Leu	Asn	Pro	Gly	Asp	Lys	Val	
		210					215					220					
25																	=00
				GAC													720
	Leu	Ala	Ala	Asp	Ser	Ala	Gly	Asn	Leu	Val		Ser	Asp	Phe	lle		
	225					230					235					240	
							100	7.00	CCT	CCT	CTTC	መመመ	ሞክሮ	GT/C	מדת	CDD	768
30				CGA													, 00
	Phe	Thr	Asp	Arg			Thr	Thr	Arg	250		FIIE	ıyı	Vai	255	010	
					245					250					200		
	200	C 7 7	C 7 7	CCC	CTT	CAA	ΔΔG	<b>ል</b> ፐር	ACC	CTC	ACC	GCC	GCT	CAC	CTC	CTT	816
7.5				Pro													
35	Thr	GIII	Gru	260	Vai	GIU	Lys		265					270			
				200					_00								
	ጥጥጥ	GTC	CTC	GAC	AAC	TCA	ACG	GAA	GAT	CTC	CAC	ACC	ATG	ACC	GCC	GCG	864
				Asp													
				- 4.													

			275					280					285				
						AGA Arg											912
5		290					295					300					
	AGC	GGT	CAG	CTT	AAA	TCT	GTC	ATC	GTG	CAG	CGG	ATA	TAC	ACG	GAG	GAG	960
	Ser	Gly	Gln	Leu	Lys	Ser	Val	Ile	Val	Gln	Arg	īle	Tyr	Thr	Glu	Glu	
	305					310					315					320	
10																	
	CAG	CGG	GGC	TCG	TTC	GCA	CCA	GTG	ACT	GCA	CAT	GGG	ACC	ATT	GTG	GTC	1008
	Gln	Arg	Gly	Ser	Phe	Ala	Pro	Val	Thr	Ala	His	Gly	Thr	Ile	Val	Val	
					325					330					335		
15	GAC	AGA	АТА	CTG	GCG	TCC	TGT	TAC	GCC	GTA	АТА	GAG	GAC	CAG	GGG	CTT	1056
						Ser											
	П	,		340			-	-	345					35C			
	GCG	CAT	TTG	GCC	TTC	GCG	CCC	GCC	AGG	CTC	TAT	TAT	TAC	GTG	TCA	TCA	1104
20						Ala											
			355					360					365				
	TTC	CTG	TCC	CCC	AAA	ACT	CCA	GCA	GTC	GGT	CCA	ATG	CGA	CTT	TAC	AAC	1152
	Phe	Leu	Ser	Pro	Lys	Thr	Pro	Ala	Val	Gly	Pro	Met	Arg	Leu	Tyr	Asn	
25		370					375					380					
	AGG	AGG	GGG	TCC	ACT	GGT	ACT	CCA	GGC	TCC	TGT	CAT	CAA	ATG	GGA	ACG	1200
	Arg	Arg	Gly	Ser	Thr	Gly	Thr	Pro	Gly	Ser	Cys	His	Gln	Met	Gly	Thr	
	385					390					395					400	
30																	
	TGG	CTT	TTG	GAC	AGC	AAC	ATG	CTT	CAT	CCT	TTG	GGG	ATG	TCA	GTA	AAC	1248
	Trp	Leu	Leu	Asp	Ser	Asn	Met	Leu	His	Pro	Leu	Gly	Met	Ser	Val	Asn	
					405					410					415		
																	1056
35	TCA	AGC	TG														1256
	Ser	Ser															

	(2)	INF	AMAC	TION	FOR	SEQ	ID	NO:6	:			•					
5		(i	(; (;	A) L B) T C) S	CE C ENGT YPE: TRAN OPOL	H: 1 nuc DEDN	425 leic ESS:	base aci sin	pai d	rs							
10		(ii	) MO:	LECU	LE T	YPE:	cDN.	A									
15		(ix	(2		E: AME/I			1425									
		(xi	) SE	QUEN	CE D	ESCR:	IPTI:	ON:	SEQ	ID N	0:6:						
20										CTA Leu 10						CTG- Leu	48
25										CCG Pro							96
										TTA Leu							144
30										GCC Ala					GAA Glu	GGG Gly	192
35										AAG Lys							240
	AAC	CCC	GAC	ATC	ATA	ттт	AAG	GAT	GAA	GAA	AAC	ACC	GGA	GCG	GAC	AGG	288

	Asn	Pro	Asp	Ile	Ile	Phe	Lys	Asp	Glu	Glu	Asn	Thr	Gly	Ala	Asp	Arg	
					85					90					95		
													mmc	200	3 mc	mcc	336
				CAG													330
5	Leu	Met	Thr	Gln	Arg	Cys	ьуs	Asp		ьeu	Asn	Ala	Leu	110	TTE	Ser	
				100					105					110			
	GTG	ATG	AAC	CAG	TGG	CCA	GGA	GTG	AAA	CTG	CGG	GTG	ACC	GAG	GGC	TGG	384
				Gln													
10			115		-			120					125				
	GAC	GAA	GAT	GGC	CAC	CAC	TCA	GAG	GAG	TCT	CTG	CAC	TAC	GAG	GGC	CGC	432
	Asp	Glu	Asp	Gly	His	His	Ser	Glu	Glu	Ser	Leu	His	Tyr	Glu	Gly	Arg	
		130					135					140					
15																	
				ATC													480
	Ala	Val	Asp	Ile	Thr	Thr	Ser	Asp	Arg	Asp		Ser	Lys	Tyr	Gly		
	145					150					155					160	
				CTG		C.T.C	CAC	ccc	ccc	mmc	CAC	ТСС	CTC	TAC	ሞውር	GAG	528
20				Leu													320
	Leu	Ата	Arg	reu	165	vai	Giu	VTO	Gry	170	1100	110	· u	- , -	175		•
					105					1.0							
	TCC	AAG	GCA	CAT	ATC	CAC	TGC	TCG	GTG	AAA	GCA	GAG	AAC	TCG	GTG	GCG	576
25	Ser	Lys	Ala	His	Ile	His	Cys	Ser	Val	Lys	Ala	Glu	Asn	Ser	Val	Ala	
				180					185					190			
				GGA													624
	Ala	Lys	Ser	Gly	Gly	Cys	Phe	Pro	Gly	Ser	Ala	Thr		His	Leu	Glu	
30			195					200					205				
															000	C.T.C.	672
				ACC													672
	Gln	_	Gly	Thr	Lys	Leu		Lys	Asp	Leu	Ser		GIY	Asp	Arg	vaı	
		210					215					220					
35				ar c	C. C. C	CT C	ccc	ccc	CTTC	CTC	መክር	אככ	GNC	ጥጥ⊂	ርሞር	ACT	720
				GAC													, 20
		Ala	Ala	Asp	ASP	230	GIÀ	Arg	neu	Pen	235	ner	voh	1116	104	240	
	225					2.JU					200					- · ·	

	TTC	CTG	GAC	CGC	GAC	GAC	GGC	GCC	AAG	AAG	GTC	TTC	TAC	GTG	ATC	GAG	768
	Phe	Leu	Asp	Arg	Asp	Asp	Gly	Ala	Lys	Lys	Val	Phe	Tyr	Val	Ile	Glu	
					245					250					255		
5	ልርር	cee	GAG	CCG	רפר	GAG	CGC	ርጥር	CTG	ርፐር	ACC	GCC	GCG	CAC	CTG	ርፐር	816
,									Leu								010
		5		260	5		3		265					270			
	TTT	GTG	GCG	CCG	CAC	AAC	GAC	TCG	GCC	ACC	GGG	GAG	ccc	GAG	GCG	TCC	864
10	Phe	Val	Ala	Pro	His	Asn	Asp	Ser	Ala	Thr	Gly	Glu	Pro	Glu	Ala	Ser	
			275					280					285				
	ሞርር	GGC	ፐርር	GGG	CCG	ርር ፕ	ፐርር	ece	GGC	GCA	CTG	GGG	ССТ	CGG	GCG	CTG	912
																Leu	712
15		290		4			295	•	•			300					
	TTC	GCC	AGC	CGC	GTG	CGC	CCG	GGC	CAG	CGC	GTG	TAC	GTG	GTG	GCC	GAG	960
	Phe	Ala	Ser	Arg	Val	Arg	Pro	Gly	Gln	Arg	Val	Tyr	Val	Val	Ala	Glu	
	305					310					315					320	
20	OCM	C A C	666	CAC	000	ccc	CTIC	CTC	ccc	ccc	CCT	CTC	CAC	ACC.	COC	NCC.	1008
									CCC Pro								1006
	Arg	Map	сту	nsp	325	ALG	nea	пеп	110	330	ALG	701	1113	261	335	1111	
25	CTA	AGC	GAG	GAG	GCC	GCG	GGC	GCC	TAC	GCG	CCG	CTC	ACG	GCC	CAG	GGC	1056
	Leu	Ser	Glu	Glu	Ala	Ala	Gly	Ala	Tyr	Ala	Pro	Leu	Thr	Ala	Gln	Gly	
				340					345					350			
																	1104
20									GCC								1104
30	inr	116	355	TTE	Asn	Arg	vai	360	Ala	sei	Cys	ıyı	365	Val	116	GIU	
			333					500					505				
	GAG	CAC	AGC	TGG	GCG	CAC	CGG	GCC	TTC	GCG	CCC	TTC	CGC	CTG	GCG	CAC	1152
	Glu	His	Ser	Trp	Ala	His	Arg	Ala	Phe	Ala	Pro	Phe	Arg	Leu	Ala	His	`` <del>`</del> .
35		370					375					380					
									GCG								1200
		Leu	Leu	Ala			Ala	Pro	Ala	Arg		Asp	Arg	Gly	Gly		
	385					390					395					400	

	AGC	GGC	GGC	GGG	GAC	CGC	GGG	GGC	GGC	GGC	GGC	AGA	GTA	GCC	CTA	ACC	1248
	Ser	Gly	Gly	Gly	Asp	Arg	Gly	Gly	Gly	Gly	Gly	Arg	Val	Ala	Leu	Thr	
					405					410					415		
5																	
	GCT	CCA	GGT	GCT	GCC	GAC	GCT	CCG	GGT	GCG	GGG	GCC	ACC	GCG	GGC	ATC	1296
	Ala	Pro	Gly	Ala	Ala	Asp	Ala	Pro	Gly	Ala	Gly	Ala	Thr	Ala	Gly	Ile	
				420					425					430			
10	CAC	TGG	TAC	TCG	CAG	CTG	CTC	TAC	CAA	ATA	GGC	ACC	TGG	CTC	CTG	GAC	1344
	His	Trp	Tyr	Ser	Gln	Leu	Leu	Tyr	Gln	Ile	Gly	Thr	Trp	Leu	Leu	Asp	
			435					440					445				
	AGC	GAG	GCC	CTG	CAC	CCG	CTG	GGC	ATG	GCG	GTC	AAG	TCC	AGC	NNN	AGC	1392
15	Ser	Glu	Ala	Leu	His	Pro	Leu	Gly	Met	Ala	Val	Lys	Ser	Ser	Xaa	Ser	
		450					455					460					
	CGG	GGG	GCC	GGG	GGA	GGG	GCG	CGG	GAG	GGG	GCC						1425
	Arg	Gly	Ala	Gly	Gly	Gly	Ala	Arg	Glu	Gly	Ala						
20	465					470					475						

(2) INFORMATION FOR SEQ ID NO:7:

25

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1622 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: both
- (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA
- 35 (ix) FEATURE:
  - (A) NAME/KEY: CDS
  - (B) LOCATION: 51..1283

# (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

	CAT	CAGC	CCA	CCAG	GAGA	CC T	CGCC	CGCC	G CT	cccc	CGGG	CTC	CCCG	GCC	ATG	TCT	56
															Met	Ser	
5															1		-
	CCC	GCC	CGG	CTC	CGG	CCC	CGA	CTG	CAC	TTC	TGC	CTG	GTC	CTG	TTG	CTG	104
	Pro	Ala	Arg	Leu	Arg	Pro	Arg	Leu	His	Phe	Cys	Leu	Val	Leu	Leu	Leu	
			5					10					15				
10																	
	CTG	CTG	GTG	GTG	CCC	GCG	GCA	TGG	GGC	TGC	GGG	CCG	GGT	CGG	GTG	GTG	152
	Leu	Leu	Val	Val	Pro	Ala	Ala	Trp	Gly	Cys	Gly	Pro	Gly	Arg	Val	Val	
		20					25					30					
15									AAA								200
	Gly	Ser	Arg	Arg	Arg		Pro	Arg	Lys	Leu		Pro	Leu	Ala	Tyr		
	35					40					45					50	
											0.00	200	200	3.00	667	200	240
									AAG								248
20	Gln	Phe	Ser	Pro		Val	Pro	GLu	Lys		Leu	стА	A.la	ser	65	Arg	
					55					60					65		
	m n m	C 2 2	000	N F C	z m.c	CCIR	ccc	N.C.C	TCC	CAC	CCC	ጥጥር	NAC.	CAC	ርሞር	ACC.	296
									Ser								250
25	Tyr	Glu	GIÀ		TTE	ATA	Arg	ser	75	GIU	Arg	rne	гуз	80	Беа	1111	
25				70					,,					00			
	CCC	<b>አ</b> አጥ	<b>ም</b> እር	አልጥ	CCA	GDC	ΔͲC	ΑΤС	TTC	AAG	GAC	GAG	GAG	AAC	ACA	GGC	344
									Phe								
	110	AJII	85	ASII	110	1101	110	90		2,0			95			1	
30			0.5					-					• -				
50	GCC	GAC	CGC	СТС	ATG	ACC	CAG	CGC	TGC	AAG	GAC	CGC	CTG	AAC	TCG	CTG	392
									Cys								
		100	•••	200			105	5		-,-		110					
35	GCT	ATC	TCG	GTG	ATG	AAC	CAG	TGG	ccc	GGT	GTG	AAG	CTG	CGG	GTG	ACC	440
									Pro								
	115					120		•		•	125	-		-		130	
	GAG	GGC	TGG	GAC	GAG	GAC	GGC	CAC	CAC	TCA	GAG	GAG	TCC	CTG	CAT	TAT	488

	Glu	Gly	Trp	Aşp	Glu	Asp	Gly	His	His	Ser	Glu	Glu	Ser	Leu	His	Tyr	
					135					140					145		
	GNG	ccc	CGC	GCG	GTG	GAC	ልሞር	ACC	ACA	ጥርል	GAC	CGC	GAC	CGC	ТАА	AAG	536
5			Arq														_
J	OI G	O <sub>1</sub>	9	150		1.01			155	-	p	• 9		160		-1 -	
	TAT	GGA	CTG	CTG	GCG	CGC	TTG	GCA	GTG	GAG	GCC	GGC	TTT	GAC	TGG	GTG	584
	Tyr	Gly	Leu	Leu	Ala	Arg	Leu	Ala	Val	Glu	Ala	Gly	Fhe	Asp	Trp	Val	
10			165					170					175				
	ጥልጥ	<b>ጥ</b> ስር	GAG	<b>ጥ</b> ር Δ	AAG	GCC	CAC	GTG	СДТ	тсс	ሞርር	GTC	AAG	TCC	GAG	CAC	632
			Glu														
	-3-	180			4		185			•		190	-				
15																	
	TCG	GCC	GCA	GCC	AAG	ACG	GGC	GGC	TGC	TTC	CCT	GCC	GGA	GCC	CAG	GTA	680
	Ser	Ala	Ala	Ala	Lys	Thr	Gly	Gly	Cys	Phe	Pro	Ala	Gly	Ala	Gln	Val	
	195					200					205					210	
20	000	0.00	67.6	».cm	ccc	ccc	CCM	CMC	ccc	mm/c	TO A	ccc	CTC	אככ	ccc	CCA	728
20			GAG Glu														120
	Arg	ъeu	GIU	Ser	215	MIG	Arg	Val	nia	220	Jer	VIO	V 0 1	nr g	225	Cly	
	GAC	CGT	GTG	CTG	GCC	ATG	GGG	GAG	GAT	GGG	AGC	CCC	ACC	TTC	AGC	GAT	776
25	Asp	Arg	Val	Leu	Ala	Met	Gly	Glu	Asp	Gly	Ser	Pro	Thr	Phe	Ser	Asp	
				230					235					240			
	omo.	ama.	3.00	mmc.	ome.	CNC	000	CAC	ccc	CAC	ክ <i>ር</i> ር	CTTC	A C A	ccc	ጥጥር	CAC	824
			ATT Ile														024
30	vaı	ьеи	245	FIIE	Leu	изр	Arg	250	110	1113	ni y	LCu	255	1114	1110	02	
50			2.10					200									
	GTC	ATC	GAG	ACT	CAG	GAC	ccc	CCA	CGC	CGC	CTG	GCA	CTC	ACA	CCC	GCT	872
	Val	Ile	Glu	Thr	Gln	Asp	Pro	Pro	Arg	Arg	Leu	Ala	Leu	Thr	Pro	Ala	
		260					265					270					
35																	
			CTC														920
		Leu	Leu	Phe	Thr		qzA	Asn	His	Thr		Pro	Ala	Ala	Arg		
	275					280					285					290	

	CGG	GCC	ACA	TTT	GCC	AGC	CAC	GTG	CAG	CCT	GGC	CAG	TAC	GTG	CTG	GTG	968
	Arg	Ala	Thr	Phe		Ser	His	Val	Gln		_	Gln	Tyr	Val		Val	
					295					300					305		
5	GCT	GGG	GTG	CCA	GGC	CTG	CAG	CCT	GCC	CGC	GTG	GCA	GCT	GTC	TCT	ACA	10 <del>16</del>
	Ala	Gly	Val	Pro	Gly	Leu	Gln	Pro	Ala	Arg	Val	Ala	Ala	Val	Ser	Thr	
				310					315					320			
	CAC	GTG	GCC	CTC	GGG	GCC	TAC	GCC	CCG	CTC	ACA	AAG	CAT	GGG	ACA	CTG	1064
10	His	Val	Ala	Leu	Gly	Ala	Tyr	Ala	Pro	Leu	Thr	Lys	His	Gly	Thr	Leu	
			325					330					335				
	GTG	GTG	GAG	GAT	GTG	GTG	GCA	TCC	TGC	TTC	GCG	GCC	GTG	GCT	GAC	CAC	1112
	Val	Val	Glu	Asp	Val	Val	Ala	Ser	Cys	Phe	Ala	Ala	Val	Ala	Asp	His	
15		340					345					350					
	CAC	CTG	GCT	CAG	TTG	GCC	TTC	TGG	CCC	CTG	AGA	CTC	TTT	CAC	AGC	TTG	1160
	His	Leu	Ala	Gln	Leu	Ala	Phe	Trp	Pro	Leu	Arg	Leu	Phe	His	Ser.	Leu	
	355					360					365					370	
20																	
						ACC											1208
	Ala	Trp	Gly	Ser	•	Thr	Pro	Gly	Giu	380 GTÀ	Val	His	Trp	Tyr	385	GIn	
					375					300					303		
25	CTG	CTC	TAC	CGC	CTG	GGG	CGT	СТС	CTG	СТА	GAA	GAG	GGC	AGC	TTC	CAC	1256
	Leu	Leu	Tyr	Arg	Leu	Gly	Arg	Leu	Leu	Leu	Glu	Glu	Gly	Ser	Phe	His	
				390					395					400			
	CCA	CTG	GGC	ATG	TCC	GGG	GCA	GGG	AGC	TGAA	AGGA	CT C	CACC	GCTG	C		1303
30	Pro	Leu	Gly	Met	Ser	Gly	Ala	Gly	Ser								
			405					410									
	CCTC	CTGG	SAA C	TGCT	GTAC	T GG	GTCC	AGAA	GCC	TCTC	AGC	CAGG	AGGG	AG C	TGGC	CCTGG	1363
															~~~	C1 CC1	1.400
35	AAGG	GACC	TG A	.GCTG	GGGG	A CA	CTGG	CTCC	TGC	CATC	TCC	TUTE	CCAT	са А	GATA	CACCA	1423
	TTGA	GACT	TG A	CTGG	GCAA	C AC	CAGC	GTCC	CCC	ACCC	GCG	TCGT	GGTG	TA G	TCAT	AGAGC	1483
	TGCA	AGCT	GA G	CTGG	CGAG	G GG	ATGG	TTGT	TGA	CCCC	TCT	CTCC	TAGA	GA C	CTTG	AGGCT	1543
					_							_					

	GGC	ACGG	CGA	CTCC	CAAC	TC A	GCCT	GCTC	r ca	CTAC	GAGT	TTT	CATA	CTC	TGCC	TCCCC	CC	1603
_	ATT	GGGA	GGG	CCCA	TTCC	С												1622
5	(2)	INF	ORMA	TION	FOR	SEQ	ID	NO:8:	:									
10		(i	()	A) L B) T	ENGT YPE: TRAN	H: 1 nuc DEDN	191 leic ESS:	ISTIC base acic both ear	pai	rs								
15		(ii	) MO	LECU	LE T	YPE:	cDN	A										
		(ix	(2	ATUR A) N B) L	AME/													
20																		
		(xi	) SE	QUEN	CE D	ESCR	IPTI	ON: S	EQ	ID N	0:8:							
25	Met				Thr			CTG Leu		Leu					Leu			48
	1				5					10					15			
								GGG Gly										96
30				20					25					30				
								CTC Leu 40										144
35	GTG	CCC	GGC	GTG	CCA	GAG	CGG	ACC	CTG	GGC	GCC	AGT	GGG	CCA	GCG	GAG		192
								Thr										

	GGG	AGG	GTG	GCA	AGG	GGC	TCC	GAG	CGC	TTC	CGG	GAÇ	CTC	GTG	CCC	AAC	240
	Gly	Arg	Val	Ala	Arg	Gly	Ser	Glu	Arg	Phe	Arg	Asp	Leu	Val	Pro	Asn	
	65					70					75					80	
5	መአሮ	አአሮ	ccc	GAC	ልጥሮ	<u>አ</u> ጥሮ	ሞጥር	AAC	CAT	CAC	CNC	እ አ ር	аст	GGA	ecc	GAC	28 <del>8</del>
J				Asp													200
	TYL	ASII	FIU	nap	85	116	1110	БуЗ	пор	90		ASII	Ser	O.L.y	95	7.55	
	ccc	CTC	አመር	ACC	CNC	ССТ	TCC	N A C	CAC	n.c.c	CTC	አልሮ	CCT	TTC	ccc	<b>አ</b> ጥጥ	336
10				Thr													330
10	Arg	пец	мес	100	Gru	лгу	Суз	цуз	105	ALG	Val	ASII	A1.G	110	AIG	110	
				100					103					110			
	GCC	GTG	ATG	AAC	ATG	TGG	CCC	GGA	GTG	CGC	CTA	CGA	GTG	ACT	GAG	GGC	384
	Ala	Val	Met	Asn	Met	Trp	Pro	Gly	Val	Arg	Leu	Arg	Val	Thr	Glu	Gly	
15			115					120					125				
	TGG	GAC	GAG	GAC	GGC	CAC	CAC	GCT	CAG	GAT	TCA	CTC	CAC	TΛC	GAA	GGC	432
				Asp													
		130			- 1		135			•		140		-		•	
20																	
	CGT	GCT	TTG	GAC	ATC	ACT	ACG	TCT	GAC	CGC	GAC	CGC	AAC	AAG	TAT	GGG	480
	Arg	Ala	Leu	Asp	Ile	Thr	Thr	Ser	Asp	Arg	Asp	Arg	Asn	ъуs	Tyr	Gly	
	145					150					155					160	
25				CGC													528
	Leu	Leu	Ala	Arg		Ala	Val	Glu	Ala		Phe	Asp	Trp	Val		Tyr	
					165					170					175		
	GAG	TCC	CGC	AAC	CAC	GTC	CAC	GTG	TCG	GTC	AAA	GCT	GAT	AAC	TCA	CTG	576
30	Glu	Ser	Arg	Asn	His	Val	His	Val	Ser	Val	Lys	Ala	Asp	Asn	Ser	Leu	
				180					185					190			
	GCG	GTC	CGG	GCG	GGC	GGC	TGC	TTT	CCG	GGA	AAT	GCA	ACT	GTG	CGC	CTG	624
				Ala													
35			195		4	•	•	200		•			205				
	TGG	AGC	GGC	GAG	CGG	AAA	GGG	CTG	CGG	GAA	CTG	CAC	CGC	GGA	GAC	TGG	672
	Trp	Ser	Gly	Glu	Arg	Lys	Gly	Leu	Arg	Glu	Leu	His	Arg	Gly	Asp	Trp	
		210					215					220					

				•	•											-		
	GTT	TTG	GCG	GCC	GAT	GCG	TCA	GGC	CGG	GTG	GTG	CCC	ACG	CCG	GTG	CTG	720	)
	Val	Leu	Ala	Ala	Asp	Ala	Ser	Gly	Arg	Val	Val	Pro	Thr	Pro	Val	Leu		
	225					230		•			235					240		
5																		-
	CTC	TTC	CTG	GAC	CGG	GAC	TTG	CAG	CGC	CGG	GCT	TCA	TTT	GTG	GCT	GTG	768	}
	Leu	Phe	Leu	Asp	Arg	Asp	Leu	Gln	Arg	Arg	Ala	Ser	Phe	Val	Ala	Val		
					245					250					255			
10				TGG													816	,
	Glu	Thr	Glu	Trp	Pro	Pro	Arg	Lys	Leu	Leu	Leu	Thr	Pro	Trp	His	Leu		
				260					265					270				
																~~~	0.54	
				GCT													864	
15	Val	Phe		Ala	Arg	Gly	Pro		Pro	Ala	Pro	GLY		Phe	Ата	Pro		
			275					280					285					
	CTC	שיייר	ccc	CGC	CCC	ርሞአ	CCC	ርርሞ	GGG	GAC	тсс	GTG	CmC	GCG	ccc	GGC	912	ı
				Arg													7.2	
20	vaı	290	ита	Arg	Ary	₽₩u	295	nia	G., 7	nop	OCI	300	Deu	11.20		CLY		
20		230					2,5											
	GGG	GAT	GCG	CTT	CGG	CCA	GCG	CGC	GTG	GCC	CGT	GTG	GCG	CGG	GAG	GAA	960	ļ
				Leu														
	305	_				310					315					320		
25																		
	GCC	GTG	GGC	GTG	TTC	GCG	CCG	CTC	ACC	GCG	CAC	GGG	ACG	CTG	CTG	GTG	1008	
	Ala	Val	Gly	Val	Phe	Ala	Pro	Leu	Thr	Ala	His	Gly	Thr	Leu	Leu	Val		
					325					330					335			
30	AAC	GAT	GTC	CTG	GCC	TCT	TGC	TAC	GCG	GTT	CTG	GAG	AGT	CAC	CAG	TGG	1056	
	Asn	Asp	Val	Leu	Ala	Ser	Cys	Tyr	Ala	Val	Leu	Glu	Ser	His	Gln	Trp		
				340					345					350				
				GCT													1104	
35	Ala	His		Ala	Phe	Ala	Pro		Arg	Leu	Leu	His		Leu	Gly	Ala		
			355					360					365					
									000		000	* Tr C	<b>~</b> ~ ~ ~	mcc	m » ~	m c m	1150	
				GGC													1152	
	Leu	Leu	Pro	Gly	СТÄ	Ala	val	GIN	Pro	Inr	GTÀ	met	nlS	rrp	ryr	ser		

380 370 375 CGG CTC CTC TAC CGC TTA GCG GAG GAG CTA CTG GGC TG 1191 Arg Leu Leu Tyr Arg Leu Ala Glu Glu Leu Leu Gly 385 390 395 (2) INFORMATION FOR SEQ ID NO:9: 10 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 1251 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: both 15 (D) TOPOLOGY: linear (ii) MOLECULE TYPE: cDNA 20 (ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1..1248 25 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9: ATG GAC GTA AGG CTG CAT CTG AAG CAA TTT GCT TTA CTG TGT TTT ATC 48 Met Asp Val Arg Leu His Leu Lys Gln Phe Ala Leu Leu Cys Phe Ile 5 10 15 1 30 AGC TTG CTT CTG ACG CCT TGT GGA TTA GCC TGT GGT CCT GGT AGA GGT 96 Ser Leu Leu Thr Pro Cys Gly Leu Ala Cys Gly Pro Gly Arg Gly 20 25 30 TAT GGA AAA CGA AGA CAC CCA AAG AAA TTA ACC CCG TTG GCT TAC AAG 144 Tyr Gly Lys Arg Arg His Pro Lys Lys Leu Thr Pro Leu Ala Tyr Lys 40 45 35

192

CAA TTC ATC CCC AAC GTT GCT GAG AAA ACG CTT GGA GCC AGC GGC AAA

	Gln	Phe	Ile	Pro	Asn	Val	Ala	Glu	Lys	Thr	Leu	Gly	Ala	Ser	Gly	Lys	
		50					55					60	I				
			000		3.00	• • • • • • • • • • • • • • • • • • • •	* ~ ~										
•			GGC														24(
5	1 y r 6 5		Gly	гуѕ	116	70	_	ASII	ser	GIU	75		гÀг	GIU	Leu	80	
	0.5					, 0					,,	'				00	
	CCG	AAT	TAT	AAT	ccc	GAT	ATC	ATC	TTT	AAG	GAC	GAG	GAA	AAC	ACA	AAC	288
	Pro	Asn	Tyr	Asn	Pro	Asp	Ile	Ile	Phe	Lys	Asp	Glu	Glu	Asn	Thr	Asn	
10					85					90					95		
														,			
			AGG -														336
	Ala	Asp	Arg		Met	Thr	Lys	Arg	-	Lys	Asp	Lys	Leu		Ser	Leu	
15				100					105					110			
15	GCC	ATA	TCC	GTC	ATG	AAC	CAC	TGG	CCC	GGC	GTG	AAA	CTG	CGC	GTC	ACT	384
			Ser														
			115					120					125				
											i						
20	GAA	GGC	TGG	GAT	GAG	GAT	GGT	CAC	CAT	TTA	GAA	GAA	TCT	TTG	CAC	TAT	432
	Glu	_	Trp	Asp	Glu	Asp		His	His	Leu	Glu		Ser	Leu	His	Tyr	
		130					135					140					
	GAG	GGA	CGG	GCA	GTG	GAC	ATC	АСТ	ACC	тса	GAC	AGG	САТ	АДА	AGC	AAG	480
25			Arg														
	145	-	,			150					155	•	•	-		160	
	TAT	GGG	ATG	CTA	TCC	AGG	CTT	GCA	GTG	GAG	GCA	GGA	TTC	GAC	TGG	GTC	528
	Tyr	Gly	Met	Leu	Ser	Arg	Leu	Ala	Val	Glu	Ala	Gly	Phe	Asp	Trp	Val	
30					165					170					175		
	m a m	m n m	CDD	m.cm	***	ccc	CAC	አጠአ	CNC	TICC.	тст	CMC	222	CCA	CNN	አአጥ	576
			GAA Glu														576
	TYT	ıyı	GIU	180	цуs	ліа	1113	116	185	Cys	Jei	val	БуЗ	190	GIU	ASII	
35																	
	TCA	GTG	GCT	GCT	AAA	TCA	GGA	GGA	TGT	TTT	CCT	GGG	TCT	GGG	ACG	GTG	624
	Ser	Val	Ala	Ala	Lys	Ser	Gly	Gly	Cys	Phe	Pro	Gly	Ser	Gly	Thr	Val	
			195					200					205				

	ACA	CTT	GGT	GAT	GGG	ACG	AGG	AAA	CCC	ATC	AAA	GAT	CTT	AAA	GTG	GGC	672
	Thr	Leu	Gly	Asp	Gly	Thr	Arg	Lys	Pro	Ile	Lys	Asp	Leu	Lys	Val	Gly	
		210					215					220					
5	GAC	CGG	GTT	TTG	GCT	GCA	GAC	GAG	AAG	GGA	AAT	GTC	TTA	ATA	AGC	GAC	720
	Asp	Arg	Val	Leu	Ala	Ala	Asp	Glu	Lys	Gly	Asn	Val	Leu	Ile	Ser	Asp	
	225					230					235					240	
												AGA					768
10	Phe	Ile	Met	Phe	Ile	qzA	His	Asp	Pro	Thr	Thr	Arg	Arg	Gln	Phe	Ile	
					245					250					255		
												ACC					816
	Val	Ile	Glu	Thr	Ser	Glu	Pro	Phe		Lys	Leu	Thr	Leu		Ala	Ala	
15				260					265					270			
																	0.54
												TCG					864
	His	Leu		Phe	Val	Gly	Astı		Ser	Ala	Ala	Ser		11.e	Thr	Ala	•
			275					280					285				
20								041	aan	C 7 T	n C n	C TO IT	mma	CTC	TCC	CNN	912
												GTT					912
	Thr		Ala	Ser	Asn	Va.L		Pro	GIA	Asp	TRE	Val	ьeu	Val	пр	GIU	
		290					295					300					
25	CAC	מרמ	ጥርር	GAG	AGC	CTC	AAG	agc.	GTT	ACA	GTG	AAA	AGG	ATT	TAC	ACT	960
23												Lys					
	305	1111	Суз	GIU	501	310	2,5	001			315		3		-	320	
	303					510											
	GAG	GAG	CAC	GAG	GGC	TCT	ттт	GCG	CCA	GTC	ACC	GCG	CAC	GGA	ACC	ATA	1008
30												Ala					
•					325					330					335		
	ATA	GTG	GAT	CAG	GTG	TTG	GCA	TCG	TGC	TAC	GCG	GTC	ATT	GAG	AAC	CAC	1056
												Val					
35				340					345					350			
	AAA	TGG	GCA	CAT	TGG	GCT	TTT	GCG	CCG	GTC	AGG	TTG	TGT	CAC	AAG	CTG	1104
	Lys	Trp	Ala	His	Trp	Ala	Phe	Ala	Pro	Val	Arg	Leu	Cys	His	Lys	Leu	
			355					360					365				

				٠.	٠,													
	ATG	ACG	TGG	CTT	TTT	CCG	GCT	CGT	GAA	TCA	AAC	GTC	AAT	TTT	CAG	GAG	1	152
	Met	Thr	Trp	Leu	Phe	Pro	Ala	Arg	Glu	Ser	Asn	Val	Asn	Phe	Gln	Glu		
		370					375					380						
5																	_	
					TGG												1	200
	-	Gly	Ile	His	Trp		Ser	Asn	Met	Leu		HIS	TTE	GIÀ	Ser	400		
	385					390					395					400		
10	CTG	CTG	GAC	AGA	GAC	TCT	TTC	CAT	CCA	CTC	GGG	ATT	TTA	CAC	TTA	AGT	1	248
	Leu	Leu	Asp	Arg	Asp	Ser	Phe	His	Pro	Leu	Gly	Ile	Leu	His	Leu	Ser		
					405					410					415			
	TGA																1:	251
15																		
	,			n T C.)]	TOD	aro.	TD 1	30.1	٦.									
	(2)	INF	JRMA'	LTOM	FOR	SEQ	10 1	NO: 10	<i>.</i>									
			(i) 3	SEOUI	ENCE	CHA	RACTI	ERIS:	rics:	:								
20		٠		(A)	LEN	NGTH:	42	ā am:	ino a	cids	5							
				(B)	'TYI	PE: 6	amino	o aci	id									
				(D)	тог	OLOC	SY: 3	linea	ar									
		(:	ii) N	MOLE	CULE	TYPE	E: pi	rote	in									
25																		
		,		FOU	ENCE	חודפנ	ים דמי	י א רי די	SEC	מד כ	NO · 1	10:						
		(2	M.I.) .	POOT	JIVOD				,									
	Met	Val	Glu	Met	Leu	Leu	Leu	Thr	Arg	Ile	Leu	Leu	Val	Gly	Phe	Ile		
30	1				5					10					15			
	Cys	Ala	Leu	Leu	Val	Ser	Ser	Gly	Leu	Thr	Cys	Gly	Pro	Gly	Arg	Gly		
				20					25					30				
												_	_		_	_		
35	Ile	Gly		Arg	Arg	His	Pro		Lys	Leu	Thr	Pro		Ala	Tyr	Lys		
			35					40					45					
	C1-	Dh -	T1-	Dro	Asn	V = 1	Δls	Glu	T.ve	<b>ጥ</b> ኮ ፦	Leu	Glv	Ala	Ser	Glv	Arg		
	GID	50	11e	PTO	ASII	Val	55	JIU	دوب	1117	±eu.	60			3	9		
		50																

30

	Туг 65	Glu	Gly	Lys	Ile	Thr 70	Arg	Asn	Ser	Glu	Arg 75	Phe	Lys	Glu	Leu	Thr 80
5	Pro	Asn	Tyr	Asn	Pro 85	Asp	Ile	Ile	Phe	Lys 90	Asp	Glu	Glu	Asn	Thr 95	Gly
10	Ala	Asp	Arg	Leu 100	Met	Thr	Gln	Arg	Cys 105	Lys	Asp	Ŀys	Leu	Asn 110	Ala	Leu
	Ala	Ile	Ser 115	Val	Met	Asn	Gln	Trp 120	Pro	Gly	Val	Lys	Leu 125	Arg	Val	Thr
15	Glu	Gly 130	Trp	Asp	Glu	Asp	Gly 135	His	His	Ser	Glu	Glu 140	Ser	Leu	His	Tyr
٠	Glu 145	Gly	Arg	Ala	Val	Asp 150	Ile	Thr	Thr	Ser	Asp 155	Arg	Asp	Arg	Ser	Lys 160
20	Туг	Gly	Met	Leu	Ala 165	Arg	Leu	Ala	Val	Glu 170	Ala	Gly	Phe	Asp	Trp 175	Val
25	Tyr	Tyr	Glu	Ser 180	Lys	Ala	His	Ile	His 185	Cys	Ser	Val	Lys	Ala 190	Glu	Asn
23	Ser	Val	Ala 195	Ala	Lys	Ser	Gly	Gly 200	Cys	Phe	F.ro	Gly	Ser 205	Ala	Thr	Val
30	His	Leu 210	Glu	His	Gly	Gly	Thr 215	Lys	Leu	Val	Lys	Asp 220	Leu	Ser	Pro	Gly
	Asp 225	Arg	Val	Leu	Ala	Ala 230	Asp	Ala	Asp	Gly	Arg 235	Leu	Leu	Tyr	Ser	Asp 240
35	Phe	Leu	Thr	Phe	Leu 245	Asp	Arg	Met	Asp	Ser 250	Ser	Arg	Lys	Leu	Phe 255	Tyr
	Val	Ile	Glu	Thr 260	Arg	Gln	Pro	Arg	265	Arg	Leu	Leu	Leu	Thr 270	Ala	Ala

	His	Leu	Leu	Phe	Val	Ala	Pro	Gln	His	Asn	Gln	Ser	Glu	Ala	Thr	Gly
			275					280					285			
5	Ser	Thr	Ser	Gly	Gln	Ala		Phe	Ala	Ser	Asn		Lys	Pro	Gly	Gln
		290					295					300				
	Arg	Val	Tyr	Val	Leu		Glu	Gly	Gly	Gln		Leu	Leu	Pro	Ala	Ser 320
10	305					310					315					J20
	Val	His	Ser	Val		Leu	Arg	Glu	Glu	Ala 330	Ser	Gly	Ala	Tyr	Ala 335	Pro
					325											
15	Leu	Thr	Ala	Gln 340	Gly	Thr	Ile	Leu	Ile 345	Asn	Arg	Val	Leu	Ala 350	Ser	Cys
13															_,	~
	Tyr	Ala	Val 355	Ile	Glu	Glu	His	Ser 360	Trp	Ala	His	Trp	Al.a 365	Phe	Ala	Pro
					01	<b>61</b>	7	T 0.11	7 l -	71-	Lou	Cue	Pro	Asn	Gly	Ala
20	Phe	Arg 370	Leu	Ala	GIN	GIÀ	375	ьеи	MIG	MIG	neu	380	110	nop	J.,	
	Tlo	Pro	Thr	Δla	Δla	Thr	Thr	Thr	Thr	Glv	Ile	His	Trp	Tyr	Ser	Arg
	385	FIO	1111	VIO	діа	390				2	395		•	-		400
25	Leu	Leu	Tyr	Ara	Ile	Gly	Ser	Trp	Val	·Leu	Asp	Gly	Asp	Ala	Leu	His
	200		- <b>,</b> -	,	405	-				410					415	
	Pro	Leu	Gly	Met	Val	Ala	Pro	Ala	Ser							
30				420					425							
	(2)	INF	ORMA'	TION	FOR	SEQ	ID :	NO:1	1:							
35			(i)													
					•				ino id		S					
				( )	1 TY	rr:	aiiiLll	u ac	<u>. u</u>							

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

5		(>	ri) S	EQUE	ENCE	DESC	CRIPT	TION:	SEÇ	) ID	NO:1	11:				
	Met 1	Ala	Leu	Pro	Ala 5	Ser	Leu	Leu	Pro	Leu 10	Cys	Cys	Leu	Ala	Leu 15	Leu
10	Ala	Leu	Ser	Ala 20	Gln	Ser	Cys	Gly	Pro 25	Gly	Arg	Gly	Pro	Val 30	Gly	Arg
	Arg	Arg	Tyr 35	Val	Arg	Lys	Gln	Leu 40	Val	Pro	Leu	Leu	Tyr 45	Lys	Gln	Phe
15	Val	Pro 50	Ser	Met	Pro	Glu	Arg 55	Thr	Leu	Gly	Ala	Ser 60	Gly	Pro	Ala	Glu
20	Gìy 65	Arg	Val	Thr	Arg	Gly 70	Ser	Glu	Arg	Phe	Arg 75	Asp	Leu	Val	Pro	Asn 80
	Туr	Asn	Pro	Asp	Ile 85	lle	Phe	Lys	Asp	Glu 90	Glu	Asn	Ser	Gl y	Ala 95	Asp
25	Arg	Leu	Met	Thr 100	Glu	Arg	Cys	Lys	Glu 105	Arg	Val	Asn	Ala	նeu 110	Ala	Ile
	Ala	Val	Met 115	Asn	Met	Trp	Pro	Gly 120	Val	Arg	Leu	Arg	Val 125	Thr	Glu	Gly
30	Trp	Asp 130	Glu	Asp	Gly	His	His 135	Ala	Gln	Asp	Ser	Leu 140	His	Tyr	Glu	Gly
35	Arg 145	Ala	Leu	Asp	Ile	Thr 150	Thr	Ser	Asp	Arg	Asp 155	Arg	Asn	Lys	Tyr	Gly 160
	Leu	Leu	Ala	Arg	Leu 165	Ala	Val	Glu	Ala	Gly 170	Phe	Asp	Trp	Val	Tyr 175	Tyr

	Glu	Ser	Arg	Asn 180	His	Ile	His	Val	Ser 185	Val	Lys	Ala	Asp	Asn 190	Ser	Leu
5	Ala	Val	Arg 195	Ala	Gly	Gly	Cys	Phe 200	Pro	Gly	Asn	Ala	Thr 205	Val	Arg	Leu
	Arg	Ser 210	Gly	Glu	Arg	Lys	Gly 215	Leu	Arg	Glu	Leu	His 220	Arg	Gly	Asp	Trp
10	Val 225	Leu	Ala	Ala	Asp	Ala 230	Ala	Gly	Arg	Val	Val 235	Pro	Thr	Pro	Val	Leu 240
15	Leu	Phe	Leu	Asp	Arg 245	Asp	Leu	Gln	Arg	Arg 250	Ala	Ser	Phe	Val	Ala 255	Val
	Glu	Thr	Glu	Arg 260	Pro	Pro	Arg	Lys	Leu 265	Ŀeu	Leu	Thr	Pro	Trp 270	His	Leu
20	Val	Phe	Ala 275	Al.a	Arg	Gly	Pro	Ala 280	Pro	Ala	Pro	Gly	Asp 285	Phe	Ala	Pro
	Val	Phe 290	Ala	Arg	Arg	Leu	Arg 295	Ala	Gly	Asp	Ser	Val 300	Leu	Ala	Pro	Gly
25	Gly 305	Asp	Ala	Leu	Gln	Pro 310	Ala	Arg	Val	Ala	Arg 315	Val	Ala	Arg	Glu	Glu 320
30	Ala	Val	Gly	Val	Phe 325	Ala	Pro	Leu	Thr	Ala 330	His	Gly	Thr	Leu	Leu 335	Val
	Asn	Asp	Val	Leu 340	Ala	Ser	Cys	Tyr	Ala 345	Val	Leu	Glu	Ser	His 350	Gln	Trp
35	Ala	His	Arg 355	Ala	Phe	Ala	Pro	Leu 360	Arg	Leu	Leu	His	Ala 365	Leu	Gly	Ala
	Leu	Leu 370	Pro	Gly	Gly	Ala	Val 375	Gln	Pro	Thr	Gly	Met 380	His	Trp	Tyr	Ser

Arg Leu Leu Tyr Arg Leu Ala Glu Glu Leu Met Gly
385 390 395

- 5 (2) INFORMATION FOR SEQ ID NO:12:
  - (i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 411 amino acids

(B) TYPE: amino acid

10 (D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

15 (xi) SEQUENCE DESCRIPTION: SEQ ID NO:12:

Met Ser Pro Ala Trp Leu Arg Pro Arg Leu Arg Phe Cys Leu Phe Leu

1 5 10 15

20

35

Leu Leu Leu Leu Val Pro Ala Ala Arg Gly Cys Gly Pro Gly Arg
20 25 30

Val Val Gly Ser Arg Arg Pro Pro Arg Lys Leu Val Pro Leu Ala . 25 40 45

Tyr Lys Gln Phe Ser Pro Asn Val Pro Glu Lys Thr Leu Gly Ala Ser 50 60

30 Gly Arg Tyr Glu Gly Lys Ile Ala Arg Ser Ser Glu Arg Phe Lys Glu 65 70 75 80

Leu Thr Pro Asn Tyr Asn Pro Asp Ile Ile Phe Lys Asp Glu Glu Asn 85 " 90 95

Thr Gly Ala Asp Arg Leu Met Thr Gln Arg Cys Lys Asp Arg Leu Asn 100 105 110

Ser Leu Ala Ile Ser Val Met Asn Gln Trp Pro Gly Val Lys Leu Arg

			115					120					125			
5	Val	Thr 130	Glu	Gly	Arg	Asp	Glu 135	Asp	Gly	His	His	Ser 140	Glu	Glu	Ser	Leu
J	His 145	Tyr	Glu	Gly	Arg	Ala 150	Val	Asp	Ile	Thr	Thr 155	Ser	Asp	Arg	Asp	Arg 160
10	Asn	Lys	Tyr	Gly	Leu 165	Leu	Ala	Arg	Leu	Ala 170	Val	Glu	Ala	Gly	Phe 175	Asp
	Trp	Val	Tyr	Tyr 180	Glu	Ser	Lys	Ala	His 185	Val	His	Cys	Ser	Val 190	Lys	Ser
15	Glu	His	Ser 195	Ala	Ala	Ala	Lys	Thr 200	Gly	Gly	Cys	Phe	Pro 205	Ala	Gly	Ala
20	Gln	Val 210	Arg	Leu	Glu	Asn	Gl.y 215	GLu	Arg	Val	Ala	Leu 220	Ser	Ala	Val	Lys
	Pro 225	Gly	Asp	Arg	Val	Leu 230	Ala	Met	Gly	Glu	Asp 235	Gly	Thr	Pro	Thr	Phe 240
25	Ser	Asp	Val	Leu	Ile 245	Phe	Leu	Asp	Arg	Glu 250	Pro	Asn	Arg	Leu	Arg 255	Ala
	Phe	Gln	Val	Ile 260	Glu	Thr	Gln	Asp	Pro 265	Pro	Arg	Arg	Leu	Ala 270	Leu	Thr
30	Pro	Ala	His 275	Leu	Leu	Phe	Ile	Ala 280	Asp	Asn	His	Thr	Glu 285	Pro	Ala	Ala
35	His	Phe 290	Arg	Ala	Thr	Phe	Ala 295	Ser	His	Val	Gln	Pro 300	Gly	Gln	Tyr	Val
,,	Leu 305	Val	Ser	Gly	Val	Pro 310	Gly	Leu	Gln	Pro	Ala 315	Arg	Val	Ala	Ala	Val 320
	Ser	Thr	His	Val	Ala	Leu	Gly	Ser	Tyr	Ala	Pro	Leu	Thr	Arg	His	Gly

325 330 335

Thr Leu Val Val Glu Asp Val Val Ala Ser Cys Phe Ala Ala Val Ala 340 345 350

5

Asp His His Leu Ala Gln Leu Ala Phe Trp Pro Leu Arg Leu Phe Pro
355 360 365

Ser Leu Ala Trp Gly Ser Trp Thr Pro Ser Glu Gly Val His Ser Tyr 10 370 375 380

Pro Gln Met Leu Tyr Arg Leu Gly Arg Leu Leu Leu Glu Glu Ser Thr 385 390 395 400

15 Phe His Pro Leu Gly Met Ser Gly Ala Gly Ser 405 410

## 20 (2) INFORMATION FOR SEQ ID NO:13:

(1) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 437 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein

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(xi) SEQUENCE DESCRIPTION: SEQ ID NO:13:

Met Leu Leu Leu Leu Ala Arg Cys Phe Leu Val Ile Leu Ala Ser Ser 1 5 10 15

35

Leu Leu Val Cys Pro Gly Leu Ala Cys Gly Pro Gly Arg Gly Phe Gly
20 25 30

Lys Arg Arg His Pro Lys Lys Leu Thr Pro Leu Ala Tyr Lys Gln Phe

			35					40					45			
5	Ile	Pro		Val	Ala	Glu	Lys		Leu	Gly	Ala	Ser 60	_	Arg	Tyr	Glu
J	Gly 65	-	Ile	Thr	Arg	Asn 70		Glu	Arg	Phe	Lys 75	Glu	Leu	Thr	Pro	Asn 80
10	Tyr	Asn	Pro	Asp	Ile 85	Ile	Phe	Lys	Asp	Glu 90		Asn	Thr	Gly	Ala 95	
	Arg	Leu	Met	Thr 100		Arg	Cys	Lys	Asp 105	Lys	Leu	Asn	Ala	Leu 110	Ala	Ile
15	Ser	Val	Met 115	Asn	Gln	Trp	Pro	Gly 120	Val	Arg	Leu	Arg	Val 125	Thr	Glu	Gly
20	Trp	Asp 130	Giu	Asp	Gly	His	His 135	Ser	Glu	Glu	Ser	Leu 140	His	Tyr	Glu	Gly
	Arg 145	Ala	Val	Asp	Ile	Thr 150	Thr	Ser	Asp	Arg	Asp 155	Arg	Ser	Lys	Tyr	Gly 160
25	Met	Leu	Ala	Arg	Leu 165	Ala	Val	Glu	Ala	Gly 170	Phe	Asp	Trp	Val	Tyr 175	Tyr
	Glu	Ser	Lys	Ala 180	His	Ile	His	Cys	Ser 185	Val	Lys	Ala	Glu	Asn 190	Ser	Val
30	Ala	Ala	Lys 195	Ser	Gly	Gly	Cys	Phe 200	Pro	Gly	Ser	Ala	Thr 205	Val	His	Leu
35	Glu	Gln 210	Gly	Gly	Thr	Lys	Leu 215	Val	Lys	Asp	Leu	Arg 220	Pro	Gly	Asp	Arg
<i>JJ</i>	Val 225	Leu	Ala	Ala	Asp	Asp 230	Gln	Gly	Arg	Leu	Leu 235	Tyr	Ser	Asp	Phe	Leu 240
	Thr	Phe	Leu	Asp	Arg	Asp	Glu	Gly	Ala	Lys	Lys	Val	Phe	Tyr	Val	Ile

					245					250					255	
5	Glu	Thr	Leu	Glu 260	Pro	Arg	Glu	Arg	Leu 265	Leu	Leu	Thr	Ala	Ala 270	His	Leu
3	Leu	Phe	Val 275	Ala	Pro	His	Asn	Asp 280	Ser	Gly	Pro	Thr	Pro 285	Gly	Pro	Ser
10	Ala	Leu 290	Phe	Ala	Ser	Arg	Val 295	Arg	Pro	Gly	Gln	Arg 300	Val	Tyr	Val	Val
	Ala 305	Glu	Arg	Gly	Gly	Asp 310	Arg	Arg	Leu	Leu	Pro 315	Ala	Ala	Val	His	Ser 320
15	Val	Thr	Leu	Arg	G1u 325	Glu	Glu	Ala	Gly	Ala 330	Tyr	Дlа	Pro	Leu	Thr 335	Ala
20	His	Gly	Thr	Ile 340	Leu	Jie	Asn	Arg	Va.). 345	Ъeu	Ala	Ser	Cys	Tyr 350	Ala	Val
	Ile	Glu	Giu 355	His	Ser	Trp	Ala	His 360	Arg	Ala	Phe	Ala	Pro 365	Phe	Arg	Leu
25	Ala	His 370	Ala	Leu	Leu	Ala	Ala 375	Leu	Ala	Pro	Ala	Arg 380	Thr	Asp	Gly	Gìу
	Gly 385	Gly	Gly	Ser	Ile	Pro 390	Ala	Ala	Gln	Ser	Ala 395	Thr	Glu	Ala	Arg	Gly 400
30	Ala	Glu	Pro	Thr	Ala 405	Gly	Ile	His	Trp	Туг 410	Ser	Gln	Leu	Leu	Tyr 415	His
35	Ile	Gly	Thr	Trp 420	Leu	Leu	Asp	Ser	Glu 425	Thr	Met	His	Pro	Leu 430	Gly	Met
<i>J J</i>	Ala	Val	Lys	Ser	Ser											

PCT/US99/20500 WO 00/15246

(2) INFORMATION FOR SEQ ID NO:14:

(i) SEQUENCE CHARACTERISTICS:

(A) LENGTH: 418 amino acids

(B) TYPE: amino acid

(D) TOPOLOGY: linear

(ii) MOLECULE TYPE: protein .

10

5

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:14:

Met Arg Leu Leu Thr Arg Val Leu Leu Val Ser Leu Leu Thr Leu Ser 15 15

1

Leu Val Val Ser Gly Leu Ala Cys Gly Pro Gly Arg Gly Tyr Gly Arg 20

25

Arg Arg Ris Pro Lys Lys Leu Thr Pro Leu Ala Tyr Lys Gln Phe Ile 20 35 40.

Pro Asn Val Ala Glu Lys Thr Leu Gly Ala Ser Gly Arg Tyr Glu Gly 50 55

25

Lys Ile Thr Arg Asn Ser Glu Arg Phe Lys Glu Leu Thr Pro Asn Tyr 70 75

Asn Pro Asp Ile Ile Phe Lys Asp Glu Glu Asn Thr Gly Ala Asp Arg 30 90 85

Leu Met Thr Gln Arg Cys Lys Asp Lys Leu Asn Ser Leu Ala Ile Ser 105 100

Val Met Asn His Trp Pro Gly Val Lys Leu Arg Val Thr Glu Gly Trp 115 120 125

Asp Glu Asp Gly His His Phe Glu Glu Ser Leu His Tyr Glu Gly Arg 130 135 140

	Ala 145		Asp	lle	. Thr	Thr 150		Asp	Arg	Asp	155		Lys	Tyr	Gly	Thr 160
5	Leu	Ser	Arg	Leu	Ala 165		Glu	ı Ala	Gly	Phe 170	-	Trp	Val	Tyr	Туг 175	
10	Ser	Lys	Ala	His 180		His	Cys	Ser	Val 185	-	Ala	Glu	Asn	Ser 190		Ala
	Ala	Lys	Ser 195	_	Gly	Cys	Phe	Pro 200	Gly	Ser	Ala	Leu	Val 205	Ser	Leu	Gln
15	Asp	Gly 210	Gly	Gln	Lys	Ala	Val 215	-	Asp	Leu	Asn	Pro 220	Gly	Asp	Lys	Val
	Leu 225	Ala	Ala	Asp	Ser	Ala 230	Gly	Asn	Leu	Val	Phe 235		Asp	Phe	īle	Met 240
20	Ph.e	Thr	Asp	Arg	Asp 245	Ser	Thr	Thr	Arg	Arg 250	Val	Phe	туr	Va.l	Ile 255	Glu
25	Thr	Gln	Glu	Pro 260	Val	Glu	Lys	Ile	Thr 265	Leu	Thr	Ala	Ala	His 270	Leu	Leu
	Phe	Val	Leu 275	Asp	Asn	Ser	Thr	Glu 280	Asp	Leu	His	Thr	Met 285	Thr	Ala	Ala
30	Tyr	Ala 290	Ser	Ser	Val	Arg	Ala 295	Gly	Gln	Lys	Val	Met 300	Val	Val	Asp	Asp
	Ser 305	Gly	Gln	Leu	Lys	Ser 310	Val	Ile	Val	Gln	Arg 315	Ile	Tyr	Thr	Glu	Glu 320
35	Gln	Arg	Gly	Ser	Phe 325	Ala	Pro	Val	Thr	Ala 330	His	Gly	Thr	Ile	Val 335	Val
	Asp	Arg	Ile	Leu 340	Ala	Ser	Cys	-	Ala 345	Val	Ile	Glu	Asp	Gln 350	Gly	Leu

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Ala His Leu Ala Phe Ala Pro Ala Arg Leu Tyr Tyr Tyr Val Ser Ser Phe Leu Ser Pro Lys Thr Pro Ala Val Gly Pro Met Arg Leu Tyr Asn Arg Arg Gly Ser Thr Gly Thr Pro Gly Ser Cys His Gln Met Gly Thr Trp Leu Leu Asp Ser Asn Met Leu His Pro Leu Gly Met Ser Val Asn Ser Ser (2) INFORMATION FOR SEQ ID NO:15: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 475 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:15: Met Leu Leu Leu Ala Arg Cys Leu Leu Val Leu Val Ser Ser Leu Leu Val Cys Ser Gly Leu Ala Cys Gly Pro Gly Arg Gly Phe Gly Lys Arg Arg His Pro Lys Lys Leu Thr Pro Leu Ala Tyr Lys Gln Phe Ile

	Pro	Asn 50	Val	Ala	Glu	Lys	Thr 55	Leu	Gly	Ala	Ser	Gly 60	Arg	Tyr	Glu	Gly
5	Lys 65	Ile	Ser	Arg	Asn	Ser 70	Glu	Arg	Phe	Lys	Glu 75	Leu	Thr	Pro	Asn	Tyr 80
	Asn	Pro	Asp	Ile	Ile 85	Phe	Lys	Asp	Glu	Glu 90	Asn	Thr	Gly	Ala	<b>Asp</b> 95	Arg
10	Leu	Met	Thr	Gln 100	Arg	Cys	Lys	Asp	Lys 105	Leu	Asn	Ala	Leu	Ala 110	Ile	Ser
15	Val	Met	Asn 115	Gln	Trp	Pro	G3.y	Val 120	Lys	Leu	Arg	Val	Thr 125	Glu	Gly	Trp
	Asp	Glu 130	Asp	GГĀ	His	His	Ser 135	Glu	Glu	Ser	Leu	His 140	Tyr	Glu	Gly	Arg
20	Ala 145	Val	Asp	Ile	Thr	Thr 150	Ser	Asp	Arg	Asp	Arg 155	Ser	Lys	Tyr	Gly	Met 160
	Leu	Ala	Arg	Leu	Ala 165	Vai	Glu	Ala	Gly	Phe 170	Asp	Trp	Val	Tvr	Туг 175	Glu
25	Ser	Lys	Ala	His 180	Ile	His	Cys	Ser	Val 185	Lys	Ala	Glu	Asn	Ser 190	Val	Ala
30			195					200					205	His		
	Gln	Gly 210	Gly	Thr	Lys	Leu	Val 215	Lys	Asp	Leu	Ser	Pro 220	Gly	Asp	Arg	Val
35	Leu 225	Ala	Ala	Asp	Asp	Gln 230	Gly	Arg	Leu	Leu	Tyr 235	Ser	Asp	Phe	Leu	Thr 240
	Phe	Leu	Asp	Arg	Asp	Asp	Gly	Ala	Lys	Lys	Val	Phe	Tyr	Val	Ile 255	Glu

	Thr	Arg	Glu	Pro 260	Arg	Glu	Arg	Leu	Leu 265	Leu	Thr	Ala	Ala	His 270	Leu	Leu
5	Phe	Val	Ala 275	Pro	His	Asn	Asp	Ser 280	Ala	Thr	Gly	Glu	Pro 285	Glu	Ala	Ser
	Ser	Gly 290	Ser	Gly	Pro	Pro	Ser 295	Gly	Gly	Ala	Leu	Gly 300	Pro	Arg	Ala	Leu
10	Phe 305	Ala	Ser	Arg	Val	Arg 310	Pro	Gly	Gln	Arg	Val 315	Tyr	Val	Val	Ala	Glu 320
15	Arg	Asp	Gly	Asp	Arg 325	Arg	Leu	Leu	Pro	Ala 330	Ala	Val	His	Ser	Val 335	Thr
	Leu	Ser	Glu	Glu 340	Ala	Ala	Gly	Ala	Tyr 345	Ala	Pro	Leu	Thr	Ala 350	Gln	Gly
20	Thr	Ile	Leu 355	Ile	Asn	Arg	Val	Leu 360	Ala	Ser	Cys	Tyr	Ala 365	Val	Ile	G.lu
	Glu	His 370	Ser	Trp	Ala	His	Arg 375	Ala	Phe	Ala	Pro	P.he 380	Arg	Leu	Ala	His
25	Ala 385	Leu	Leu	Ala	Ala	Leu 390	Ala	Pro	Ala	Arg	Thr 395	Asp	Arg	Gly	Gly	Asp 400
30	Ser	Gly	Gly	Gly	Asp 405	Arg	Gly	Gly	Gly	Gl.y 410	Gly	Arg	Val	Ala	Leu 415	Thr
	Ala	Pro	Gly	Ala 420	Ala	Asp	Ala	Pro	Gly 425	Ala	Gly	Ala	Thr	Ala 430	Gly	Ile
35	His	Trp	Tyr 435	Ser	Gln	Leu	Leu	Tyr 440	Gln	Ile	Gly	Thr	Trp 445	Leu	Leu	Asp
	Ser	Glu 450	Ala	Leu <sub>.</sub>	His	Pro	Leu 455	Gly	Met	Ala	Val	Lys 460	Ser	Ser	Xaa	Ser

Arg Gly Ala Gly Gly Gly Ala Arg Glu Gly Ala
465 470 475

(2) INFORMATION FOR SEQ ID NO:16:

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- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 411 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear

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- (ii) MCLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:16:
- 15 Met Ser Pro Ala Arg Leu Arg Pro Arg Leu His Phe Cys Leu Val Leu
  1 5 . 10 . 15
  - Leu Leu Leu Val Val Pro Ala Ala Trp Cly Cys Gly Pro Gly Arg

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- Val Val Gly Ser Arg Arg Pro Pro Arg Lys Leu Val Pro Leu Ala 35 40 45
- Tyr Lys Gin Pne Ser Pro Asn Val Pro Glu Lys Thr Leu Gly Ala Ser

  50 55 60
  - Gly Arg Tyr Glu Gly Lys Ile Ala Arg Ser Ser Glu Arg Phe Lys Glu
    65 70 75 80
- 30 Leu Thr Pro Asn Tyr Asn Pro Asp Ile Ile Phe Lys Asp Glu Glu Asn 85 90 95
  - Thr Gly Ala Asp Arg Leu Met Thr Gln Arg Cys Lys Asp Arg Leu Asn 100 105 110

35

- Ser Leu Ala Ile Ser Val Met Asn Gln Trp Pro Gly Val Lys Leu Arg 115 120 125
- Val Thr Glu Gly Trp Asp Glu Asp Gly His His Ser Glu Glu Ser Leu

		130					135					140				
5	His 145	Tyr	Glu	Gly	Arg	Ala 150	Val	Asp	Ile	Thr	Thr 155	Ser	Asp	Arg	Asp	Arg 160
J	Asn	Lys	Tyr	Gly	Leu 165	Leu	Ala	Arg	Leu	Ala 170	Val	Glu	Ala	Gly	Phe 175	Asp
10	Trp	Val.	Tyr	Tyr 180	Glu	Ser	Lys	Ala	His 185	Val	His	Cys	Ser	Val 190	Lys	Ser
	Glu	His	Ser 195	Ala	Ala	Ala	Lys	Thr 200	Gly	Gly	Cys	Phe	Pro 205	Ala	Gly	Ala
15	Gln	Val 210	Arg	Leu	Glu	Ser	Gly 215	Ala	Arg	Val	Ala	Leu 220	Ser	Ala	Val	Arg
20	Pro 225	Gly	Asp	Arg	Val	Leu 230	Ala	Met	Gly	Glu	Asp 235	Gly	Ser	Pro	Thr	Phe 240
20	Ser	Asp	Val	Leu	Ile 245	Phe	Leu	Asp	Arg	Glu 250	Pro	His	Arg	Leu	Arg 255	Ala
25	Phe	Gln	Val	Ile 260	Glu	Thr	Gln	Asp	Pro 265	Pro	Arg	Arg	Leu	Ala 270	Leu	Thr
	Pro	Ala	His 275	Leu	Leu	Phe	Thr	Ala 280	Asp	Asn	His	Thr	Glu 285	Pro	Ala	Ala
30	Arg	Phe 290	Arg	Ala	Thr	Phe	Ala 295	Ser	His	Val	Gln	Pro 300	Gly	Gln	Туг	Val
35	Leu 305	Val	Ala	Gly	Val	Pro 310	Gly	Leu	Gln	Pro	Ala 315	Arg	Val	Ala	Ala	Val 320
<i>33</i>	Ser	Thr	His	Val	Ala 325	Leu	Gly	Ala	Tyr	Ala 330	Pro	Leu	Thr	Lys	His 335	Gly
	Thr	Leu	Val	Val	Glu	Asp	Val	Val	Ala	Ser	Cys	Phe	Ala	Ala	Val	Ala

Asp His His Leu Ala Gln Leu Ala Phe Trp Pro Leu Arg Leu Phe His Ser Leu Ala Trp Gly Ser Trp Thr Pro Gly Glu Gly Val His Trp Tyr Pro Gln Leu Leu Tyr Arg Leu Gly Arg Leu Leu Elu Glu Gly Ser Phe His Pro Leu Gly Met Ser Gly Ala Gly Ser (2) INFORMATION FOR SEQ ID NO:17: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 396 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:17: Met Ala Leu Leu Thr Asn Leu Leu Pro Leu Cys Cys Leu Ala Leu Leu Ala Leu Pro Ala Gln Ser Cys Gly Pro Gly Arg Gly Pro Val Gly Arg Arg Arg Tyr Ala Arg Lys Gln Leu Val Pro Leu Leu Tyr Lys Gln Phe Val Pro Gly Val Pro Glu Arg Thr Leu Gly Ala Ser Gly Pro Ala Glu Gly Arg Val Ala Arg Gly Ser Glu Arg Phe Arg Asp Leu Val Pro Asn

	Tyr	Asn	Pro	Asp	Ile 85		Phe	. Lys	Asp	G1 u 90		Asn	Ser	Gly	Ala 95	
5	Arg	Leu	Met	Thr 100		Arg	Cys	Lys	Glu 105	Arg	Val	Asn	Ala	Leu 110	Ala	Ile
10	Ala	Val	Met 115		Met	Trp	Pro	Gly 120	Val	Arg	Leu	Arg	Val 125	Thr	Glu	Gly
10	Trp	Asp 130		Asp	Gly	His	His 135	Ala	Gln	Asp	Ser	Leu 140	His	Tyr	Glu	Gly
15	Arg 145	Ala	Leu	Asp	Ile	Thr 150	Thr	Ser	Asp	Arg	Asp 155	Arg	Asn	Lys	Tyr	Gly 160
	Leu	Lev	Ala	Arg	Leu 165	Ala	Val	Glu	Ala	Gly 170	Phe	Asp	Trp	Val	Tyr 175	Tyr
20	Glu	Ser	Arg	Asn 180	His	Val	His	Val	Ser 185	Val	Lys	Ala	Asp	Asn 190	Ser	Leu
25	Ala	Val	Arg 195	Ala	Gly	Gly	Cys	Phe 200	Pro	Gly	Asn	Ala	Thr 205	Val	Arg	Leu
25	Trp	Ser 210	Gly	Glu	Arg	Lys	Gly 215	Leu	Arg	Glu	Leu	His 220	Arg	Gly	Asp	Trp
30	Val 225	Leu	Ala	Ala	Asp	Ala 230	Ser	Gly	Arg	Val	Val 235	Pro	Thr	Pro	Val	Leu 240
	Leu	Phe	Leu	Asp	Arg 245	Asp	Leu	Gln	Arg	Arg 250	Ala	Ser	Phe	Val	Ala 255	Val
35	Glu	Thr	Glu	Trp 260	Pro	Pro	Arg	Lys	Leu 265	Leu	Leu	Thr	Pro	Trp 270	His	Leu
	Val	Phe	Ala 275	Ala	Arg	Gly	Pro	Ala. 280	Pro	Ala	Pro	Gly	Asp 285	Phe	Ala	Pro

Val Phe Ala Arg Arg Leu Arg Ala Gly Asp Ser Val Leu Ala Pro Gly Gly Asp Ala Leu Arg Pro Ala Arg Val Ala Arg Val Ala Arg Glu Glu Ala Val Gly Val Phe Ala Pro Leu Thr Ala His Gly Thr Leu Leu Val Asn Asp Val Leu Ala Ser Cys Tyr Ala Val Leu Glu Ser His Gln Trp Ala His Arg Ala Phe Ala Pro Leu Arg Leu Leu His Ala Leu Gly Ala Leu Leu Pro Gly Gly Ala Val Gln Pro Thr Gly Met His Trp Tyr Ser Arg Leu Leu Tyr Arg Leu Ala Glu Glu Leu Leu Gly (2) INFORMATION FOR SEQ ID NO:18: (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 416 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:18: Met Asp Val Arg Leu His Leu Lys Gln Phe Ala Leu Leu Cys Phe Ile 

Ser Leu Leu Thr Pro Cys Gly Leu Ala Cys Gly Pro Gly Arg Gly

-				20	•		-		25					30	1	
5	Tyr	Gly	Lys 35	_	Arg	His	Pro	Lys 40	-	Leu	Thr	Pro	Leu 45		Туг	Lys
3	Gln	Phe 50		Pro	Asn	Val	Ala 55		Lys	Thr	Leu	61 y		Ser	Gly	Lys
10	Tyr 65		Gly	Lys	Ile	Thr 70	_	Asn	Ser	Glu	Arg 75		Lys	Glu	Leu	Ile 80
	Pro	Asn	Tyr	Asn	Pro 85	Asp	Ile	Ile	Phe	Lys 90		Glu	Glu	Asn	Thr 95	Asn
15	Ala	Asp	Arg	Leu 100		Thr	Lys	Arg	Cys 105	Lys	Asp	Lys	Leu	Asn 110	Ser	Leu
20	Ala	Πe	Ser 115		Met	Asn	His	Trp 120	Pro	Gly	Val	Lys	Leu 125	Arg	Val	Thr
20	Glu	Gly 130	Trp	Asp	Glu	Asp	Gly 135	His	His	Leu	Glu	Glu 140	Ser	Leu	His	Tyr
25	Glu 145	Gly	Arg	Ala		Asp 150	Ile	Thr	Thr	Ser	Asp 155	Arg	Asp	Lys	Ser	Lys 160
	Tyr	Gly	Met	Leu	Ser 165	Arg	Leu	Ala	Val	Glu 170	Ala	Gly	Phe	Asp	Trp 175	Val
30	Tyr	Tyr	Glu	Ser 180	Lys	Ala	His	Ile	His 185	Cys	Ser	Val	Lys	Ala 190	Glu	Asn
35	Ser	Val	Ala 195	Ala	Lys	Ser	Gly	Gly 200	Cys	Phe	Pro	Gly	Ser 205	Gly	Thr	Val
	Thr	Leu 210	Gly	Asp	Gly	Thr	Arg 215	Lys	Pro	Ile	Lys	Asp 220	Leu	Lys	Val	Gly
	Asp	Ara	Val	Leu	Ala	Ala	Asp	Glu	Lys	Gly	Asn	Val	Leu	Ile	Ser	Asp

	225					230					235					240
5	Phe	Ile	Met	Phe	Ile 245	Asp	His	Asp	Pro	Thr 250	Thr	Arg	Arg	Gln	Phe 255	Ile
,	Val	Ile	Glu	Thr 260	Ser	Glu	Pro	Phe	Thr 265	Lys	Leu	Thr	Leu	Thr 270	Ala	Ala
10	His	Leu	Val 275	Phe	Val	Gly	Asn	Ser 280	Ser	.Ala	Ala	Ser	Gly 285	Ile	Thr	Ala
	Thr	Phe 290	Ala	Ser	Asn	Val	Lys 295	Pro	Gly	Asp	Thr	Val 300	Leu	Val	Trp	Glu
15	Asp 305	Thr	Cys	Glu	Ser	Leu 310	Lys	Ser	Val	Thr	Val 315	Lys	Arg	Ile	Tyr	Thr 320
20	Glu	Glu	His	Glu	Gly 325	Ser	Phe	Alā	Pro	Val 330	Thr	Ala	His	Gly	Thr 335	lle
20	Ile	Val	Asp	Gln 340	Val	Leu	Ala	Ser	Cys 345	Tyr	Ala	Val	Ile	G].u 350	Asn	His
25	Lys	Trp	Ala 355	His	Trp	Ala	Phe	Ala 360	Pro	Val.	Arg	Leu	Cys 365	His	Lys	Leu
	Met	Thr 370	Trp	Leu	Phe	Pro	Ala 375	Arg	Gl.u	Ser	Asn	Val 380	Asn	Phe	Gln	Glu
30	Asp 385	Gly	Ile	His	Trp	Tyr 390	Ser	Asn	Met	Leu	Phe 395	His	Ile	Gly	Ser	Trp 400
35	Leu	Leu	Asp	Arg	Asp 405	Ser	Phe	His	Pro	Leu 410	Gly	Ile	Leu	His	Leu 415	Ser

(2) INFORMATION FOR SEQ ID NO:19:

(i) SEQUENCE CHARACTERISTICS:

			- (2	וע (א	FINGI	n. I	a TO 1	Jase	Par.	LS							
	<ul><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: both</li><li>(D) TOPOLOGY: linear</li></ul>																
			((	C) Si	rani	DEDNI	ESS:	botl	h								
			(1	D) T	OPOL	OGY:	lin	ear									
5																	-
		(ii)	) MO	LECU!	LE T	YPE:	cDN.	Ą									
										٠							
		(ix)	) FE	ATURI	Ξ:												
10			(2	A) N	AME/I	KEY:	CDS										
			(1	B) L	OCAT:	ION:	1	1413									
		(xi)	SE	QUEN	CE DI	ESCR:	IPTI	ON: S	SEQ :	ID NO	0:19	:					
15																	
				CAC													48
	Met	Asp	Asn	His		Ser	Val	Pro	Trp		Ser	Ala	Ala	Ser		Thr	
	1				5					10					1.5		
									~		m.m.o.		(Om ⊃	ca c	mme:	COR CO	96
20				CTG													90
	Cys	Leu	Ser	Leu	Gly	Cys	GIn	Met		Gin	Fne	GIN	Pne	30	Pne	GIII	•
				20					25					.50			
	0.00	~ n n	D (1)	CGC	7.00	CAC	CTC	CAT	CTC	ccc	ሌ አ C	CCC	CCA	מממ	ACA	ACG	144
25				Arg													
25	Leu	GIII	35	Arg	Ser	GIU	пец	40	пеа	my	шys	110	45	9	• 9		
			دد					30									
	۵۵	ACG	ATG	CGC	CAC	АТТ	GCG	CAT	ACG	CAG	CGT	TGC	CTC	AGC	AGG	CTG	192
				Arg													
30	017.	50	.,,,,	9			55					60					
50																	
	ACC	TCT	CTG	GTG	GCC	CTG	CTG	CTG	ATC	GTC	TTG	CCG	ATG	GTC	TTT	AGC	240
				Val													
	65					70					75					80	
35																	
	CCG	GCT	CAC	AGC	TGC	GGT	CCT	GGC	CGA	GGA	TTG	GGT	CGT	CAT	AGG	GCG	288
				Ser													
					85					90					95		

	CGC	AAC	CTG	TAT	CCG	CTG	GTC	CTC	AAG	CAG	ACA	ATT	CCC	AAT	CTA	.TCC	336
	Arg	Asn	Leu	Tyr	Pro	Leu	Val	Leu	Lys	Gln	Thr	Ile	Pro	Asn	Leu	Ser	
				100					105					110			
5	GAG	TAC	ACG	AAC	AGC	GCC	TCC	GGA	CCT	CTG	GAG	GGT	GTG	ATC	CGT	CGG	384
	Glu	Tyr	Thr	Asn	Ser	Ala	Ser	Gly	Pro	Leu	Glu	Gly	Val	Ile	Arg	Arg	
			115					120					125				
	GAT	TCG	CCC	AAA	TTC	AAG	GAC	CTC	GTG	CCC	AAC	TAC	AAC	AGG	GAC	ATC	432
10	Asp	Ser	Pro	Lys	Phe	Lys	Asp	Leu	Val	Pro	Asn	Tyr	Asn	Arg	Asp	Ile	
		130					135					140					
		TTC															480
		Phe	Arg	Asp	Glu		Gly	Thr	Gly	Ala	-	Gly	Leu	Met	Ser	-	
15	145					150					155					160	
											<b></b>		a.a.a				500
		TGC															528
	Arg	Cys	rys	GLu	_	ren	Asn	vai	Leu		Tyr	ser	vai	Met		GIU	
20					165					170					175		
20	mcc.	020	666	N CTC	ccc	Cmc	cmc	CTC	7.00	~n~	N.C.C	TICC.	CAC	cac	CAC	<b>ም</b> ል. ን	E7 <i>6</i>
		CCC															576
	пр	Pro	GIĀ	180	Arg	ьеи	Leu	Val	185	GIU	Ser	irb	изр	190	изр	ığı	
				100					103					190			
25	ጥፈጋ	CAC	ദേദ	CAG	GAG	TCG	CTC	CAC	TAC	GAG	GGC	CGA	GCG	GTG	ACC	АТТ .	624
		His															
•			195					200	-1-		1	9	205				
	GCC	ACC	TCC	GAT	CGC	GAC	CAG	TCC	AAA	TAC	GGC	ATG	CTC	GCT'	CGC	CTG	672
30		Thr															
		210		-	-		215		-	•	-	220			_		
	GCC	GTC	GAG	GCT	GGA	TTC	GAT	TGG	GTC	TCC	TAC	GTC	AGC	AGG	CGC	CAC	720
		Val															
35	225				-	230	-	-			235					240	
	ATC	TAC	TGC	TCC	GTC	AAG	TCA	GAT	TCG	TCG	ATC	AGT	TCC	CAC	GTG	CAC	768
		Tyr															
		<b>.</b> -	•		245	-		٠		250					255		

							•	• •										
	GGC	TGC	TTC	ACG	CCG	GAG	AGC	ACA	GCG	CTG	CTG	GAG	AGT	GGA	GTC	CGG	8	116
	Gly	Cys	Phe	Thr	Pro	Glu	Ser	Thr	Ala	Leu	Leu	Glu	Ser	Gly	Val	Arg		
				260		•			265					270				
5																		
	AAG	CCG	CTC	GGC	GAG	CTC	TCT	ATC	GGA	GAT	CGT	GTT	TTG	AGC	ATG	ACC	8	64
	Lys	Pro	Leu	Gly	Glu	Leu	Ser	Ile	Gly	Asp	Arg	Val	Leu	Ser	Met	Thr		
			275					280					285					
10	GCC	AAC	GGA	CAG	GCC	GTC	TAC	AGC	GAA	GTG	ATC	CTC	TTC	ATG	GAC	CGC	9	12
	Ala	Asn	Gly	Gln	Ala	Val	Tyr	Ser	Glu	Val	Ile	Leu	Phe	Met	Asp	Arg		
		290					295					300						
	AAC	CTC	GAG	CAG	ATG	CAA	AAC	TTT	GTG	CAG	CTG	CAC	ACG	GAC	GGT	GGA	. 9	60
15	Asn	Leu	Glu	Gln	Met	Gln	Asn	Phe	Val	Gln	Leu	His	Thr	Asp	Gly	Gly		
	305					310					315					320		
															CÄG		1.0	800
	Ala	Val	Leu	Thr	Val	Thr	Pro	Ala	His	Leu	Val	Ser	Val·	Trp	Gln	Pro		
20					325					330					335			
															GAG		10	)56
	Glu	Ser	Gln	Lys	Leu	Thr	Phe	Val		Ala	His	Arg	Ile		Glu	Lys		
				340					345					350				
25												020	ame.	7.00	ccc.	CAC	11	0.4
															CCC		11	.04
	Asn	Gln		Leu	Val	Arg	Asp		Glu	Thr	GTÀ	GIU		Arg	Pro	GIII		
			355					360					365					
••		ar -	ama	220	mm^	ccc	a cm	CTC	ccc	λCm	ስ አ <i>ር</i>	CCC	GTC	GTC	GCG	CCG	11	.52
30																		
	Arg		Val	гàг	Leu	GIY		vaı	Arg	ser	гÃ2		Vai	vai	Ala	110		
		370					375					380						
				~~~	000	7.00	z mm	CTC.	CTC	ስ ክ C	TICC.	CTC	ccc	CCC	AGT	ፕርር	. 12	200
25															Ser			. • •
35		Thr	Arg	GIU	сту		116	Val	Val	ASII		Vai	ALG	nια	001	400		
	385					390					395					100		
	m	000	O.T.C	7 m c	7 7 C	አርጥ	CNC	TICC.	CTC	CCC	ርእር	ሞርር	GGA	ርሞር	GCT	CCC	12	48
																	12	
	Tyr	Ala	val	TTE	Asn	ser	GID	ser	ьеи	мта	uis	ттр	GTÀ	neu	Ala			

410 415 405 ATG CGC CTG CTG TCC ACG CTG GAG GCG TGG CTG CCC GCC AAG GAG CAG 1296 Met Arg Leu Leu Ser Thr Leu Glu Ala Trp Leu Pro Ala Lys Glu Gln 420 425 TTG CAC AGT TCG CCG AAG GTG GTG AGC TCG GCG CAG CAG CAG AAT GGC Leu His Ser Ser Pro Lys Val Val Ser Ser Ala Gln Gln Gln Asn Gly 445 440 435 10 ATC CAT TGG TAT GCC AAT GCG CTC TAC AAG GTC AAG GAC TAC GTG CTG 1392 Ile His Trp Tyr Ala Asn Ala Leu Tyr Lys Val Lys Asp Tyr Val Leu 460 450 455 1416 15 CCG CAG AGC TGG CGC CAC GAT TGA Pro Gln Ser Trp Arg His Asp 465 470 (2) INFORMATION FOR SEQ ID NO:20: 20 (i) SEQUENCE CHARACTERISTICS: (A) LENGTH: 471 amino acids (B) TYPE: amino acid (D) TOPOLOGY: linear 25 (ii) MOLECULE TYPE: protein (xi) SEQUENCE DESCRIPTION: SEQ ID NO:20: 30 Met Asp Asn His Ser Ser Val Pro Trp Ala Ser Ala Ala Ser Val Thr 5 10 1 Cys Leu Ser Leu Gly Cys Gln Met Pro Gln Phe Gln Phe Gln Phe Gln 30 20 35 Leu Gln Ile Arg Ser Glu Leu His Leu Arg Lys Pro Ala Arg Arg Thr 45 35

	Gln	Thr 50	Met	Arg	His	Ile	Ala 55	His	Thr	Gln	Arg	Cys 60	Leu	Ser	Arg	Leu
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105

110

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5	As	p Ar 13	g Asp O	Xaa	Ser	Lys	Туг 135	Gly	Xaa	Leu	Xaa	Arg 140	Leu	Ala	Val	Glu
	Al 14		y Phe	Asp	Trp	Val 150	Tyr	Tyr	Glu	Ser	Lys 155	Ala	His	Ile	His	Cys 160
10	Se	r Va	l Lys	Ala	Glu 165	Asn	Ser	Val	Ala	Ala 170	Lys	Ser	Gly	Gly	Cys 175	Phe
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30			LECUL													
	(v	) FR	AGMEN'	r TY	PE: i	.nter	nal									
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